



CORK UNIVERSITY HOSPITAL LABORATORY MEDICINE USER HANDBOOK

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| Approver(s): | Dr Vitaliy Mykytiv, Ms Sinead Creagh | Approval Date: | 16/10/2023 11/10/2023 |

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| | Approved By: | Dr Vitaliy Mykytiv, Ms | Sinead Creagh |
| | Author: | Mr Paul Cantwell | |

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1 AMENDMENT TABLE

The Laboratory Medicine User Handbook is controlled in accordance with local quality management system requirements. Amendments to the last two revisons are listed in the table below. The full amendment history is available by contacting the Laboratory Medicine Quality Manager (refer to section 4.3: Contact Details).

| nended Section(s) | Amendment |
|-------------------------------------|---|
| Section 3.3 Contact Details | Updated for Department of Pathology |
| Section 4.6.3 Types of clinical | POCT Creat and POCT HbA1c added to section 4.6 |
| services offered by the lab | Details re spare POCT COBAS Liat held in Covid lab removed |
| services offered by the lab | from section 4.6 |
| Section 4.4.2 Types of clinical | Haematology: Added anticardiolipin and b2 glycoprotein 1 to |
| services offered by the lab | lupus anticoagulant screen |
| services offered by the lab | |
| Section 4.4.2 Types of clinical | Haematology: Added Kleihauer Testing to the list |
| services offered by the lab | |
| Section 7.4 Additional requirements | Remove the additional requirements for sending on ice |
| Section 8.2 Critical Results | Updated the phoning requirements for the following in |
| Reporting | Biochemistry: Triglcyerides, Calcium (Paeds), Ethanol, |
| | Bilirunin (conjugated) and Direct Sodium. Urate was |
| | removed from the list. |
| Section 8.2 Critical Results | Include requirement for appropriate contact details for |
| Reporting | critical results |
| | POCT Blood gas critical results added to list |
| | POCT Creatinine critical results added to list |
| Section 9.3 "Instructions for using | Remove section 9.3 - "Instructions for using Lab Enquiry / |
| Lab Enquiry / Netterm" | Netterm" as no longer applicable. Blood component collection |
| Eus Enquiry / Netterni | slips now generated through the EBTS system |
| Section 12 Test Directory | Ammonia- updated the comment to the following: Fill |
| Section 12 Test Directory | |
| | specimen to the top and transport to the laboratory |
| | immediately as sample needs to be spun within 30 minutes |
| | of collection |
| | Alcohol (Ethanol)- added the following comment: |
| | Alcohol measurement is provided for clinical purposes only. |
| | Samples will not be accepted for medicolegal or workplace |
| | testing |
| | Anti cardiolipin: changed sample volume to 3.5 ml serum |
| | sample |
| | Bence Jones Proteins; updated the comment to the |
| | following: |
| | As of June 6th requests for BJP are limited to Haematology Consultant request only |
| | Beta 2 glycoprotein : changed sample volume to 3.5 ml |
| | serum sample and changed TAT to 4- 6 weeks |
| | Activated Partial Thromboplastin Time (APTT), reference |
| | |
| | range for 90 day infant corrected to 24-50 seconds. |
| | Biochemistry: TAT for the following referral tests from |
| | Biochemsitry were updated: Zinc |
| | Vitamin E, Vitamin A, Copper, Vitamin K, B 2 Microglobulin, |
| | AAC, CSF Oligoclonal Bands, Urinary Organic Acids, Urinary |
| | Catecholamines, TPMT, 17 OH Progesterone, |
| | 3 Hydroxybutyrate, AAT Phenotype, CSF Pyridoxal |
| | Phosphate, Gastrin, IGD, Chromium, Insulin-like-growth |
| | factor BP3, Tetanus antibodies (IgG) |
| | Purine/pyrimidine Screen, Amyloid A, DHEAS, Flecanide, |
| | Urinary Steroid Profile Metanephrines, |
| | Karyotype/Chromosome analysis (>5y old), |
| | Karyotype/Chromosome analysis (<5y old), |
| | Haemophilus influenzae B Antibodies (IgG), Mitotane |

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| | Aution | Pill Taul Calitwell | | | | |
| | Biochemistry: foll | owing referral tests a | dded to Test Directory | | | |
| | | of specimen require | | | | |
| | laboratory and tu | | | | | |
| | | e, CSF Pyridoxal Pho | sphate. | | | |
| | | ne, Cystine (WBC), M | | | | |
| | | | nidinoacetate/creatine, | | | |
| | Free Fatty Acids, Mycophenolate and Urinary Steroid Profile, | | | | | |
| | | | ests, Dexamethasone | | | |
| | | and Dynamic functio | | | | |
| | | culated Globulin add | | | | |
| | | id (CSF) – Culture an | | | | |
| | | | d where requested by | | | |
| | | g green Microbiology | | | | |
| | | | e Toxin testing carried | | | |
| | | RGET DETECTED sam | | | | |
| | Creatinine (POCT |) added toTest Direct | ory | | | |
| | | stopathology Specim | ens | | | |
| | GIT biopsies | | | | | |
| | Upper GIT P | | | | | |
| | Upper GT P | | | | | |
| | Upper GIT P | | | | | |
| | | c PO4 TAT = 10 wor | | | | |
| | Lower GIT P0 | | | | | |
| | Lower GIT P0 | | | | | |
| | Gynae P01 Gynae P04 | TAT = 12 wor | | | | |
| | Gynae P04Skin P01 | TAT = 12 work | | | | |
| | | $\frac{\text{TAT} = 14 \text{ work}}{\text{ded the following refe}}$ | | | | |
| | | el for Inherited | | | | |
| | | | ford, Churchill Hospital) | | | |
| | | | of AML, CML and ALL | | | |
| | (Munich leukaemi | | ·····, ···· | | | |
| | • | | H, for the diagnosis of | | | |
| | | | eloma, Plasmocytoma- | | | |
| | (Munich leukaemi | | | | | |
| | Cytogenetics (Chi | omosome banding) f | or the diagnosis of AML, | | | |
| | CML, ALL and MD | S-(Munich leukaemia | lab) | | | |
| | Haematology: Up | dated GATA Mutation | al analysis referral lab | | | |
| | | Bristol NHS Trust, | | | | |
| | | ab, Pathology Science | | | | |
| | · · · · | ry-On-Trym, Bristol, I | | | | |
| | | dded toTest Director | | | | |
| | | | d the comment to the | | | |
| | _ | | ch must be separated | | | |
| | within 30 minutes | | | | | |
| | | creased TAT to 7 wor | | | | |
| | - | | - updated comment | | | |
| | | and sample acceptan | се | | | |
| | Renal Biopsy- add | | | | | |
| | | | re processed in-house | | | |
| | | s are then referred to | | | | |
| | | weeks for renal tran | splant case referred to | | | |
| | Beaumont. | Chaine II II II | Discolor | | | |
| | | Chains added to test | | | | |
| | | | llowing comment: Drug | | | |
| | | | clinical purposes only. | | | |
| | | be accepted for me | dicolegal or workplace | | | |
| | testing | | · - | | | |
| | | | comment: Drug screen | | | |
| | | | urposes only. Samples | | | |
| | will not be accept | ed for medicolegal or | workplace testing | | | |

| Approved By: Dr Vitaliy Myktyiu, Ms Sinead Creagh Author: Mr Paul Cantwell Urine microscopy has resulted in a change of container requirements. Vitamin B12 assay-Biological reference range changing from 120 - 650 ng/L to 140 - 844ng/L , indeterminate reference range changing from 100 - 150ng/L to 120 - 170 ng/L. Haematology: FMH by flow cytometry to include the changes that all postnatal samples with bleeds ≥4mls are referred. Also updated Kielnauer Test for Foetal Cells FMH-in line with bleeds ≥4mls are NOT referred. Kleihauer on a rhesus D Neg mother of a baby with a weak D Ag are NOT referred. Also updated Kleihauer Test for Foetal Cells FMH-in line with bleeds ≥4mls are NOT referred. Kleihauer on a rhesus D Neg mother of a baby with a weak D Ag are NOT referred. Also updated Kleihauer Test for Foetal Cells FMH-in line with bleeds ≥4mls are NOT referred. Haematology: TAT's for the following referred tests wer updated: Cerebrospinal Fluid (CSF) - Immunophenotyping primary CNS lymphoma or CNS involvement by Leukaemia, lymphoma-40 days GLL Prognostic Markers (TP53 and IGVH mutation status)-67 days; Fibrinogen Phenotyping and Genetic Analysis-80 days; GEPD Assay-50 days; Hearophilia MH Research-120 days Heparin ./PF4 Antibody Test (HIT; Heparin Induce Thrombocytopenia screening test)-36 days; HLH Granul release assay (Haemophagocytic Lympho Histocytosis)-21 days; fibrinogen Mutimers / Collager binding - 90 days; 140 days Warfarin Plasma Resistance Concentration and gene - 2: days; Md days Molecular Pathology: Next Generation Sequencing cTNA Plasma Molecular testing in the pathology laboratory in Berformed on request from | Title: Laboratory Medicine User Handbook | Reference: | PPG-CUH-PAT-31 | Revision: 22 | | | |
|---|--|-----------------------|-------------------------|-------------------------|--|--|--|
| Author: Mr Paul Cantwell Urine microscopy (Microbiology): Introduction of automated urine microscopy has resulted in a change of container requirements. Vitamin B12 assay-Biological reference range changing from 120 - 650 ng/L to 140 - 844ng/L, indeterminate reference range changing from 100 - 150ng/L to 120 - 170 ng/L. Haematology: FMH by flow cytometry to include the changes that all postnatal samples with bleeds ≥4mls are referred to the Rotunda for flow cytometry. Antenatal patients with bleeds ≥4mls are referred to the Rotunda for flow cytometry. Antenatal patients with bleeds ≥4mls are referred to the Rotunda for flow cytometry. Antenatal patients with above changes Haematology: TAT's for the following referred tests were updated: Cerebrospinal Fluid (CSF) - Immuophenotyping primary CNS lymphoma or CNS involvement by Leukaemia, lymphoma-40 days CLL Prognostic Markers (TP53 and IGVH mutation status)-65 days; Flamophilia MH Research-120 days GPD Assay-60 days; Haemophilia MH Research-120 days; Heparin /PF4 Antibody Test (HIT; Heparin Inducer Thrombocytopenia screening test)-36 days; Flut Granul release assay (Haemophila MT Research-120 days; Wutation analysis for inherited bleeding disorders Haemophilia carrier testing for direct mutational detection mutation analysis for inherited Teator VIII or Factor 12 days; Wutation analysis for inherited bleeding on gene; PMN Paroxysmal nocturnal haemoglobinuria -60 days; Uon Willebrand Multimers / Collagen binding -90 days / 140 days; Warfarin Plasma Resistance Concentration and gene: 2 days/80 days Molecular Pathology: Next Generation Sequencing cfTNA Plasma Molecular testing in the pathology laboratory UH Plasox of as a molecular testing from Consultar Histopathologists on plasma samples from pat | | Active Date: | 03/11/2023 | Page: 6 of 206 | | | |
| Urine microscopy (Microbiology): Introduction of automated urine microscopy has resulted in a change of container requirements. Vitamin B12 assay-Biological reference range changing from 120 - 650 ng/L to 140 - 844ng/L , indeterminate reference range changing from 100 - 150ng/L to 120 - 170 ng/L. Haematology: FMH by flow cytometry to include the changes that all postnatal samples with bleeds 24mls are referred to the Rotunda for flow cytometry. Antenatal patients with bleeds 24mls are NOT referred. Kleihauer on a rhesus D Neg mother of a baby with a weak D Ag are NOT referred. Also updated kleihauer Test for Foetal Cells FMH-in line with above changes Haematology: TAT's for the following referred tests were updated: Cerebrospinal Fluid (CSF) - Immunophenotyping primary CNS lymphoma or CNS involvement by Leukaemia lymphoma-40 days CLL Prognostic Markers (TP53 and IGVH mutation status)-6: days; Fibrinogen Phenotyping and Genetic Analysis-80 days G6PD Assay-60 days; Haemophilia MH Research-120 days Heparin /PF4 Antibody Test (HIT; Heparin Induced Thrombocytopenia screening test)-36 days; HLH Granul release assay (Haemophagocytic Lympho Histocytos)-21 days; Mutation analysis for inherited bleeding disorders Haemophilic carrier testing for inherited bleeding disorders Haemophilic arrier testing for inherited on days; VON Willebrand Multimers / Collagen binding -90 days; 140 days Warfarin Plasma Resistance Concentration ad gene - 2: days/80 days Molecular Pathology: Next Generation Sequencing dTNA Plasma Molecular testing in the pathology laboratory CH is performed on request from Consultant Histopathologists on plasma samples from patients with Lung cancer. The cut-off for receipt of these samples into the laboratory is 15:00 2 K2 EDTA Blood tubes (must reach lab within 4 hours) OR at least 1 Roche drDNA blood tube Please contact the laboratory prior t | | | | s Sinead Creagh | | | |
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| requirements. Vitamin B12 assay-Biological reference range changing from 120 - 650 ng/L to 140 - 844ng/L, indeterminate reference range changing from 100 - 150ng/L to 120 - 170 ng/L. Haematology: FMH by flow cytometry to include the changes that all postnatal samples with bleeds 2-4mls are ROT referred. Kleihauer on a rhesus D Neg mother of a baby with a weak D Ag are NOT referred. Also updated Kleihauer Test for Foetal Cells FMH-in line with above changes Haematology: FMH back and the composition of the comparison of the | | | | | | | |
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| ref lab supplier | | | | | | | |
| | Section 15 | | emoved ThromboGen | omics Cambridge as a | | | |
| Added MLL and Oxford genetic lab as new suppliers. | | | wford conctining the | now cuppliant | | | |

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| | Author: | Mr Paul Cantwell | |

2 INTRODUCTION

2.1 Overview

The profile of laboratory services offered has changed dramatically in recent years and continues to evolve as new technologies and methodologies are discovered. It is our hope that this User Handbook will familiarise the user with departmental policies as well as specific test requirements.

Laboratory policy statements include brief descriptions of each laboratory, location for specimen delivery, key contact personnel, the hours of operation and instructions concerning specimen collection and transportation to the laboratory. Specific criteria for refusal of requests for examination of specimens should be noted. Regretably service may not be provided if acceptance criteria are not fulfilled. Other special instructions are also included as well as details of the out-of-hours (on-call) service.

In order to obtain the best possible laboratory services, it is essential to ensure that all specimens are collected properly, and that both the specimen and request form are labelled with the appropriate information.

All tests are listed alphabetically in the "Laboratory Medicine Test Directory" with complete ordering information including the name of the test, department that will process the specimen, specimen and container required, reference intervals (where appropriate), special comments and turnaround times.

The information in this handbook is subject to change and will be updated to keep the information current.

2.2 Disclaimer

This handbook has been prepared by laboratory staff at Cork University Hospital and every care has been taken in its compilation. This handbook is intended to be used as a guide only. Practitioners should use this handbook as a guide to individual testing on the basis of clinical findings, not as a complete or authoritative statement of such testing.

Laboratory Medicine shall not be liable to users of the handbook nor to any other person, firm, company or other body for any loss, direct, indirect, or consequential, in contract or in tort or for any negligent mis-statement or omission contained herein, by reason of, arising from or in relation to any such user, other person, company or body relying or acting upon or purporting to rely or act upon any matter contained in this handbook.

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| | Author: | Mr Paul Cantwell | |

2.3 Major Objectives

Laboratory Medicine is committed to providing the highest quality diagnostic and consultative services for all its users.

Major Objectives

- 1. To provide examinations that are fit for their intended use;
- 2. To provide all employees with the knowledge, training, and tools necessary to allow for the completion of accurate and timely work;
- 3. To provide an effective service to its users;
- 4. To uphold professional values and conduct;
- 5. To provide safe and suitable conditions for all staff and visitors to the laboratory;
- 6. To procure and maintain equipment and other resources needed for the provision of the service;
- 7. To ensure that all personnel are familiar with the contents of the Quality Manual and all procedures relevant to their work;
- 8. To collect, transport and handle of all specimens in such a way as to ensure the correct performance of laboratory examinations;
- 9. To report results of examinations in ways which are timely, confidential, accurate and clinically useful;
- 10. To operate a quality management system to integrate the organisation, procedures, processes and resources.

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| | Author: | Mr Paul Cantwell | |

3 GENERAL INFORMATION

3.1 The location of the laboratory

Laboratory Medicine at Cork University Hospital is situated on the ground floor of the main Cork University Hospital building and can be accessed via the ground floor of the main hospital building.

The postal address of the CUH laboratory service is: Laboratory Medicine Cork University Hospital Wilton Cork City Ireland T12 DC4A

There are six Departments within CUH Laboratory Medicine whose main activities are described below.

| | Department /Section | Location |
|----|--|---|
| 1. | Blood Transfusion | Ground floor, Laboratory building |
| 2. | Clinical Biochemistry | Ground floor, Laboratory building. |
| | Molecular Genetics | Ground floor on the link corridor between outpatients and laboratory reception |
| 3. | Clinical Microbiology | First floor, Laboratory building |
| | Infectious Diseases Serology | Located on the ground floor, opposite Physiotherapy department. |
| | Covid Laboratory | Stand alone purpose built laboratory beyond the Goods inwards entrance for stores |
| 4. | Haematology and Coagulation | Ground floor, Laboratory building |
| | Haematinics | Ground floor, by outpatients |
| | Molecular Genetics | Ground floor on the link corridor between outpatients and laboratory reception |
| 5. | Pathology | |
| | Histopathology Cytopathology Molecular Pathology | First Floor, Laboratory building (Swipe access only)* |
| | Electron Microscopy /Renal Next Generation Sequencing | Ground Floor, CUH (Adjacent to Theatre 9) |
| | Post Mortem | Ground Floor, Laboratory building adjacent to Biochemistry |
| | Neuropathology | Ground floor on the link corridor between outpatients and laboratory reception |
| 6. | Autoimmune Serology | Autoimmune Serology shares the ground floor of the Laboratory building with the Haematology and Biochemistry Departments. |

*It is advisable that external couriers have contact numbers for laboratories, as laboratories are swipe access only.

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3.2 Opening Hours and Laboratory Telephone Extension Numbers

Prefix (021) 49 for direct access from outside Cork University Hospital. The telephone enquiry service should be used for emergency enquiries only.

Sample Deadline denotes the cut-off for receipt of routine samples. A detailed list of on-call tests is outlined in the section "On-Call Tests".

| Blood Transfusion | Contact No | Opening Hours | Sample Deadline | |
|---|----------------------|---|------------------|--|
| Blood Transfusion Laboratory | Ext. 22537 | 08 :00-20 :00 | 17 :00 (Mon-Fri) | |
| | | Mon-Fri | 09 :30 (Sat) | |
| | | 09 :00-12 :00 Sat | | |
| Antenatal Section of Laboratory | Ext: 22668 | | | |
| Blood Transfusion Laboratory Fax Number: | (021) 4922004 | Only emergency samp during the out-of-hour | | |
| Medical Scientist On-call | Bleep:199 | - | | |
| | | appropriately and proc working day. | | |
| Clinical Biochemistry | Contact No | Opening Hours | Sample Deadline | |
| Clinical Biochemistry | Ext. 20173 | 08:00-20.00 Mon-Fri | 16:30 Mon-Fri | |
| Specific Proteins / Immunology | Ext. 22535 | Only emergency samp | | |
| Molecular Genetics | Ext. 22361 /22531 | | | |
| | | | | |
| Medical Scientist on call | Bleep: 376 | | | |
| | | | | |
| Clinical Microbiology | Contact No | Opening Hours | Sample Deadline | |
| Clerical Office –Results/Enquiries | Ext. 22501 | 09:00-17:00 Mon-Fri | 16:30 Mon-Fri | |
| Main Laboratory | Evt 22503 | Limited service after 17:00 | | |

| Clerical Office –Results/Enquiries | Ext. 22501 | 09:00-17:00 Mon-Fri | 16:30 Mon-Fri | | |
|------------------------------------|-----------------------|---|-------------------------|--|--|
| Main Laboratory | Ext. 22503 | Limited service after 17:00 | | | |
| Routine Bacteriology, Mycology | /22505 | Only emergency samples will be process | | | |
| and Antibiotic Assays | | during the out-of-hours service. A detailed | | | |
| Infectious Diseases Serology | Ext. 22506 | | outlined in the section | | |
| Category 3 Laboratory – TB | Ext. 22823 | | urgent specimens will | | |
| Category 3 Laboratory – Enterics | Ext. 22821 | | ely and processed the | | |
| Infection Control | Ext. 28074 / 28075 | next working day. | | | |
| Covid Laboratory | Ext. 22139 | | | | |
| Medical Scientist on call: | Bleep: 375 | | | | |
| Haematology and Coagulation | Contact No | Opening Hours | Sample Deadline | | |
| Clerical Office –Results/Enquiries | Ext. 22541 | Routine hours are | 16:30 Mon-Fri | | |
| | | defined as 09:00 to | 12 :00 Sat | | |
| | | 17:00, except for | | | |
| | | the following tests | | | |
| | | FBC and routine | | | |
| | | Coagulation which | | | |
| | | are analysed | | | |
| | | between 08:00 to | | | |
| | | 20:00 | | | |

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| | | 1 1 | | |
|---|----------------------|---|---------------------|--|
| | | Mon-Fri, and | | |
| | | 09:00 to 12:00 Sat | | |
| Main Laboratory | Ext. 20172 | Only emergency samples will be process | | |
| Haematinics | Ext. 22128 | during the out-of-hou | | |
| Specimen reception | Ext. 22547 | list of on-call tests is o | | |
| Flow Cytometry Laboratory | Ext. 21351 | "On-Call Tests". Non u be stored and process | | |
| | | - | ed the next working | |
| Medical Scientist on call (Haematology): | Bleep: 377 | day. Only emergency samples will be processed during the out-of-hours service. A detailed list of on-call tests is outlined in the section "On-Call Tests". Non urgent specimens will be stored and processed the next working | | |
| | | day. | | |
| Pathology | Contact No | Opening Hours | Sample Deadline | |
| Histopathology (Laboratory) | Ext:22792 | 08 :00-18 :00 Mon- | 16:30 Mon-Fri | |
| | | Fri | Fixed & unfixed | |
| Secretariat | Ext:22514 | 09 :00 12 :00 Sat | specimens | |
| | / 22510 | 08 :00-18 :00 Mon- | 11:45 Sat. | |
| Breast Secretariat | Ext: 20497 | Fri | | |
| | | 08 :00-18 :00 Mon- Fri | | |
| Cytopathology | Ext. 22511 | 9am 5pm Mon Fri | 4.30pm | |
| , | | No service on Sat | | |
| Specimen Reception | Ext. 22792 | | | |
| Consultant Pathologist/clerical | Ext.22514/ | | | |
| office | 22510/ | | | |
| | 20497 | | | |
| Post Mortem /Mortuary Services | Ext. 22525 /22883 | 24 hour service | 11am cut-off | |
| Perinatal Pathology Team | 087 3691513 | 8-4pm Mon-Fri (exl. bank holidays) | Contact PNP team | |
| Renal Pathology/Electron | Ext 21315 | 08:00-16:00 Mon-Fri | Mon – Fri 8am to | |
| Microscopy | | | 15:30pm | |
| | | | | |
| Out of hours contact Pathologist on | | | | |
| Neuropathology Office | Ext 22520 | 09:00-17:00 Mon-Fri | 16:00 Mon-Fri | |
| Neuropathology Laboratory | Ext 22519 | | | |
| Mobile for Consultant Neuropathol | ogist on call: (| Contact CUH switchboa | ard | |
| Immunology | Contact No | Opening Hours | Sample Deadline | |
| Autoimmune Serology | Ext. 22535 | 08:00-17:00 Mon-Fri | 16:30 Mon-Fri | |
| - • | | No service on Sat | | |
| Laboratory Medicine Information Systems | Contact No | Opening Hours | Sample Deadline | |
| Laboratory Information Systems | Ext. 20150 | 09:00-17:00 Mon-Fri | N/A | |
| Helpdesk <u>cuhit.pathology@hse.ie</u> | | No service on Sat | | |
| Point of Care Testing | Contact No | Opening Hours | Sample Deadline | |
| Point of Care Testing | Ext. 20262 | 09:00-17:00 Mon-Fri | N/A | |
| cuh.pochelpdesk@hse.ie | | No service on Sat | | |

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3.3 Contact Details

| Name | Position | Tel Ext. | E. mail |
|----------------------------|--|--------------|----------------------------|
| General Laboratory Me | dicine | | |
| Ms Sinead Creagh | Laboratory Manager | 22532 | sinead.creagh@hse.ie |
| Mr Paul Cantwell | Laboratory Quality Manager | 20089 | paul.cantwell@hse.ie |
| Ms Brid O'Mahony | Chief Medical Scientist – ICT | 20150 | brid.oMahony1@hse.ie |
| Ms Margaret O'Mahony | Chief Medical Scientist – ICT | 20150 | margaret.omahony4@hse.ie |
| Department of Blood Ti | • | | |
| Dr Oonagh Gilligan | Consultant Haematologist | 20111 | Oonagh.Gilligan@hse.ie |
| Dr Mary Cahill | Consultant Haematologist | 22546 | MaryR.Cahill@hse.ie |
| Dr Cleona Duggan | Consultant Haematologist | 22545 | Cleona.Duggan@hse.ie |
| Dr Derville O'Shea | Consultant Haematologist | 22548 | Derville.Oshea@hse.ie |
| Dr Vitaliy Mykytiv | Consultant Haematologist | 20111 | Vitaliy.Mykytiv@hse.ie |
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| Ms Bridget Lane | Specialist Medical Scientist: Haemovigilance Co-ordinator | 22668 | Bridget.lane@hse.ie |
| Greg O'Connor | Haemovigilance Officer (CUH) | 086 0453551 | Greg.Oconnor@hse.ie |
| Deirdre Harrington | Haemovigilance Officer (CUH) | 086 0453551 | Deirdre.Harrington@hse.ie |
| Ms Connie Foley | Haemovigilance Midwife (CUMH) | 086 7872160 | Connie.Foley@hse.ie |
| Ms Patricia O'Leary | Haemovigilance Midwife (CUMH) | 086 7872163 | Patricia.Oleary@hse.ie |
| Medical Scientist on cal | I in Blood Bank: Bleep No: | 199 | |
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| Dr Sean Costelloe | Consultant Clinical Biochemist | 22530 | Sean.Costelloe@hse.ie |
| Dr Aidan Ryan | Consultant Chemical Pathologist | 34401 | Aidan.ryan1@hse.ie |
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| Ms Katherine Hooley | Chief Medical Scientist | 22535 | Katherine.hooley@hse.ie |
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| Dr Deirdre Broderick | Consultant Microbiologist | | Deirdre.broderick@hse.ie |
| D.I.Ts | Microbiology Registrars / SHO | 22504 /22694 | /20076 |
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| Mr Eddie McCullagh | Chief Medical Scientist | 22505 | Eddie.McCullagh@hse.ie |
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| Mr Liam Blake | Senior Medical Scientist (Surveillance) | 21318 | Liam.blake@hse.ie |
| Medical Scientist on cal | I Bleep No: | 375 | |

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| Dr Viyaliy Mykytiv | Consultant Haematologist | 20111 | Vitaliy.Mykytiv@hse.ie |
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| | (Flow Cytometry) | 21001 | |
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| Katherine Hooley | Chief Medical Scientist | 22535 | Katherine.Hooley@hse.ie |
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| Di Michael W. Definett | Histopathologist | 20490 | mendel.bernett@nse.ie |
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| , | Pathology | | |
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| | Pathology | | |
| Mr Stephen Power | Chief Medical Scientist, | 22572 | stephen.power2@hse.ie |
| | Pathology | | |
| Ms Susan Dineen | Perinatal Specialist Medical | 087 3691513 | cuh.perinatalpath@hse.ie |
| Ms Therese Brosnan | Scientists | | |
| Ms Bríd O'Sullivan | | | |

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| Name | Position | Tel E | ĸt. | E. mail |
|---|---|-------|-----|-------------------------|
| Mr Dan Collins | Mortuary Services Manager | 22525 | | daniel.collins@hse.ie |
| Mr Kevin Lynch | Senior Anatomical Pathology Technician | 22524 | • | kevin.lynch@hse.ie |
| Neuropathology | | | | |
| Dr Niamh Bermingham | Consultant Neuropathologist | 20474 | ŀ | niamh.bermingham@hse.ie |
| Dr Michael Jansen | Consultant Neuropathologist | 20475 | 5 | Michael.jansen@hse.ie |
| An urgent on call service is provided weekdays from 9.00 am Monday to 5.00 pm Friday and a limited on call at certain weekends only. For Neuropathologist on call rota and mobile contact nos. please check with Hospital Switchboard. | | | | |
| Point of Care Testing Department | | | | |
| | | | | Mark.Butler@hse.ie |

3.4 Availability of clinical advice on ordering of examinations and on interpretation of results

- 1. Clinical advice on ordering of examinations and on interpretation of examination results is available and can be obtained by contacting the appropriate clinical team (refer to section 3.3).
- 2. Interpretation and clinical advice is provided on the report where appropriate.
- 3. Refer to section 5.0 for further information regarding the ordering of examinations.
- 4. Refer to the A-Z Test Directory for a list of tests performed, samples required, primary sample volumes, special precautions, turnaround time, biological reference intervals, and clinical decision values.
- 5. Haematology Virtual Clinic provides a service to referring GP's, outpatient clinics, other CUH medical/surgical departments and outside hospitals whereby they receive advice and helpful guidelines from the Consultant Haematologists. The main purpose of this service is to save patients unnecessary trips to the haematology outpatient clinics which are already heavily overbooked. It allows GP's etc to follow up and treat their patients in the community as a result of the advice they receive from the haematology consultants.

3.5 The laboratory's complaint procedure

The goal of Laboratory Medicine is to ensure that our users receive accurate, reliable, meaningful and timely laboratory results. It is your right as a service user of the HSE to make a complaint if you believe that standards of care, treatment or practice fall short of what is acceptable. If you need to make a complaint, we want the process to be easy, effective and fair.

In order to help you to do so please contact the appropriate Department, the Laboratory Manager or the Quality Manager (refer to 4.3 for contact details) or one of the Hospital complaints offiers:

<u>https://www.hse.ie/eng/about/qavd/complaints/officers/hospital/</u>

HSE policy and procedures for 'The Management of Consumer Feedback to include Comments, Compliments and Complaints in the Health Service Executive' can be accessed through the HSE website or by clicking on the following link:

<u>https://www.hse.ie/eng/services/yourhealthservice/feedback/complaints/policy/</u>

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3.6 Policy on protection of personal information

Laboratory Medicine is committed to protecting the privacy of personal information of its service users and patients. In the course of their work, health service staff are required to collect and use certain types of information about people, including 'personal data' as defined by the Data Protection Act 2018. The HSE has a responsibility to ensure that this personal data is;

- obtained fairly
- recorded correctly, kept accurate and up-to-date
- used and shared both appropriately and legally
- stored securely
- not disclosed to unauthorised third parties
- disposed of appropriately when no longer required

All staff working in the HSE are legally required under the Data Protection Act 2018 to ensure the security and confidentiality of all personal data they collect and process on behalf of service users and employees.

Data Protection rights apply whether the personal data is held in electronic format or in a manual or paper based form.

HSE policy and procedures with regards to Data Protection can be obtained through the following link:

http://www.hse.ie/eng/services/yourhealthservice/info/DP/

3.7 Instructions for transportation of samples, including any special handling needs

Instructions for the transport of specimens to the Laboratory are described in a separate procedure for Sample Transportation: PPG-CUH-PAT-36.

NOTE: All Urgent Biochemistry samples should be brought directly to the Biochemistry Laboratory and handed directly to a member of staff

Urgent samples from GP's should be sent in the bag specifically labelled 'Biochemistry Urgent Samples' to allow for prompt processing. A supply of labelled bags is available from Biochemistry.

Please contact the laboratory for information on the correct procedure for centrifugation and specimen storage prior to transport to the laboratory.

All GP Coagulation and Urgent Haematology specimens must be put into a separate transport/delivery bag, labelled 'Coagulation and Urgent Haematology Specimens only' to allow for prompt processing.

Samples for specialised coagulation must arrive into the laboratory within 4 hours of phlebotomy.

Samples for COVID 19 testing and all CSF samples must be delivered directly to Microbiology, the pneumatic tube system should never be used.

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4 TYPES OF CLINICAL SERVICES OFFERED BY THE LABORATORY

4.1 Autoimmune Serology

Autoimmune serology provides a service for the screening and diagnosis of a large range of autoantibody associated diseases. These diseases include Rheumatoid arthritis, Systemic Lupus Erythematosis and Coeliac disease. Immunofluorescence, Elisa and other methodologies are undertaken in this section to detect the presence of autoantibodies in the serum of patients with suspected Autoimmune disease.

While Autoimmune Serology strives to provide a comprehensive in-house service for the more commonly encountered Autoimmune diseases, some auto antibodies - associated with less frequently encountered clinical conditions require off-site analysis. These serum samples are sent to external accredited laboratories for autoantibody determination. Please note that the use of external laboratories will increase the Turn Around Times (TAT's) for these assays.

Examinations referred to other laboratories: Tests not done on-site are referred to outside laboratories for analysis. Test information is included in the test directory.

Information regarding in-house and referred tests is available in the Test Directory. Stated volumes required apply to adult patients. For paediatric samples please send as much blood (up to adult volume) as possible.

Because individual tests are often grouped into profiles, and secondary confirmatory assays are often undertaken, small blood volumes may result in incomplete analysis.

4.2 Department of Clinical Biochemistry

Clinical services offered (including examinations referred to other laboratories) Clinical Biochemistry is a consultant led service that provides a diagnostic, analytical and interpretative service for a large range of analytes in body fluids. Clinical Biochemistry deals with the biochemical basis of disease and the use of biochemical tests for its diagnosis, prognosis, screening and management. The laboratory provides a reliable analytical service and advice on the management of patients with metabolic disturbances.

As well as routine diagnostic work, the Department is actively involved in teaching students of medical science, science, and medicine. The Department has research and teaching links with the Departments of Medicine and Pathology of UCC and with Cork Institute of Technology Biological Sciences Department. The Laboratory is involved in collaborative research with clinical colleagues, international collaborators in the EU IST framework and postgraduate research is also carried out. Staff members contribute as lecturers and project mentors to the UCC/CIT MSc. in Biomedical Sciences. The Royal College of Pathologists recognises the department for higher specialist training in Clinical Biochemistry.

Information regarding in-house and referred tests is available in the Test Directory. Services offered include:

- Routine Clinical Biochemistry e.g. liver, renal, cardiac, bone, glucose
- Lipids, e.g. cholesterol, triglycerides, lipoproteins
- Endocrinology, e.g. thyroid function, infertility testing, pituitary disorders
- Specific proteins, e.g. immunoglobulins, allergies, acute phase proteins

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- Therapeutic drugs
- Cardiac markers
- Toxicology
- Molecular Genetics, e.g. Haemochromatosis

Tests not done on-site are referred to outside laboratories for analysis. Test information is included in the test directory.

For advice on molecular genetic investigations, contact Principal Biochemist (ext 22531).

4.3 Department of Clinical Microbiology

Clinical services offered (including examinations referred to other laboratories)

Clinical Microbiology is a consultant led service that offers a comprehensive range of diagnostic services in routine Bacteriology, Mycobacteriology, Mycology, Parasitology, Infectious Diseases Serology and Molecular Diagnostics as well as consultation in microbiology, infectious diseases and antibiotic utilisation and provision of statistical and cumulative data for infectious disease monitoring. The medical team is available at all times for consultation on any aspect of microbiology and infection control.

In addition to diagnostic services, education and training are an integral part of the daily routine of the department, with established links to the Medical and Science Faculties at University College Cork and the Biological Sciences Department of the Cork Institute of Technology. The laboratory is also involved in teaching both medical and biomedical science students and is involved in collaborative research work with clinical colleagues. The department is accredited by the Royal College of Pathologists for specialist training in Clinical Microbiology.

Information regarding in-house and referred tests is available in the Test Directory. Services offered include:

- 1. Routine Bacteriology: Examination of Urine, Sputum, Blood, CSF and Swabs etc.
- 2. Serological testing for hepatitis, HIV, syphilis, leptospirosis, etc. Please refer to the Test Directory for acceptable sample types for each test. Only the sample types specified will be tested. Any other sample types will be rejected and will NOT be tested.
- 3. Molecular testing for *Chlamydia trachomatis, N. gonorrhoea* and enteric pathogens is performed in-house. SARS CoV 2 and Influenza testing and Respiratory multiplex of performed in-house. Carbapenemase Producing Enterobacteriales (CPE) as approved by the Microbiology Medical Team.
- 4. Parasitology includes the investigation of faeces specimens for evidence of infestation.
- 5. Mycology: Examination of specimens such as skin scrapings and specimens from systemic infections for the presence of pathogenic fungi.
- 6. TB Laboratory: The investigation of specimens for Mycobacterium spp.

Tests not done on-site are referred to outside laboratories for analysis. Test information is included in the test directory.

General collection and transport guidelines:

1. Where possible, collect the specimen prior to the administration of antimicrobial therapy.

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- 2. Collect the specimen with as little contamination from indigenous microbial flora as possible to ensure that the specimen will be representative of the infective site.
- 3. Collect the specimen using sterile equipment and aseptic technique to prevent the introduction of contaminating micro-organisms.
- 4. Collect an adequate amount of the specimen. Insufficient specimens may yield falsenegative results.
- 5. Most specimens collected with a swab and transported dry are unacceptable.
- 6. Identify the specimen source and/or specific site correctly so that proper culture media will be selected during processing in the laboratory. Special requests such as Diphtheria, Actinomyces, Nocardia etc. should be noted on the microbiology request form.
- 7. Specimens should be transported as soon as possible.
- 8. If processing is delayed, refrigeration is preferable to storage at ambient temperature, with the following exceptions:
 - Blood cultures hold specimen at room temperature
 - CSF hold specimen at room temperature do not transport through pneumatic tube system
 - Specimens for the detection of gonococci (keep GC specimens at room temperature)
 - Mycology specimens
- 9. Microbial cultures submitted by other laboratories for further identification should be submitted in pure culture on the appropriate medium in a sealed, screw-capped slope. Petri plates are acceptable if properly sealed for immediate transport.
- 10. Include foreign travel stating country as certain diseases/infections are associated with certain parts of the world.

Note: Telephone the laboratory if the proper procedure is in doubt.

4.4 Department of Haematology and Coagulation

Clinical services offered (including examinations referred to other laboratories) The Haematology Department is a consultant led service that provides a comprehensive range of laboratory tests and clinical support for the management of haematological disorders.

Haematology is a regional laboratory service, in addition to stat and urgent service provision to the theatres, day services, cancer care and accident and emergency departments of CUH/CUMH, the laboratory accepts samples from Cork Dental Hospital, other citywide hospitals which have no laboratory facility (e.g. St. Finbarr's Hospital, South Infirmary Victoria Hospital), and General Practitioners. The Haematology laboratory is the referral laboratory for other HSE-South hospitals Bantry and Mallow and Kerry General Hospital, in which full range of testing is not available. The laboratory serves a catchment area of just over 450,000 for non-routine testing

As well as providing the diagnostic services provided, education and training are an integral part of the daily routine within the laboratory with established links to the Medical and Science faculties at UCC and the Biological Sciences department of the Munster Technological University (MTU). Members of staff regularly teach at both institutions. In

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addition an Irish Committee of Higher Medical Training/Royal College of Pathologists approved structured training programme for Non Consultant Hospital Doctors (NCHDs) is well established within the laboratory as are trainee medical scientist programmes approved by the Academy of Medical Laboratory Science. The laboratory is also involved in both intradepartmental and collaborative research.

Information regarding in-house and referred tests is available in the Test Directory. Services offered include:

1. Routine Full Blood Counts, ESR and Blood films

- FBC consists of a full blood count, which includes the number of red blood cells, white cels, and platelets as well as white cell differential.
- FBC may show evidence of: iron deficiency or Vitamin B12 deficiency anaemias, infection or inflammation, bleeding or clotting disorders, and possible haemolytic anaemias (in conjunction withof hypochromic RBCs, Reticulocyte count, and RBC morphology.
- ESR (Erythorocyte Sedimentation Rate) detects the presence of inflammation caused by one or more conditions such as; infection, tumours or autoimmune disorders or to assist in the diagnosis and monitoring of specific conditions such as temporal arteritis, systemic vasculitis, polymyalgia rheumatic or rheumatoid arthritis. ESRs must be processed within 12 hours of phlebotomy unless stored at 4 ° C.

2. Coagulation

- PT and INR to monitor Warfarin and Di-coumarin therapy
- APTT to monitor intravenous Heparin therapy and the investigation of inherited and acquired bleeding.
- Routine Screen for investigation of bleeding disorders: INR, APTT, Fibrinogen and Platelet Count. In the event of abnormal results occurring in the Intrinsic or Extrinsic Pathways the relevant Factor deficiencies are investigated including screens for Von Willebrand's disease and Inhibitor screens
- Anti-Factor Xa to monitor Low Molecular Weight Heparin therapy
- Platelet function abnormalities are investigated by performing Platelet Function Tests.
- Lupus Anticoagulant screen: PT, APTT, Fibrinogen assay, AFSL, and DVVT. Anticadiolipin and Beta 2 Glycoprotein antibodies are also part of the lupus screen.
- Direct Oral Anticoagulant (Apixaban and Rivoroxaban): do not require routine monitoring. However, monitoring may be required in certain circumstances e.g. when there is concern about adsorption, acute renal impairment, potential drug interactions, to estimate drug levels in the setting of bleeding. Levels should not be used to guide the acute management of a bleed as this can lead to a delay in treatment but can be helpful to differentiate the causes of prolonged bleeding (failure to clear the drug vs consumptive coagulopathy etc.).
- 3. Thrombophilia

Appropriate ordering for Thrombophilia for the investigation of thrombotic episodes must be 6 weeks post thrombotic episode. Patients on anticoagulants are not suitable for Thrombophilia screening. Check BCSH guidelines published December 2010 to prevent unnecessary testing of patients, copy and paste following link to browser for

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guidelines:

www.bcshguidelines.com/documents/Heritable_thrombophilia_bjh_07_2010.pdf Thrombophilia request form FOR-CUH-PAT-1575, including documentation of patient consent, must be received with all requests and is available on the CUH website.

The TAT's cited in the directory for the assays involved in the Thrombophilia Screen, refers to the time that the results are available in the Haematology Laboratory. The TAT for the full report is 3 - 4 weeks.

4. Bone marrow investigations

Bone marrow examinations are undertaken when investigating patients for Leukaemia, Lymphoma, Myeloma, Myelofibrosis and Platelet abnormalities e.g. Thrombocytopenia / Thrombocytosis.

Bone Marrow investigations for add on tests: contact Haematology Laboratory.

5. Flow Cytometry

Flow cytometry is used in the diagnosis and classification of acute leukaemia, chronic lymphoid leukaemia and Non-Hodgkin's lymphoma. The technique employs flurochrome-labelled monoclonal antibodies directed against specific cellular antigens. Abnormal cell populations are characterised by multiparameter analysis, using forward light scatter, side scatter and fluorescence signals to classify /identify each cell type (immunophenotype). Other applications of this technique include immune monitoring and lymphocyte subset analysis, e.g. CD4 count for HIV.

6. Haematinic Assays

Haematinic studies consist of serum B12, Folate and Ferritin assays.

Vitamin B12 and Folate assays are carried out in the investigation of macrocytic anaemias. B12, Folate and Ferritin should be requested for investigation of abnormal FBC results and relevant clinical syndromes.

Use of haematinics for screening of well patients is not recommended. **Requests should be accompanied by clinical details.** When B12 results are low Intrinsic Factor Antibody investigation is carried out. Serum Ferritin assays are performed when microcytic hypochromic anaemia is suspected, or cases of suspected Haemachromatosis. See BCSH guidelines.

The diagnosis of B12 and folate deficiency

http://onlinelibrary.wiley.com/doi/10.1111/bjh.12959/pdf and

Laboratory Diagnosis of Functional Iron Deficiency

http://onlinelibrary.wiley.com/doi/10.1111/bjh.12311/pdf

N.B. Interference in these assays may occur in patients receiving or having diagnostic procedures utilizing monoclonal antibodies.

7. Haemoglobinopathy Screening and Glycosylated Haemoglobin Assays:

Investigation of possible haemoglobinopathy includes the following tests:

- HbS Screening test
- HbA2 Quantitation
- Hb Electrophoresis
- Hb F Quantitation
- HbS Quantitation

Determined using HPLC / Electrophoresis Technologies

Glycosylated Haemoglobin assays are used in monitoring diabetic patients as the levels reflect time-averaged blood glucose levels. HbA1c is an objective test of metabolic

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control, which is independent of the patient's cooperation, the time of day, insulin administration, meals, or exercise and provides the physician with an unbiased indication of the efficacy of prescribed therapy.

8. Kleihauer testing for the estimation of feto-maternal haemorrhage and kleihauer testing for pregnancy loss

Emergency Specimens

Laboratory must be informed of specimens which are emergencies and they will be processed within time frame stated for emergencies for each test.

Examinations referred to other laboratories:

Test information is included in the test directory.

4.5 Department of Pathology

Pathology is a comprehensive consultant led service, which includes Histopathology, Frozen Section, Direct Immunofluorescence, Electron Microscopy, Diagnostic Cytopathology, Neuropathology, Molecular Pathology and a Post mortem service.

Information regarding in-house and referred tests is available in the Test Directory.

Autopsies /Post-Mortems

All persons who die in Cork University Hospital (and CUMH adult deaths) are initially transferred to the mortuary, even if an autopsy is not indicated. A body cannot be released from the mortuary and funeral arrangements cannot be finalised until the mortuary staff can verify whether or not an autopsy will be required.

Please contact the Anatomical Pathology Technician at Ext: 22525 as soon as possible after ALL deaths to help clarify these issues.

Under no circumstances should anyone commit to either scheduling a post mortem or releasing a deceased person, as this is the responsibility of the post-mortem room staff.

Coroner's Autopsies

The following types of death must be reported to the Coroner.

- Where the death may have resulted from an accident, suicide or homicide.
- Where any question of misadventure arises in relation to the clinical or pharmaceutical treatment of the deceased.
- Where a patient dies before a clinical diagnosis is made.
- Where a patient dies within 24 hours of admission to hospital.
- Where the death occurred while a patient was undergoing an operation, or was under the effect of an anaesthetic, or following an operation.
- Where the death occurred during, or as a result of, any procedure.
- Where the death resulted from any industrial disease.
- Where the death was due to neglect or lack of care (including self-neglect)
- Where the death occurred due to hospital service acquired infection
- All deaths occurring in patients who have been referred from a Nursing Home or long term residential care facility
- All deaths in association with Intracerebral haemorrhage
- All deaths occurring in Intensive Care Unit
- All deaths occurring in the Accident and Emergency Department

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• Where death is due to or a contributing factor of alcohol / toxin related cirrhosis / steatosis of the liver or viral cirrhosis of the liver due to IV drug use

Do not ask the next of kin for consent to perform an autopsy examination if any of the above circumstances apply. If you have any doubt as to whether or not a death is properly reportable, consult with the Coroner who will advise accordingly. The fact that a death is reported to the Coroner does not mean that an autopsy will always be required. The Cork City Coroner (Philip Comyn) contactable through the swtichboard.

Cremation

If the family wishes to have the body cremated, the arrangements must be made by them through the Funeral Director/Anatomical Pathology Technician.

It is the policy of Cork University Hospital to refer all documents relating to cremation to the Coroners office for completion. Cardiac pacemakers and/or any radioactive implant must be removed prior to a cremation (and, if appropriate, this action notified to the Coroner).

Consented / Hospital autopsies

Do not ask next of kin for consent to perform an autopsy examination if the death is properly reportable to the Coroner. (See "Coroner's autopsies" above.) The family member granting consent should be the next of kin. Other immediate family members must not object to the examination. The doctor seeking consent (preferably SpR or Consultant) should explain fully to the next of kin the reasons for the examination, the answers sought etc. An information booklet "Information for next of kin/relatives on a hospital request postmortem examination" EXT-CUH-PAT-665 (Form 452) is available which outlines the autopsy examination procedures at CUH and should be offered to the next of kin who is giving the consent.

The Consent to a Post Mortem Examination form (FOR-CUH-PAT-1109 (Form 450)) is quite detailed, but each section is critically important and must be completed in full. Incompletely or incorrectly filled Consent forms will not be accepted.

A Request for Post Mortem Examination form (FOR-CUH-PAT-1214 (Form 451)) must also be completed in full. Provide a brief clinical summary, the presumed cause of death, and list the specific problems to be examined.

The a) Consent form (FOR-CUH-PAT-1109 (Form 450)), b) Request form (FOR-CUH-PAT-1214 (Form 451)) and c) Medical Chart should be delivered to the post mortem room at the earliest opportunity. In addition the case should always be discussed in advance with the pathologist on PM duty.

A Consented/Hospital autopsy service is available at CUH on weekdays. This service is not available at weekends or Bank Holidays. Please note that an autopsy examination requires significant scheduling. Requests received after 11.00a.m. are unlikely to be performed that same day.

Perinatal Autopsy Examination

In the case of neonatal deaths, stillborn infants and foetuses >12 weeks gestational age, the protocol is as for an adult (see above section). Fully informed signed consent of the parent is required.

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Neuropathology

Neuropathology provides a Consultant -provided quality diagnostic service mainly to Cork University Hospital for Neurosurgery, Neurology and Specialised Ophthalmology, outside referrals for approximately 1/3 of the country including all of the Cork hospitals, Tralee and Bantry and referrals from Limerick.

The following information is designed to help you use the Department:

Investigations: These include neurosurgical biopsies, neuromuscular biopsies, temporal artery biopsies, ophthalmic biopsies, CSF for Cytology, CSF for S100, 14-3-3 protein & RTQuiIC and blood for antineuronal antibodies. For advice regarding investigations contact the Consultant Neuropathologist ext 22520.

Request Forms. Please use the designated neuropathology request form for all requests. This is light grey (copies available from the Dept. extension 22520)

Patient Details. Please fill out the patient details correctly. Sticky labels are the best. Essential information for tissues must include patients MRN, full name, address, date of birth, nature of the specimen, hospital location, consultant to whom the report should be sent and relevant clinical information.

Protocols. Protocols for most investigations including muscle and nerve biopsy are available. Neurological/medical teams requesting surgeons to perform a biopsy should complete all the details on the neuropathology request form to accompany the patient to theatre. Please indicate the doctor to whom the results should go.

Autopsies/Brain referrals. For post mortems /Brain referrals on CNS disease cases please contact the Consultant Neuropathologist on duty. (Ext 22520). Coroner's cases and Consent Autopsy protocols are shared with Histopathology (see Histopathology section). Post mortem examinations that are required for investigation of unexplained or incompletely investigated rapidly progressive neurodegenerative disease/ dementia [i.e. where prion disease (transmissible spongiform encephalopathy) has not been satisfactorily excluded from the differential diagnosis) are not carried out in this institution as required biocontainment facilities are not available. For information please ring ext 22520 or the post mortem room ext 22525.

High Risk Cases. Special precautions are required for investigations on atypical dementia and other high risk cases. Fresh CNS, CSF or tissue samples must be treated carefully and decontaminated according to recommended guidelines. Please consult the Neuropathologist on duty for advice. (ext 22520)

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4.6 Point of Care Testing (POCT)

The Point of Care Testing Department consists of a Chief Medical Scientist, Senior Medical Scientist and a Senior Biomedical Engineer to oversee the day-to-day running of POCT devices. POCT devices are situated outside the laboratory and give high quality results if used and maintained correctly. POCT Devices **MUST NOT** be used unless you have been trained. Training courses are organised periodically by the Point of Care Testing Department. Follow the instructions for the disposal of waste in order to minimise health, safety and cross infection risks.

- Blood Gas Analysers: Analysers are located at all Critical Care Areas and in excess of 100,000 Blood Gases are performed annually in CUH. Blood Gas Analysers are located in the Emergency Department, Intensive Care Units (General and Cardiac/HDU), Theatres, CUMH Neonatal Units and Labour Wards, Cath Labs, Ward 5B and Ward GC.
- Blood Glucose Meters: Blood Glucose Meters are located throughout the Hospital to monitor known diabetics and to detect Hyperglycaemia and Hypoglycaemia. Glucometers are not to be used for the diagnosis of diabetes mellitus, for which blood specimens must be sent to the laboratory (Fasting and 2 hr Post-Prandial samples). 250,000 POCT Glucose measurements are performed annually in CUH.
- 3. **PCR Testing for SARS-CoV2/ FluA/Flu B:** COBAS Liat POCT analysers are located in the Emergency Department for POCT SARS-CoV2/ **FluA/Flu B** testing. This POCT service is to support Laboratory Testing and provides short turnaround times that can improve patient triage processes.
- POCT Creatinine: iStat Alinity is located in Radiology department for POCT Creatinine testing. This service is only to be used where a recent laboratory Creatinine measurement is not available.
- 5. **POCT HbA1c:** DCA Vantage for POCT HbA1c testing is located in the Paediatric Diabetic Day Unit and must only be used for patients who are attending this clinic.

Point of Care Testing Steering Group: The purpose of the Point of Care Testing Steering Group is to provide Clinical Governance of the POCT Service by ensuring that systems and processes for monitoring and improving the quality of POCT services are in accordance with best practice. Membership includes the Clinical Director of Diagnostics, Consultant Clinical Biochemist, Consultant Microbiologist, Members of Hospital and Laboratory management, Chief Medical Scientist POCT, Nurse Management, Hospital IT, Biomedical Engineering and Clinical Representatives from relevant areas. Applications for new POCT Services, or extensions to existing services, can be submitted to the POCT steering group for consideration.

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5 INSTRUCTIONS FOR PATIENT-COLLECTED SAMPLES

5.1 Faeces / Stool Sample Collection

- 1. Specimen containers are available from the clinical area or general practitioner. Faeces /stool specimens are submitted for microbiology from patients with diarrhoea or stomach upset. Sometimes, a stool is sent on a person that has had close contact with a person that has had diarrhoea.
- 2. The container should be labelled with your full name, date of birth (or your Hospital Chart number if you have it), date / time of collection and the sample type, i.e. Faeces.
- 3. The sterile container should not be opened until you are ready to collect the sample.
- 4. Wash and dry your hands.
- 5. Do not submit faeces contaminated with urine or toilet water. Urinate into the toilet if needed.
- 6. Place plenty of lavatory paper in a clean potty or in the lavatory pan. Make sure there is no trace of disinfectant or bleach present, as this will interfere with the test. Faeces (a bowel movement) should then be passed on to the toilet paper. Do not send stool wrapped in toilet paper to the laboratory
- 7. **Note:** If you have severe diarrhoea or a watery stool, a potty may be needed to collect the initial sample.
- 8. Open the container and, using the 'spoon' that is provided, transfer enough stool in order to fill approximately 1/3 of the container. Do not overfill the container. Also please ensure that the outside of the container is not soiled with stool.
- 9. You should ensure that the lid of the container is firmly closed. Note that a leaking container may be infectious. Place the container into the specimen bag attach to the laboratory request form.
- 10. Flush away the remaining paper and faeces down the lavatory.
- 11. Wash and dry hands thoroughly with soap and warm water.
- 12. Specimens should be brought to the laboratory as soon as possible.

5.2 Mid Stream Urine (MSU) Collection

- 1. Specimen containers are available from the clinical area or general practitioner.
- 2. The aim of collecting a mid stream urine sample is to help the doctor decide if you have a urinary tract infection (UTI or "kidney infection"). A 'mid-stream' sample is the best sample as the first urine you pass may be contaminated with bacteria from the skin.
- 3. The container should be labelled with your full name, date of birth (or your Hospital Chart Number if you have it), date / time of collection and the sample type, i.e. MSU.
- 4. The sterile container should not be opened until you are ready to collect the sample.
- 5. Prior to collection the genital area should be cleaned with tap water. Antiseptics should not be used. If the area is soiled, use soap and water and rinse thoroughly.
- 6. You should pass some urine into the toilet (discard the initial part of the urine sample); then without stopping the flow of urine, catch some urine in the sterile container (approximately half full). You should then finish passing urine into the toilet. Some specimen bottles contain boric acid preservative (red top container

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with white powder in it). Do not discard the white powder. Fill boric acid container to the line marked, close the lid and mix well. This gives the correct concentration of preservative. Do not use urinary dipstick on boric acid samples as this leads to erroneous results.

- 7. You should ensure that the lid of the container is firmly closed and place the container into the specimen bag attached to the laboratory request form.
- 8. Specimens should ideally be brought to the doctor's surgery or laboratory within 2 hours of collection. If that is not possible the sample should be refrigerated until it can be brought to the doctor's surgery or laboratory.
- 9. Wash and dry hands thoroughly with soap and warm water.

5.3 24 hour collection of urine

Key Points;

- Ensure that you are provided with a collection bottle (brown container) for the 24 hour urine collection before you leave the hospital.
- All of the urine passed during the 24 hour period should be collected. Failure to collect all urine may invalidate result.
- An exact timing of the 24 hour period is required.
- Ensure container is labelled with patient's full name, date of birth, date of collection and time collection was started and time collection was finished.
- Do not void urine directly into the 24 hour container but into a suitable clean detergent free container and then pour urine into the 24 hour container.
- If the container contains a preservative, please exercise care when adding urine to the 24 hour container avoiding splashing.
- Keep container away from children at all times.

Procedure;

- 1. Empty your bladder at 8am on rising or at a more convenient time and discard that sample. The collection period has now started. Write start time on container.
- 2. Collect all urine passed during the next 24 hours and place in container.
- 3. On the following morning empty your bladder at 8am on rising (must be the same time as starting time) and add this sample to the collection. The collection is now complete. Write the finish time on the container.
- 4. Close the container cap securely and ensure container and request form contain required information
- 5. Bring collection to the laboratory on the day of completion.

Incomplete collections;

- 1. If you forget and lose a sample down the toilet, then discard all urine collected up to that time and start collection again.
- 2. If the collection requires a preservative return the container to the laboratory and request a new container.

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5.4 Sputum Sample

- 1. Specimen containers are available from the clinical area or general practitioner. Sputum samples are submitted for microbiology from patients with a chest infection
- 2. The container should be labelled with the your full name, date of birth (or your Hospital Chart number if you have it), date / time of collection and the sample type, i.e. Sputum
- 3. Gargle and rinse mouth with tap water to remove food particles and debris. DO NOT use mouthwash or brush teeth with toothpaste immediately before collection.
- 4. Open the container and hold very close to mouth.
- 5. Take as deep a breath as possible and cough deeply from within the chest. DO NOT spit saliva into the container. Saliva is not a suitable specimen for examination. The specimen should look thick and be yellow or green in colour. There may be fluid with some green or yellow material.
- 6. Avoid contaminating the outside of the container. Close the lid tightly when specimen has been obtained.
- 7. Place specimen in plastic bag section of request form and seal bag.
- 8. Bring the container and form to your GP or the laboratory as soon as possible.
- 9. If there is unavoidable delay in transporting the specimen to the GP or Laboratory, it may be stored in a refrigerator prior to transportation. Prolonged delays will affect test results.
- 10. All sputum specimens should be transported to the laboratory in tightly capped containers placed in the plastic bag (attached to the form).
- 11. This should ideally then be placed in another leak-proof container before transport to the laboratory.
- 12. Specimens for TB testing:
 - a. Three specimens are usually required. Take the specimens on 3 consecutive days. The ideal time to collect the specimens is early in the morning just after getting out of bed.
 - b. Collect and transport all specimens as described above.

5.5 HbA1c collection

- 1. Wash your hands and dry thoroughly
- 2. Increase the needle size of your testing pen by two markers
- 3. Remove the top from the PINK blood bottle
- 4. Prod your finger
- **5.** Blood needs to be dripped into the bottle
- 6. Ensure SMALL label with all relevant details is stuck to the smaller PINK topped bottle
- 7. Place small bottle in the larger universal container (MSU bottle), then in specimen bag
- 8. Seal plastic bag and fill in all details on form provided
- 9. Place in a padded/well protected envelope
- 10. Post the specimen/deliver to: CODE UN 3773, Haematology Dept, Cork University Hospital

Blood sample must be submitted at least 2 weeks before clinic visit

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6 ORDERING LABORATORY EXAMINATIONS

6.1 Instructions for completion of the request form

- 1. For accurate identification of patients and specimens, it is essential that request forms be completed fully, legibly and accurately. Please remember that inadequate information on request forms makes it impossible to issue a report to the correct location or contact the doctor in case of urgent or unexpected results.
- 2. The laboratory has a number of different request forms most of which are colour coded for the department. Multiple tests for one department can be sent on one request form but separate specimens and request forms are required if tests are being sent to a different department or where the sample types are different. Request forms are issued from Hospital Stores. Order supplies in advance to facilitate timely delivery.
- 3. The electronic request using Dedalus Clinical Manager (iCM): Refer to section 10: Information Technology.
- 4. The use of patient addressograph labels on request forms is recommended, except for Blood Transfusion Laboratory requests which must be hand written. On all requests forms, complete the following:
 - a. Patient's Full Surname and Forename
 - Patient's MRN (Medical Record Number). If a MRN is not available or relevant (i.e. GP patients) a date of birth and address must be supplied on the form and specimen label.
 - c. Patient's Date of Birth
 - d. Patient's Sex and Title
 - e. Date and time of specimen collection
 - f. Name of the Requesting Consultant
 - g. Location to where the results should be reported
 - h. Type of specimen collected and if appropriate, the anatomical site of origin or tick the relevant box
 - i. Clinical information relevant to or affecting sample collection, examination performance or result interpretation (e.g. history of administration of drugs).
 - j. Name and bleep number of requesting doctor
 - k. Analysis required
- 5. If a specimen is urgent please indicate on request form and the request will be prioritised. If results are extremely urgent please contact the relevant department to discuss your requirement. Overuse of the urgent service will adversely affect the turnaround time for all urgent tests.
- 6. Clinical details and relevant treatment information and details of foreign travel are extremely useful to the laboratory in interpreting results.
- 7. Refer to the A-Z Test Directory in this User Handbook for a list of tests performed, the sample required, turnaround time and other information regarding specimen collection. The pathologist, clinical biochemist and/or laboratory staff should be consulted where uncertainty exists about the availability, appropriateness, or selection of tests, the nature of the specimen required, or the interpretation of results.

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6.2 Format of Addressographs

The format of the labels should meet the following criteria. The type size should be a **minimum of font size Arial 12** and follow the format

| First name | Surname |
|--------------------|------------------|
| Date of birth | Sex |
| Patient address | |
| ***Space*** | |
| Date and time of s | ample collection |

Please complete the clinician location code and clinician name code on the right hand side of the request form in the space provided. Contact the clerical office to find out your clinician and location codes if you do not have them. It is important that the clinician name does not appear above the patient name as this will inevitably lead to errors.

Contact your software provider to ensure that your labels meet our minimum requirements.

6.3 Criteria for accepting and rejecting samples

The laboratory makes every effort to ensure that samples are processed as requested. However samples must be appropriate for the requested investigation, the safety of laboratory staff must not be threatened and there must be no ambiguity as to the identification of the patient. The criteria for sample acceptance, as described below, are strictly adhered to in the interest of patient safety. Failure to provide the required data shall lead to rejection of the specimen and request form.

| 6.3.1 Biochen | nistry, Haematology, Microbiology, Path | ology |
|----------------------------|---|--|
| Labelling Requirements* | Essential Information | Desirable Information |
| Request Form | Patients full name or proper coded identifier** D.O.B. and/or Patient's Medical Record Number (MRN/RID) Patient's location or destination for report or patient's consultant or GP | Patient's address Patient's sex Clinical details, relevant therapy and foreign travel (antibiotic treatment important for Microbiology), travel and prophylaxis history for Malaria |
| | Specific requirements of individual departments: Biochemistry: Date and time of specimen collection Clinical details Note: Certain analytes may not be processed if mandatory fields are incomplete Request must come from a Qualified Healthcare Professional. Patient's address Patient's sex - | Date and time of specimen collection (timing in relation to antibiotic dose essential for Antibiotic Assays and for some Chemical Pathology tests) Pathology: Date and time specimen taken. Previous relevant Histopathology Numbers (CUH/MUH) if applicable). Signature of clinician / nursing staff (pp) Clinician's bleep number |
| | Haematology /Microbiology: - Test Request Pathology/Cytopathology: | Clinical Information |

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| | Requesting Clinician, | |
|----------------|---|--|
| | Patient's address, | |
| | Patient's location, | |
| | Nature and site of specimen (including Right or Left) | |
| | Destination for report | |
| Sample | Patients full name or proper coded identifier** D.O.B. and/or Patient's Medical Record Number (MRN/RID) All non-blood samples: sample type or exact site Neuropathology: Autopsy brain specimens must be labelled with the PM number, the referring Pathologist and the date of the PM. Further details are at discretion of referring | Pathology: Date and time specimen taken. |
| | Pathologist. Perinatal UHK and CUMH specimens: The | |
| | CUMH uses the MN_CMS Millennium Electronic record. The number of the label on | |
| | the container must match the order number of the request. | |
| Requests using | Samples requested using iCM have no accompa | nying forms. |
| iCM | Details must be complete on the sample contair | ner. |

* The identifiers which appear on the sample container must match the information provided on the accompanying request form **e.g. HIV specimens

| 6.3.2 Blood T | ransfusion | |
|----------------------------|---|---|
| Labelling Requirements* | Essential Information | Desirable Information |
| Request Form | Addressographs on forms <u>not</u> accepted. Patient's Forename [§] Patient's Surname [§] Patient's Sex D.O.B. Medical Record Number (MRN/RID) Patient Address for Out-patients. Destination for report. Patient's consultant or GP. Identity of person taking the samples (Doctor's MCRN or Nurse/Midwife Bord Altranais PIN if possible) including contact details of person taking the sample (e.g. Bleep or telephone). Date and time of specimen collection. Tests Required. [§] For patient's whose identity is unknown (e.g. Unconscious or Major Emergency scenario) the use of pseudonyms/MRNs as per Emergency Department protocols will be accepted. Note: The CUMH uses the MN_CMS Millennium Electronic record. Transfusion forms generated correctly through the MN_CMS EHR are accepted in the CUH Blood Transfusion Department. | Clinical details. Previous address & patient's maiden name Transfusion & obstetric history & relevant therapy. |
| Sample | Addressographs on samples <u>not</u> accepted. Patient's Forename [§] | |

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| | Patient's Surname [§] |
|----------------|---|
| | Patient's Sex |
| | D.O.B. |
| | Medical Record Number (MRN/RID). |
| | Identity of person taking the samples |
| | Date and time of specimen collection. |
| | |
| | [§] For patient's whose identity is unknown (e.g. Unconscious or Major Emergency scenario) the use of pseudonyms/MRNs as per Emergency Department protocols will be accepted. |
| | Note: The CUMH uses the MN_CMS Millennium |
| | Electronic record. Transfusion specimen lables |
| | generated correctly through the MN_CMS EHR are |
| | accepted in the CUH Blood Transfusion Department. |
| Requests using | Blood Transfusion Samples are not to be Requested using iCM and will not be |
| iCM | processed. |

*The identifiers, which appear on the sample container, must match the information provided on the accompanying request form

6.4 Time limits for requesting additional examinations

Users may request additional examinations on specimens already sent to the laboratory. To request the add-on tests use the form titled "Request Form for Additional Tests on Sample Previously sent to Laboratory Medicine" reference FOR-CUH-PAT-1732.

Analyses for additional tests are subject to the stability of the analyte. The analysis will be performed provided the specimen has been stored appropriately and there is sufficient specimen remaining to perform the additional tests.

| Department | Time Limit | | | | |
|------------------------|--|--|--|--|--|
| Autoimmune Serology | Within the 14-day specimen retention time (dependant on storage facilities) and subject to individual analyte stability. | | | | |
| Biochemistry | CK CSF LDH CG-B P | | | | |
| | Total and Direct Bilirubin Oestradiol | | | | |
| | Please contact the laboratory with any queries. | | | | |
| Haematology | Not all add-on tests can be accommodated; the factors influencing the capability of requesting Add-On Tests include storage requirements and stability of parameters measured. Please contact the laboratory if in doubt. The following is not an exhaustive list: - Retics on FBC specimens <12 hours post phlebotomy - ESR <12hrs | | | | |
| | - Blood Films: Manual differential and Red cell morphology <12 hrs, slide Platelet check <72 hrs. | | | | |
| | DDI on Coagulation Sodium Citrate <24 hours post phlebotomy APTT on Coagulation, Sodium Citrate specimens <4 hours post phlebotomy HbA1c on FBC specimens 48 hours after receipt in laboratory Haemoglobinopathies on FBC specimens 48 hours after receipt in laboratory | | | | |
| | Haematinics on clotted specimens – extra assays 48 hours after receipt in laboratory | | | | |

The time limit for requesting additional examinations for each department is given below:

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| | Flow Cytometry on FBC specimens - contact laboratory Fibrinogen <12 hours post phlebotomy Malaria on an FBC sample (12 hours of phlebotomy) Kleihauer: the time limit is 72 hrs post delivery G6PD <24 hours | |
|-----------------------|--|--|
| Microbiology | Infectious Diseases Serology – Blood samples are stored for approximately 1 week from reception date, therefore, additional testing can be requested at any stage during this time. CSF samples are stored for approximately 2 week from reception date, therefore, additional testing can be requested at any stage during this time. | |
| Molecular Genetics | Factor V Leiden and Prothrombin gene mutations - add on not possible as separate specimens always required for genetic testing | |

Please contact the appropriate laboratory for more detail on the time limits for requesting additional examinations

6.5 List of factors known to significantly affect the performance of the examination or the interpretation of the results

Many sources of error exist that could affect the examination result. Refer to the A-Z Test Directory in this User Handbook for any special rejection criteria that may apply. Listed below are some of the major pre-examination reasons for test cancellation or delay.

Request form problems that will cause test cancellation or delay:

- Illegible patient demographics, illegible name of ordering clinician or incorrect ward /location
- Absent or incorrect patient identifier (e.g. MRN/RID or PPI)
- Absent or incorrect time and date of request
- Unclear or totally absent marking of test request boxes
- Type of body fluid not identified
- Form contaminated by specimen

Specimen problems that will cause test cancellation or delay:

- Leaking containers (rejected because of infection risk)
- Sample is unlabelled, incorrectly labelled or does not match the accompanying form
- Too few specimens or an insufficient volume for analysis. Send separate samples for each department. Split a CSF sample when requesting both cell count/culture and biochemistry. Send separate samples for in-house and send-out (reference laboratory) tests
- Misrouting of specimens e.g. inappropriate laboratory
- Incorrect lab request form used
- Sample collected into an incorrect preservative/anticoagulant
- iCM labels containing bar codes must be aligned with the original container label Note: Large loose labels on specimens cause loss and damage to samples and costly damage to analysers

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7 SPECIMEN COLLECTION

7.1 Instructions for preparation of the patient

Patients can help to ensure that their lab tests are accurate by following pre-testing instructions carefully and by providing complete medical histories, including lists of medications to their health care providers.

Variables that could affect test results

- Patient variables including exercise, diet, age, sex, circadian variation, posture, obesity, stress, smoking and medication may affect laboratory test results.
- An individual's diet and lifestyle may affect laboratory test results. It is generally recommended that the night before laboratory tests patients avoid high-fat foods, alcohol and strenuous exercise.
- Patients should ask their doctors if certain medications should be stopped prior to lab testing as certain medications may interfere with the laboratory test results.

Blood Tests

- Patients may need to fast prior to certain blood tests. For example, patients should not eat or drink anything except water for 9 to 12 hours prior to glucose and lipid profile tests.
- The amount of blood drawn at the time of collection for laboratory testing depends on the tests that are ordered. Usually the amount collected is very small (around 3-6 teaspoons.)
- Some patients become anxious when they have their blood drawn. Patients should tell the health care professional who is drawing the blood if they feel faint or sick. Slow deep breaths prior to the needle stick may help to alleviate anxiety.
- After a blood draw, the phlebotomist makes sure that all signs of bleeding have stopped. A bandage is applied to the arm for a minimum of 15 minutes.
- Aspirin or other anticoagulant (blood thinners) drugs can prolong bleeding. In such cases, patients may need continued applied pressure until the bleeding has stopped. A cold pack may be necessary to reduce swelling and bruising.
- After a patient has blood drawn, even when bleeding has stopped, patients should not carry or lift a heavy object with that arm for a minimum of one hour.

SARS CoV 2 sampling

 Refer to HSE link below for video <u>https://www.hpsc.ie/a-</u> <u>z/respiratory/coronavirus/novelcoronavirus/guidance/infectionpreventionandcontrolgui</u> <u>dance/sampling</u>

Collecting Specimens at Home

- Patients must follow all instructions exactly for collection of specimens performed at home then brought to the laboratory for testing.
- Special containers with a powder or liquid preservative may be provided for urine collection. Patients should never empty or discard any powder or liquid from the container before beginning the collection of a specimen.
- Specimens should be delivered to the laboratory in the prescribed timeframe in order to assure accurate results.

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Results

- Depending on the laboratory work performed, test results may be available within a few hours to as long as several weeks.
- Laboratory test results are often reported with a reference interval to assist the clinician in interpreting them. These reference intervals reflect the values in the majority of healthy individuals; however, a small number of healthy people (5%) may have results that are higher or lower than those in the reference range. Therefore, laboratory results should interpreted by clinicians who can decide whether or not the results indicate a medical condition.
- Clinicians consider personal medical history, family history, and results from physical examination when interpreting an individual patient's laboratory test results.

7.2 Phlebotomy Service at Cork University Hospital

Senior Phlebotomist: Ms Lynne Heeney

Contact Numbers: Phone: 22415 (Blood Room) 22353 (Phlebotomy office)

Phlebotomy is based in the Out-Patients Department for Warfarin clinic and Oncology Clinics. All other Out-Patients and GP patients are required to attend Blood room in St.Catherines by appointment only.

Wards: The service is Monday to Sunday Electronic orders **must be placed before 6.30**. Weekend /Bank Holiday for non-routine bloods, limited services.

Awbeg suit Blood Room: Warfarin and Oncology patients **ONLY**

Warfarin Clinic Monday – Friday 7:30 – 13:00 (except Wednesday) Oncology Bloods Monday – Friday 9:00 - 13:00 Monday – Thursday 14:00 – 16:30

Cedar Building Blood Room

Diabetes & Endocrine patients **ONLY** By apointment **ONLY ONLINE booking available <u>HERE</u>.**

Phone number 021/423-4910

Tuesday 7:30am-13:00pm and 2:00pm-4:30pm Wednesday 7:30am-13:00pm Thursday 7:30am-13:00pm and 2:00pm-4:30pm

St. Catherines Blood Room:

All other Consultant clinics By apointment **ONLY ONLINE booking available <u>HERE</u>**. Phone number: 021/423-4910 Monday – Thursday 7:30 – 16:30 Friday 7:30 – 16:00

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The Phlebotomy Department provides a varied service within the hospital. It covers the Paediatric wards, all the adult wards, the psychiatric unit and the Emergency Department. The Blood Room clinic provides an important Paediatric out-patients service to the General Practitioners in the City and County.

Health and Safety

- Universal precautions are adhered to at all times.
- Gloves to be used when dealing with patients.
- Gloves to be changed after each patient.
- Needles not to be recapped after use.
- Needles and Holders to be disposed of safely.
- Sharp bins provided for disposal of sharps.
- Clinical waste bags provided for any bloodstained material.
- Spillages /blood Appropriate disinfectant to be used to clean and disinfect.
- Large spillages of blood /body fluid contact Housekeeping (protocols laid down by infection control)

Prion Disease:

- 1. It is essential that all CSF samples from patients who have Prion Disease in their differential diagnosis be managed in the following manner
- 2. Each laboratory likely to receive the CSF must be informed.
- 3. The sample and form should be appropriately labelled.
- 4. Information regarding suspected Prion disease MUST be indicated on the request form
- 5. The CSF, in a universal container, is double-bagged and marked with a biohazard label.

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7.3 Phlebotomy blood collection order of draw

| Specimen Volume | Order Of Draw | Closure Colour | Tube Contents | Assays |
|--------------------|------------------|-------------------|---|--|
| 5ml | 2 | | | Prior to collecting blood samples from a newly inserted peripheral venous |
| 3ml | | Blue | Trisodium Citrate solution | Coagulation Studies |
| 4ml | 0 | Red | Separation Gel Clotting Accelerator | Biochemistry Profiles, Viral Studies, Hormone Studies, Immunology, Anti Cardiolipin AB., B12, Folate, Ferritin, RA, Intrinsic Factor AB, Iron Studies, CRP's, TDM (Therapeutic Drug Monitoring), |
| 4ml | | Red | Clotted (Gel free) | Cryoglobulins, Methotrexate |
| 4ml | | Green | Heparin | Chromosomes, Lead Levels, DNA Analysis |
| 3ml | | Purple | EDTA | FBC, HBA1C, Hb. Electrophoresis, Malaria Parasites, Sickle Cell, Reticulocyte Count, Coombs Test, Cyclosporin,Tacrolimus ESR, Immunophenotyping, PTH, Cryogobulins |
| 6ml | 0 | Pink | EDTA | Crossmatch, Group & Antibody Screen |
| 4ml | | Grey | EDTA sodium fluroide | Glucose, Fluid Glucose, Glucose Tolerance,Lactate, Alcohol Levels |
| 9ml | | Yellow | ACD-A | HLA Typing |

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7.4 Minimum Sample requirements for Paediatric/neonatal patients

The volume of serum/plasma obtained from blood depends on the haematocrit; therefore measurement of these analytes may require a larger volume of blood from patient with high haematocrit.

| Test | Sample Type | Minimum Volume | Additional Requirements |
|--|--|-----------------------------------|--|
| U/E, Creat, Ca, Mg, Phos,Bili, Lfts | Li Heparin or clotted sample (orange top/clear top) | 1ml | |
| TFT's | Li Heparin or clotted sample (orange/clear top) | 0.75ml | |
| Glucose | Fluoride oxalate (yellow top) | 0.5ml | |
| Ammonia | Li Heparin (orange top) | 0.5ml | |
| Blood amino acids | Li Heparin (orange top) | 150ul | |
| Urine amino acids | Urine | 4mls | |
| Organic Acids | Urine | 4mls | |
| Acylcarnitine | Blood spot | | |
| Very long chain fatty acids | EDTA or Lithium Heparin | 2ml | |
| Lysosomal enzymes | EDTA | 5ml | 16 enzymes measured here, specific enzymes can be requested with a sample volume of 3ml |
| Transferrin isoforms | Clotted sample (Clear top) | 0.75ml | Not for babies <3 weeks |
| Biotinidase | Li Heparin | 0.5ml | Frozen in <1hour |
| Free fatty acids and β- hydroxybutyrate | Fluoride oxalate | 2ml | |
| Insulin and C-peptide | Clotted sample | 2ml | Haemolysed samples unsuitable |
| Growth Hormone | Li heparin or clotted sample | 1ml | |
| Cortisol | Li heparin or clotted | 0.75ml | |
| 17-hydroxyprogesterone | Li heparin or clotted | 1 ml | Only after 48hrs poat birth |
| Mycophenolate | EDTA | 1ml | Spin <6hrs |
| Haematology Test: FBC | EDTA | 1mL purple or 1.3 mL red | |

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7.5 Sample Storage Conditions

Biochemistry

- 1. Store blood and urine samples at **room temperature**, unless otherwise specified.
- 2. For the addition of test requests to existing samples, please contact the laboratory for advice on sample integrity.
- 3. If a delay arises, please contact the laboratory for advice on sample integrity (Tel: 021-4922528)

Haematology

- 1. If delays are unavoidable, Haematology specimens can be preserved by refrigeration at 2-8°C in a designated specimen fridge e.g. Full Blood Counts, HbA1c, Haematinics
- 2. Coagulation samples for INR must be stored at 18-22°C (Refrigeration may lead to cold activation of coagulation factors)
- 3. Addition of test requests to existing samples is not recommended due to issues of sample integrity. Contact laboratory for advice.

Exceptions to this include:

- a. Coagulation specimens for APTT need to be assayed within 4 hours of phlebotomy
- b. Samples for Flow Cytometry should be sent to the Haematology ASAP, ideally on the day of Venesection, at room temperature. If a delay is anticipated and is needed to be kept overnight, store at 2-8°C in a designated specimen
- c. Malaria tests must be examined on the day of venesection, therefore is not suitable for storage
- d. Bone marrows and Kleihauer (Foetal cells) must be sent immediately to Haematology

Microbiology

- 1. In most cases, if delays are unavoidable, microbiology specimens can be preserved by refrigeration at 2-8°C in a designated specimen fridge, as this maintains the viability of the pathogens present and prevents the overgrowth of non-pathogenic bacteria. Exceptions to this include:
 - a. Blood Cultures Do not refrigerate or place on radiators, incubators or direct sunlight. The pneumatic tube can be utilised to transport **plastic** blood culture vials and is preferable to avoid unnecessary delays.
 - b. CSF should be held at room temperature.
 - c. Samples specifically for the isolation of *Neisseria gonorrhoea*. (i.e. cervical or urethral specimens) should be stored at room temperature. The viability of *N. gonorrhoeae* is lost over time.
 - d. Faeces Samples for Ova, Cyst and Parasite investigation should not be refrigerated (should be stored at room temperature).
 - e. Molecular Investigation: Viral swabs for SARS CoV-2 and other Respiratory Viruses are provided directly from the Microbiology Department and should be transported to the laboratory without delay. If delay is unavoidable, please store at 2-8°C.
 - f. Collection swabs for Molecular Investigation of Carbapenemase Producing Enterobacteriales (CPE), will be provided by the Microbiology Derpartment by liaising with Medical Microbiology Team and should be transported to the laboratory without delay. If delay is unavoidable, please store at 2-8°C.

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Microbiology (Infectious Diseases Serology)

Clotted Blood and EDTA Blood for Molecular Investigations

Serum and plasma must be removed and frozen at \leq -20°C by the laboratory within 24 hours of venepuncture to maintain the integrity of viral nucleic acid. Therefore, samples must be sent to the laboratory without delay. Samples received greater than 24 hours from collection will NOT be processed.

Clotted Blood for Serological Investigations

Specimens should be transported to the laboratory without delay. If delay is unavoidable, please store at 2-8°C.

<u>Oral Fluid</u>

Oral fluid specimens should be collected using commercially available collection devices such as OraCol[™] or OraSure[™]. Please contact the laboratory for further information. Please transport without delay (particularly for molecular investigations). If delay is unavoidable, please store at 2-8°C.

Respiratory Secretions

Respiratory viruses are extremely thermolabile and therefore should be transported to the laboratory without delay. The quality of the sample is a major determinant in identifying the causative agent. If delay is unavoidable, please store at 2-8°C.

<u>Stool</u>

For molecular detection of viruses associated with gastroenteritis, specimens should be transported to the laboratory as soon as possible post collection. Alternatively, specimens may be stored at 2-8°C for up to 72hrs before dispatch.

Stool for Strongyloides culture or Ova, Cyst and Parasite investigation must NOT be refrigerated. Send to the laboratory without delay.

<u>Urine</u>

Specimens should be transported without delay (particularly for molecular investigations). If delay is unavoidable, please store at 2-8°C.

Viral Swabs

Swabs should be transported to the laboratory without delay. If delay is unavoidable, please store at 2-8°C.

Pathology

Prolonged formalin fixation may have an adverse effect on subsequent molecular techniques. Specimens in Buffered Formal Saline should be stored at ambient temperature.

Neuropathology:

- 1. CSF/CNS fluids should be stored at 4°C if any delay occurs prior to delivery to the laboratory.
- 2. Any details of storage conditions should be recorded on the form.

Cytopathology:

Samples for cytological examination will deteriorate with time and should therefore be transported to the laboratory as soon as possible. In the event of a delay, samples should be stored at 2-8°C.

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8 **REPORTING OF RESULTS**

8.1 Turnaround Times

Turnaround time (TAT) is given as the maximum number of working hours/days between sample receipt and issuing a report either in the computer or by phone under normal operating conditions. In addition to the routine service each department operates an "urgent" system whereby the target turnaround time is shorter. The turnaround time for individual tests is given in the A-Z Test Directory in this User Handbook.

Overuse of the urgent service will adversely affect the turnaround time for all urgent tests. Many specialised tests are performed on a weekly basis; if such tests are required urgently please phone the appropriate laboratory to discuss the request.

TAT are routinely monitored as part of the laboratories quality improvement program.

8.2 Critical Results Reporting

Critical results will be communicated by the laboratory, therefore it is essential that up to date contact details are available for the routine day and out of hours. The laboratory requires phone details that are appropriate to receive critical results in a timely manner from all users.

| Biochemistry | , | | |
|---------------------------|--|---|--|
| Test | Result | Test | Result |
| ALT | >510 U/L (Female) >675 U/L (Male) | Glucose | <2.5 mmol/L >25 mmol/L ≥15 mmol/L if <16 y.o.) >30 mmol/L in known DM |
| AST | >630 U/L | Potassium (K) ^{3,4} | <2.5 mmol/L >6.5 mmol/L |
| Ammonia | >100 µmol/L | Lactate | >4.0 mmol/L |
| Amylase | >600 U/L | Lithium | >1.5 mmol/L |
| Bicarbonate | <10 mmol/L | Magnesium | <0.4 mmol/L |
| Bilirubin (conjugated) | >25 µmol/L (Neonates only) | Sodium (Na) (Including Direct Sodium) | <120 mmol/L (<130 mmol/L if < 16 y.o.) >160 mmol/L |
| Calcium (adjusted) | <1.8 mmol/L >3.0 mmol/L | Paracetamol | >30 mg/L (4 hours post ingestion) |
| Calcium (Paeds) | <1.8 mmol/L >3.0 mmol/L | Phosphate | <0.35 mmol/L |
| Cortisol ¹ | <50 nmol/L | Phenytoin | >28 mg/L |
| Creatinine ² | >345 μ mol/L (\geq 200 μ mol/L if <16 y.o.) An increase of 1.5 times from the lowest value in the last 0-7 days. | Salicylate | >300 mg/L |
| CK (total) | ≥5000 U/L | Triglycerides | >20 mmol/L |
| CRP | 300 mg/L (primary care only) | Theophylline | >25 mg/L |
| Digoxin | >2.5 µg/L | Troponin (ED only) ⁵ | >34 ng/L (Male) >16 ng/L (Female) |
| Ethanol | 400 mg% (Please note mg% is the same as mg/dL) | Urea | >30 mmol/L |
| FT4 | <5, >50 pmol/L (Unless CRAD) | | (≥ 10 mmol/L if <16 y.o.) |

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| Haematology | / | | | |
|---|--|--|---|---|
| Test | Result | | Test | Result |
| WBC x 10 ⁹ /I | <1.00 | | HB g/dl | <7.0 |
| WBC x 10 ⁹ /I | >35 (GP), >50 |) (Ward) | HB g/dl | >17(F), >19(M) |
| PLT x 10 ⁹ /I | <50 | | $PLT \times 10^{9}/I$ | >800 (GP), >1000 (Ward) |
| Neutrophils | < 0.5 x 10 ⁹ /l | (0.5 - 1.0) | CD4 | CD4 <200 absolute count |
| neuci opinio | phoned next d | • | | (unexpected or 1 st time) |
| Kliehauer | Foetal bleed > | | Fibrinogen | |
| | | 12 mis | Factor Xa | <1.0 |
| APTT | > 100 secs | | Factor Xa | >1.0 IU/mL |
| D-Dimer | >35.2mg/L FE | U | DOACs | Rivoroxaban >419 ng/ml Apixaban >321 ng/ml |
| INR | >4.5 (>4.5 an | d < 5.0 and GP - | Next morning Oł | (all others to Sth doc) |
| Any significan baseline Hb is | | B level e.g.>2g/ | 'dl if baseline Hb | is = 8.0 g/dl and 3g/dl |
| Positive sickle | cell screens in p | atients with <u>pre-</u> | op indicated on f | orm |
| Positive HCGs | in hospitalised in | n-patients | | |
| Urgent Factor | | | | |
| 5 | emic Syndrome | | | |
| | ed Leukaemia's | | | |
| · · · · · · · · · · · · · · · · · · · | | | | |
| Positive Malari | | | | |
| | spot Screening te | est | | |
| Equivocal Prec | | | | |
| Microbiology | | | | |
| aspirat • New ZI | | - | F's and normally | sterile body fluids, e.g. joint |
| Culture | | | | |
| | e blood cultures | | | |
| | e CSF cultures | | | |
| | a cultured of porr | | a | |
| | | | y fluids, e.g. joint | |
| | RSA, VRE or othe | er multi drug resi | y fluids, e.g. joint istant organisms | |
| Gonoco | RSA, VRE or othe occi (except to S | er multi drug resi TI clinic) | | |
| GonocoNew M¹ | RSA, VRE or othe occi (except to S ⁻ ycobacterial cult | er multi drug resi TI clinic) ure positives | stant organisms | |
| Gonoco New M¹ Skin ar | RSA, VRE or othe occi (except to S ⁻ ycobacterial cult | er multi drug resi TI clinic) | stant organisms | |
| Gonoco New M Skin ar Enterics | RSA, VRE or othe occi (except to S ⁻ ycobacterial cultu nd soft tissue Gro | er multi drug resi TI clinic) ure positives oup A Streptococ | stant organisms ci | |
| Gonoco New M Skin ar Enterics New po | RSA, VRE or othe occi (except to S ycobacterial cultu nd soft tissue Gro ositive results: ba | er multi drug resi TI clinic) ure positives oup A Streptococ acterial, viral or p | stant organisms ci | |
| Gonoco New M Skin ar Enterics New po Infectious Di | RSA, VRE or othe occi (except to S ycobacterial cultu nd soft tissue Gro ositive results: ba seases Serolog | er multi drug resi TI clinic) ure positives oup A Streptococ acterial, viral or p | stant organisms ci parasitic | |
| Gonoco New M ¹ Skin ar Skin ar Enterics New po Infectious Di Laborator | RSA, VRE or othe occi (except to S ⁻ ycobacterial cultu nd soft tissue Gro ositive results: ba seases Serolog ry Test | er multi drug resi TI clinic) ure positives oup A Streptococ acterial, viral or p IY Result | istant organisms ci barasitic Category | Comment |
| Gonoco New M Skin ar Skin ar Enterics New po Infectious Di Laborato Toxoplasr | RSA, VRE or othe occi (except to S ⁻ ycobacterial cultu nd soft tissue Gro ositive results: ba seases Serolog ry Test na IgM | er multi drug resi FI clinic) ure positives oup A Streptococ acterial, viral or p IY Result Positive | istant organisms ci barasitic Category C | Comment Pregnant patient |
| Gonoco New M Skin ar Enterics New po Infectious Di Laborator Toxoplasr CMV I | RSA, VRE or othe occi (except to S ⁻ ycobacterial cultu nd soft tissue Gro ositive results: ba seases Serolog ry Test na IgM gM | er multi drug resi FI clinic) ure positives oup A Streptococ acterial, viral or p NY Result Positive Positive | istant organisms ci Darasitic Category C C | Comment Pregnant patient Pregnant patient |
| Gonoco New M Skin ar Enterics New po Infectious Di Laborator Toxoplasr CMV I Rubella | RSA, VRE or othe occi (except to S ⁻ ycobacterial cultu nd soft tissue Gro seitive results: ba seases Serolog ry Test na IgM gM IgM | er multi drug resi FI clinic) ure positives oup A Streptococ acterial, viral or p NY Result Positive Positive Positive Positive | istant organisms ci Darasitic Category C C C | Comment Pregnant patient Pregnant patient Pregnant patient Pregnant patient |
| Gonoco New M Skin ar Skin ar Enterics New po Infectious Di Laborato Toxoplasr CMV I Rubella Parvovirus | RSA, VRE or othe occi (except to S ⁻ ycobacterial cultu nd soft tissue Gro seases Serolog ry Test na IgM gM IgM B19 IgM | er multi drug resi TI clinic) ure positives oup A Streptococ acterial, viral or p DY Result Positive Positive Positive Positive Positive | istant organisms ci Darasitic Category C C C C C | Comment Pregnant patient Pregnant patient Pregnant patient Pregnant patient Pregnant patient |
| Gonoco New M Skin ar Skin ar Enterics New po Infectious Di Laborator Toxoplasr CMV I Rubella Parvovirus HIV Ag | RSA, VRE or othe occi (except to S ⁻ ycobacterial cultured soft tissue Groupsitive results: bases Serolog ry Test na IgM gM IgM IgM IgM IgM IgM IgM IgM IgM | er multi drug resi TI clinic) ure positives oup A Streptococo acterial, viral or p DY Result Positive Positive Positive Positive Positive Positive Positive | stant organisms ci Darasitic C C C C C C C C C | Comment Pregnant patient Pregnant patient Pregnant patient Pregnant patient New detection |
| Gonoco New M Skin an Skin an Enterics New po Infectious Di Laborato Toxoplasr CMV I Rubella Parvovirus HIV Ag HBs/ | RSA, VRE or othe occi (except to S ⁻ ycobacterial cultured soft tissue Gro positive results: bases serolog ry Test na IgM gM IgM B19 IgM /Ab | er multi drug resi FI clinic) ure positives oup A Streptococ acterial, viral or p Positive Positive Positive Positive Positive Positive Positive Positive Positive Positive | istant organisms ci Darasitic C C C C C C C C C C C C | Comment Pregnant patient Pregnant patient Pregnant patient Pregnant patient Pregnant patient New detection New detection |
| Gonoco New M Skin ar Skin ar Enterics New po Infectious Di Laborator Toxoplasr CMV I Rubella Parvovirus HIV Ag HBsA Anti-H | RSA, VRE or othe occi (except to S ⁻ ycobacterial cultu nd soft tissue Gro ositive results: ba seases Serolog ry Test na IgM gM IgM B19 IgM /Ab yg ICV | er multi drug resi FI clinic) ure positives oup A Streptococo acterial, viral or p IV Result Positive Positive Positive Positive Positive Positive Positive Positive Positive Positive Positive | stant organisms ci Darasitic C C C C C C C C C C C C C C C C C C C | Comment Pregnant patient Pregnant patient Pregnant patient Pregnant patient New detection New detection New detection New detection New detection |
| Gonoco New M Skin ar Skin ar Enterics New po Infectious Di Laborato Toxoplasr CMV I Rubella Parvovirus HIV Ag HBs/ | RSA, VRE or othe occi (except to S ⁻ ycobacterial cultu ad soft tissue Gro sitive results: ba seases Serolog ry Test na IgM gM IgM B19 IgM Ab Sg CV ntibody | er multi drug resi FI clinic) ure positives oup A Streptococ acterial, viral or p Positive Positive Positive Positive Positive Positive Positive Positive Positive Positive | stant organisms ci barasitic C C C C C C C C C C C C | Comment Pregnant patient Pregnant patient Pregnant patient Pregnant patient Pregnant patient New detection New detection |

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| All positive ten | nporal artery bi | opsies (Neurop | athology) | |
|----------------------|---------------------------------|-------------------------------|----------------------------------|--------|
| • | at the discretior | n of the reporti | ng Pathologist | |
| POCT Blood Gas Sa | mnles | | | |
| | Inples | | | |
| Test | Critical Results Arterial | Critical Results Venous | Critical Results Capillary | Units |
| рН | <7.2 >7.6 | <7.2 >7.6 | <7.2 >7.6 | |
| pCO2 | <2.6 >9.3 | | | kPa |
| PO2 | <6 | | | kPa |
| Na+ | <120 >160 | <120 >160 | <120 >160 | mmol/L |
| K+ | <2.8 >6.2 | <2.8 >6.2 | <2.8 >6.2 | mmol/L |
| iCa ⁺⁺ | <0.5 >1.58 | <0.5 >1.58 | <0.5 >1.58 | mmol/L |
| Glu | <2.2 >24.9 | <2.2 >24.9 | <2.2 >24.9 | mmol/L |
| Lac | >2 | >2 | >2 | mmol/L |
| Bicarb | <10 >40 | <10 >40 | <10 >40 | mmol/L |
| Hb | <7.0 | <7.0 | <7.0 | g/dL |

Note: It is the responsibility of the POCT Operator to act immediately on any critical results and/or inform the appropriate clinician.

For unexpected significant results that are not consistent with the clinical picture, where the results require clinical intervention, or where the Operator is not reassured by the POCT result, a repeat sample should be run or a sample should be sent to the lab for confirmation. Advice on critical results may be obtained from Duty Biochemist at Ext: 22870

POCT Creatinine

Pathology

Frozen section reports

| Test | Units | Critical Result |
|-----------------|---------------------------|-----------------|
| POCT Creatinine | µmol/L | ≥ 300 |
| POCT eGFR | ml/min/1.73m ² | ≤ 30 |

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8.3 Printed Reports

- 1. Reports are printed with reference ranges and/or suitable comments wherever appropriate, to aid interpretation of results. Reports will only be given to the submitter. Private individuals will not receive reports.
- 2. Please note the printed authorised report (or an amended subsequent report) issued by Laboratory Medicine is the medico-legal document within the patient record.

Posted

Posted

Collected daily

Collected daily

Collected daily Collected daily

Collected daily

South Infirmary porter collects reports

periodically throughout the day.

- 3. Printed reports are delivered by the portering staff to CUH wards.
- 4. External hospitals are printed and issued as follows:
 - Bon Secours Hospital
 - Mallow General Hospital
 - Mercy University Hospital
 - St. Mary's Campus
 - St. Finbarr's Hospital
 - South Infirmary Hospital
 - University Hospital Waterford
 - University Hospital Kerry
 - University of Limerick Hospital Posted (to UHL) and collected daily Groups
- 5. Results for General Practitioners are printed and posted daily.
- 6. Emergency, critical and urgent positive reports are phoned directly to the wards and/or ordering clinician.
- 7. Results are electronically sent to some General Practitioners who have registered with GP messaging for more information (see below).

Pathology: Responsibility for receipt of report lies with the requesting clinican

GP Messaging - Electronic delivery of laboratory reports to the GP practice

Laboratory Medicine facilitates the issue of electronic reports to GP practices. This is facilitated using Healthlink messaging. Healthlink is the national standard for messaging between Hospitals and General Practitioners. Laboratory Results can be either viewed directly on Healthlink or integrated into Practice Management Software

Electronic laboratory facilitated reports are issued for Biochemistry, Haematology and Microbiology only.

Electronic reports are issued from Laboratory Medicine in real time. To avoid reports going to the wrong GP practice it is best to clearly print your laboratory GP location code on any test request forms being sent to Laboratory Medicine. Some practices have their laboratory GP location code incorporated into their practice stamp or on their computer generated address labels.

If you do not know your laboratory GP location code contact Laboratory Medicine at CUH on 021-4921309.

For those who are using Healthlink messaging, it is vital to regularly check reports imported into your PMS with either printed or from the Healthlink website.

This is to ensure that results, reference ranges, demographics etc are being transferred correctly from Laboratory Medicine to your PMS.

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If you have any problems with any aspect of GP messaging your first point of contact is your GPPMS software provider or the Healthlink (01) 828 7115 or email support.healthlink@healthmail.ie

8.4 Electronic Reports within CUH/CUMH

The Laboratory Information System (iLaboratory) has HL7 interfaces to the following Clinical Information Management Systems thus allowing the transmission of laboratory results immediately upon authorisation in the lab.

> DAWN

DAWN AC (DINR) Anticoagulation software is a medical application designed for managing large anticoagulation clinics. It determines the patients wafarin dosage based on their INR result.

> eMed (Renal)

eMED*Renal* is a national clinical and patient management software system designed for renal patients.

> iCIP

The IntelliVue Clinical Information Portfolio (iCIP) is a software suite designed to centralise patient data so clinicians have access at the patient's bedside in specific locations e.g. ICU, to the information they need to make clinical decisions. The patients MRN must contain a '**C**' prefix and they must be admitted to ICU in order for reports to download to this system.

≻ iCM

The iSOFT.Clinical Manager (iCM) application is an electronic health record for patients. It has many features to help organise patient information. These include placing electronic orders for tests and viewing their results. The patients MRN must contain a '**C**' prefix in order for reports to download to this system.

> Maternity System

The MN-CMS is an Electronic Health Record (EHR) for all women and babies who access the Maternity Services in Ireland. This system provides accurate and up to date clinical information to all those involved in the care of mothers and babies in our maternity units, allowing for their information to be shared with the relevant health care providers that need to access the data for the provision of care.

9 INFORMATION TECHNOLOGY

9.1 Laboratory Medicine Results Access Policy and Confidentiality Guidelines

Laboratory medicine results are stored on a Laboratory Information System [LIS]; the system is currently i.Laboratory. All hospital medical, nursing and relevant clerical staff are granted access to the full range of patient data held, subject to the terms and conditions as outlined in this policy. Non hospital HSE contracted medical, nursing and relevant clerical staff are also granted access – either to data restricted and relevant to patients in their practice area e.g. Community hospitals and GPs; or to the entire range of patient data, e.g. public health staff.

The applicant will ensure that there is tight control on access to patient pathology results via Lab Enquire in their ward, office *etc*.

<u>Please note: Histopathology results are only for look up/internal purposes and are not official Histopathology results and should not be used in any correspondence.</u>

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The applicant is responsible for the proper use of the facility.

- Usernames and Passwords must not be shared.
- Any patient specific information gained through work or on receiving reports from Laboratory Medicine is strictly confidential and must not be relayed or discussed with any third party unless they are specifically authorized to receive the information.
- Never examine any material or report that is not pertinent to your work.
- Only a doctor may authorise Laboratory Medicine information being passed to a third party. The points outlined in the Medical Council Guidelines section 31.03 should be borne in mind by any doctor passing information to a third party.
- All patient identifiable information must be held securely and locked away when not personally attended; such data must never be stored on removable storage devices (USB memory key, floppy disk, CD/DVD).
- If patient identifiable information is entered on computer, that computer should be password protected
- Never transmit confidential named patient data by email with the exception of @hse.ie accounts or to the following addresses: <u>Voluntary Hospitals:</u>
 - AMNCH, Tallaght @amnch.ie
 - Beaumont Hospital @beaumont.ie
 - Cappagh National Orthopaedic Hospital @cappagh.ie
 - Coombe Women & Infants University Hospital @coombe.ie
 - Mater Public, Dublin @mater.ie
 - Marymount University Hospital and Hospice, Cork @marymount.ie
 - Mercy University Hospital, Cork @muh.ie
 - National Maternity Hospital, Holles Street, @nmh.ie
 - National Rehabilitation Hospital, @nrh.ie
 - Our Lady's Hospice, Harold's Cross, Dublin @olh.ie
 - Our Lady's Children's Hospital, Crumlin @olchc.ie and @olhsc.ie
 - Rotunda Maternity Hospital, Dublin @rotunda.ie
 - South Infirmary Victoria University Hospital, Cork @sivuh.ie
 - St. Francis Hospice, Dublin @sfh.ie
 - St. James's Hospital, Dublin @stjames.ie
 - St. John's Hospital, Limerick @stjohnshospital.ie
 - St. Luke's Hospital, Rathgar, Dublin @slh.ie
 - St. Vincent's Hospitals Group @st---vincents.ie, @svuh.ie, @stmichaels.ie, @svhg.ie
 - Temple Street Children's University Hospital @cuh.ie

Private Hospitals And Clinics

- Aut Even Hospital, Kilkenny @auteven.ie
- Bon Secours Hospital, Tralee @bonsecours.ie
- St. Vincent's Private Hospital, Dublin @svph.ie
- Whitfield Clinic, Waterford @whitfieldclinic.ie

Agencies:

- Central Remedial Clinic (Dublin, Limerick & Waterford) @crc.ie
- Department of Health @health.gov.ie
- Health Products Regulatory Authority @hpra.ie
- Healthlink, National Messaging Broker @healthlink.ie, @healthlink.doh.ie
- SouthDoc @southdoc.ie
- Caredoc, caredoc@healthmail.ie
- NEDOC North East Doctor On Call nedoc@healthmail.ie
- National Cancer Registry Ireland <u>ncri@healthmail.ie</u>

If you have a query about any other location enquire at <u>https://www.healthmail.ie/support.cfm</u>

• All printed or written records with personal data should be shredded as soon as they are no longer needed.

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• Each employee is personally responsible for the security and confidentiality of all types of paper and electronic information which they come in contact with during the course of their work.

Each member of staff with access to Laboratory Medicine results **MUST** adhere to the following HSE policy:

Information Security Policy and Information Technology Acceptable Usage Policy <u>http://hsenet.hse.ie/OoCIO/Service_Management/PoliciesProcedures/Policies/HSE_I_T_Security_Policy.pdf</u>

9.2 Confidentiality Undertaking for Staff having Access to, or Receiving, Laboratory Results

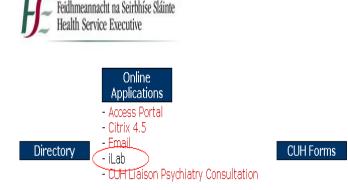
I understand that, in the course of my work, I may come into contact with, or have access to, confidential information relating either to individual patients, members of staff or to general public health issues. I understand that misuse of this information, especially its disclosure to people or agencies that are not specifically authorised to receive it would constitute a breach of confidentiality. I also understand that the use and securing of personal information is subject to the provisions of the Data Protection Act and that unauthorized disclosure of personal information is an offence under the act.

I confirm that I have read the above Laboratory Medicine guidelines on confidentiality and that I agree to comply with them as formally undertaken by signing the On-Line Laboratory Medicine Results and Confidentially Guidelines form.

9.3 Instructions i.Laboratory/Web Browser

Please note the icon for this application can be found on Staff Directory under Online applications, or by clicking on the following link

http://10.54.128.107/apex/mgwms32.dll?MGWLPN=APEX&APP=PCOMB&APPDIR= /APEX



- 1. Enter the Username and Password (if you have a problem logging on check if pop blocker is on).
- 2. Where prompted Patient Number enter C for Cork PIMS registered patients OR T for Tralee PIMS registered patients followed by the patients Medical Record Number
- 3. Under surname enter the first three letters of the patient's surname.

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4. Then click the grey "NUMBER SEARCH" button on the right hand side of the screen.

Note: If an MRN/RID is unavailable enter the patients Surname, Forename and DOB and click Search. Patients matching your search information will be returned select the patient required by clicking on the patient MRN/RID in the PATIENT RECORD NUMBER column

- 5. On selecting a patient the user can select specific discipline\specimen date or continue for most recent result.
- 6. All the lab results on the patient selected will be displayed. The most recently authorised report from the lab will appear at the top of the list. Select the specimen results you are looking for by clicking once on the appropriate date and time box in the Specimen Dare & Time column.
- 7. The results on the specimen selected will be displayed. Use the scroll bar on the right hand side of the screen to look for tests not displayed on the first screen. High or low results will be highlighted in a different colored box. Usually light blue for just outside the normal range and dark pink for well outside the range. Single or double arrows pointing up or down will also be displayed for results outside the reference range.
- 8. To review another specimen on that patient click once the <<Select Order Specimen button.
- 9. When Finished click the LOG-OFF button.
- 10. The i.Laboratory report font size can be enlarged on your pc screen hold Ctrl on the keyboard and rolling the mouse wheel up alternatively select Ctrl and +

How To Change the Lab Enquiry password (automatic account deactivation after three months if not updated

- 1. On iLaboratory log in screen click Change password button.
- 2. Enter your current username, current password and new password where prompted.

Note: The new password cannot be the same as the last and must contain at least five letters and one number.

- 3. Then click the Ok button. This new password takes immediate effect.
- 4. The password will be valid for three months and you will get a warning on screen every time you log on starting 20 days from the expiry date.
- 5. If you have any problems changing your password contact the Laboratory Information Systems Helpdesk by e-mail at <u>CUHIT.Pathology@hse.ie</u> on by phone on 021-4920150

9.4 iClinical Manager (iCM)

i.Clinical Manager (iCM) is the electronic patient record used in CUH. It provides order comms for Biochemistry, Auto Immune Serology, Haematology or Microbiology.

NB for full details on use of iCM please refer to the ICT User Manual

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All iCM user data including how to apply for an account, logging onto iCM and searching for patient data can be found on Staff Directory under Guidelines \rightarrow iCM Users Guidelines or by clicking on the following link:

http://100.24.9.212/Menu ApplicationForms/UserAccountRequestFormDoctors/Us erGuides.asp

9.4.1 Logging on to iCM

- 1. Staff directory → Citrix→ National StorefrontPortal enter your windows password → Hosted apps → ICM-SSWHG
- 2. This opens the iCM Log-On Screen Log into iCM please note the Username format is different from Citrix as it does not contain a dot between firstname and surname.e.g. If you log into Citrix as test.frank then your ICM log in will be testfrank.

9.4.2 Selecting a Patient

- 3. On logging into ICM the Patient List displays a list of current patients in a specified area.
- 4. The List Displayed is shown in the Current List dropdown box which can be changed by selecting a different dropdown option. To select a patient click on chosen patient so their details will display on the header.

9.4.3 Ordering of Laboratory Specimens on ICM

- 1. Obtain specimen from patient.
- 2. Select patient from appropriate list on ICM.
- 3. Go to Orders Tab.
- 4. Click Enter Order Icon on header or Enter Order button to open Order Browse.
- 5. Use Relevant Order Set or predictive text option at the 'Type to enter' field to find appropriate investigation and
- 6. Select or deselect components of Order Set as required.
- 7. Ensure Order is submitted on behalf of Consultant.
- 8. Add order.
- 9. To prioritise samples select URGENT REQUEST as the Collection Time
- 10. Amend clinical details (inadequate details can cause laboratory process delays)
- 11. Click OK.
- 12. Submit Orders Pending.

9.4.4 Collection of Specimen

- 1. On Orders Screen Add Specimen and select performing Department
- 2. Tick boxes to confirm investigations.
- 3. Amend number of labels if multiples required e.g. Blood Cultures
- 4. Click OK.
- 5. Ensure that labels printed match the details of patient identified for phlebotomy.
- 6. Ensure labels are affixed to correct bottles. Do not cover specimen blood volume or container `fill to' marks.
- 7. Specimen Type on label should match Specimen Type on Bottle.
- 8. Bag Specimen

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9.4.5 Results Viewing

- 1. Results are available in iCM once all parts of the request profile are authorised by Lab
- 2. Click on the Results tab for a selected patient
- 3. Results outside of normal parameters are flagged with red arrows.

NB As Microbiology results and Positive/Negative text based abnormal results are not flagged

A \blacksquare button in a result field indicates that there is an expanded result –right click to view entire comment

| 🔉 TESTLAB, AUDIT - ISOFT CI | | <u></u> |
|----------------------------------|--|-------------------------------------|
| File Edit View GoTo Actions Pr | references Tools Help | |
| 18 🗃 🖣 👬 🕨 🖿 | S & P P 🥄 \$ 2 X EI B 🗈 🏈 🚧 🏵 🗤 🖉 🕌 🗰 🗱 🖂 ? 🌮 | 3 4 🕖 🕖 |
| TESTLAB, AUDIT Administrative | 2049740 / P2049740 | 62y Male |
| Patient List Orders Results Docu | ments Observations Patient Info Summary | |
| Chart | Laboratory results - Performed since 12-Mar-2012 | 2 |
| All Available 💌 | Anti-Thrombin 3 101 | [80-120 %] |
| Since | Protein C 102 Protein S 65 | [70-120 %] |
| C Received Performed | Anti-Cardiolipin IgG 2.0 | [65-101 %] [0-10 GPL/mL] |
| 12-Mar-2012 - | Anti-Cardiolipin IgM 3.0 | [0-7 MPL/mL] |
| Six months ago 🔹 | 05-Sep-2012 13:25 INR, APTT & PT | 1 or more Final Results Received |
| Retain for next patient | APTT 32 | |
| Result Selection | INR 1.0 | [0.9-1.1] |
| Display Category Headers | | [9.7-11.3 sec] |
| Abnormal 🗖 Show Pending | PT /APTT:These normal ranges do NOT apply to patients on O5-Sep-2012 13:26 FBC | 1 or more Final Results |
| New Results | | Received |
| | Expanded Result | × 12/L] |
| Display Format | | |
| Report by Order Graph Summary | PT/APTT: These normal ranges do NOT apply to patients on | |
| Trend View | I anticoagulants. Therapeutic ranges are decided by | |
| | dinicians. | |
| | | r9/L1 |
| | | 9/1 |
| | | [^9/L] |
| | | 9/L] |
| | Bi Ec | 9/L] |
| | 05-5 | e Final Results |
| | | d |
| | St | L) |
| | | T |
| | Sc OK | [L] |
| | 05-Sep-2012 13:44 Ferritin. | 1 or more Final Results Received |
| | Ferritin. 1495 | ↑ [17-320 ng/mL] |
| | 10-Sep-2012 10:38 Dust Mite | 1 or more Final Results |

A \blacksquare in a result filed indicates that a result has been modified - right click to view previous result

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|----------------------------|---|---------------------------------|-------------------------|------------------------------------|-------------------------------|--------------|----|
| Administrative | | 2049740/R2 | 2049740 | | | 62y Mal | le |
| t List Orders Results Docu | ments Observations Pa | ient Info Summary | | | | | |
| | V | | Laboratory res | ults - Performed since 05-Sep-2012 | | | |
| vailable 🔻 | Dog Dander | | | 0.50 | ♦ [0-0.35 kU/L] | | |
| | 10-Sep-2012 10:38 | Hens Egg White | | | 1 or more Fi | inal Results | |
| eceived Performed | Hens Egg White | | | 12.00 | Received 10-0.35 kU/L1 | | |
| ep-2012 🕂 💌 | 10-Sep-2012 10:38 | Sesame Seed | | 12.00 | 1 or more Fi | inal Results | |
| week ago | Sesame Seed | | | 2.70 | Received 10-0.35 kU/L1 | | |
| etain for next patient | 10-Sep-2012 10:38 | Pea | | 2.70 | 1 or more Fi | inal Results | |
| It Selection | | | | | Received | | |
| - | Pea 10-Sep-2012 10:38 | Peanut | | 5.80 | [0-0.35 kU/L] 1 or more Fi | inal Results | |
| isplay Category Headers | | | | | Received | | |
| New Results | Peanut 10-Sep-2012 11:06 | Brazil Nut | | 0.40 | [0-0.35 kU/L] 1 or more Fi | inal Results | |
| new nesults | | Diala Hak | | | Received | | |
| ay Format | Brazi Nut | pdated Results by Received Date | | 0.05 | X | al Results | |
| ort by Order 🛛 🗖 Graph | 10-3ep-2012 11:00 | | | | | | |
| mary nd View | Almond | Order: Thyroid Function Tests. | 11-Sep-2012 09:00 Corre | ected Results | | | |
| | 10-Sep-2012 11:06 | Results Received | | | | al Results | |
| | Coconut | 12-Sep-2012 12:46 | | | | | |
| | 10-Sep-2012 14:29 | Free-T4 | 25. | 3 🐈 [12-22 pmol/L] | | al Results | |
| | Hepatitis B sulface | 12-Sep-2012 12:45 Free-T4 | 15. | 2 [12-22 pmol/L] | | | - |
| | Hepatitis C an ibodi 10-Sep-2012 14:29 | Fiee-14 | 15. | : [12-22 pmovu] | | al Results | |
| | | | | | | ui ricsuits | |
| | HIV 1 and 2 10-Sep-2012 14:29 | | | | | al Results | |
| | 10-Sep-2012 14:23 | | | | | ainesuits | |
| | Mumps IgG Antiboc | | | | | | |
| | Rubella IgG antiboo Measles IgG Antibo | | | | | L | |
| | 10-Sep-2012 14:29 | | | | | sults | |
| | Measles IgM Antibo | | | | | | |
| | 10-Sep-2012 14:29 | I | | | | sults | |
| | Mumps IgM Antiboc | Order Dietaïs Iter | m Info | Close Help | | | |
| | 10-Sep-2012 14:29 | | | | _ | al Results | |
| | Rubella IgM antibody | | negative | | Heceived | | |
| | 11-Sep-2012 09:00 | Thyroid Function Tests | neyauve | | Corrected R | lesults | |
| | Free-T4 | | 1) | 25.3 | [12-22 pmol/L] | | |
| | TSH | | / | 2.30 | [0.4-3.8 mIU/L] | | |

This view can be modified to select a specified date range or performing laboratory or test by selectively choosing options on the left hand sidebar

9.4.6 Contingency

Submitting Orders

Users should revert to manual contingency i.e. use paper forms for any requests submitted during downtimes (either iCM or Laboratory Information System {LIS}) Result Viewing

If iCM is down results will be available on iLaboratory

If LIS is down only results authorised prior to downtime will be available on iCM. Laboratories can be contacted for URGENT results.

Remember

Patient identity must be confirmed before phlebotomy

Samples must be labelled at all times

For training, fault logging, etc please contact the ICT Helpdesk on 28000 or email <u>cuhit.helpdesk@hse.ie</u>

9.5 Maternal & Newborn Clinical Management System (MN-CMS)

The MN-CMS Project is the design and implementation of an Electronic Health Record (EHR) for all women and babies in maternity services in Ireland. Cerner are the EHR provider chosen to deliver the system. The solution is called Cerner Millennium® and has been in use in CUMH since 2016. It provides order comms for Biochemistry, Auto Immune Serology, Haematology or Microbiology.

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NB for full details on use of MN-CMS please refer to the MN-CMS Familiarisation Recordings available on CUH Staff Directory under Guidelines → Maternal Newborn Clinical Management System or by clicking the following link: <u>http://10.54.129.212/Menu_PolicyProcedure/MNCMS.asp</u>

All MN-CMS user data including how to apply for an account, logging onto MN-CMS and searching for patient data can be found on Staff Directory under Guidelines→ Maternal Newborn Clinical Management System or by clicking on the following link: http://10.54.129.212/Menu PolicyProcedure/PDFs/MNCMS/MN-CMS%20Information%20Governance%20and%20Security%20Leaflet August%20 2016.pdf

9.5.1 Logging on to MN-CMS

- 1. Staff directory → Citrix→ National StorefrontPortal enter your windows password → Hosted apps → Powerchart
- 2. This opens the Cerner Millenium Log-On Screen. Log into MN-CMS please note the Username format is different from Citrix as it does not contain a dot between firstname and surname.e.g. If you log into Citrix as test.frank then your MN-CMS log in will be testfrank.

9.5.2 Selecting a Patient

- 1. On logging into MN-CMS the Maternity Whiteboard displays a list of current patients in a specified area.
- 2. To select a patient click on chosen patient so their details will display on the header and their chart opens on the default screen of **Maternity View**.
- 3. Alternatively, search for the patient using the MRN or surname using the appropriate dropdown in top right hand corner search field.

9.5.3 Ordering of Laboratory Specimens on MN-CMS

Ordering laboratory tests on a patient can be carried out by one of two ways: (a) USING QUICK REQUESTS

- 13. Obtain specimen from patient.
- 14. On the Maternity View screen, select the Quick Requests option, which opens a new screen.
- 15. Select the required order under Lab Order Selection
- 16. Multiple orders can be selected by clicking on them which highlights the required orders.
- 17. These orders then have to be signed to place the order successfully, select the green Orders for Signature option
- 18. Click on the Sign option
- 19. An Ordering Clinician window pops up, enter Clinician Surname and search, then select the appropriate option.
- 20. The Order Date/Time and the Communication type default.
- 21. Click OK
- 22. The selected orders appear in a new window. Before you can Sign the order, the required missing details need to be entered.
- 23. Click on the Missing Required Details on the bottom left hand side of the window, to display any further required information to be entered.

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- 24. Enter the details required in the fields seen. Mandatory fields appear in yellow and occasionally in white. An order cannot be signed until all the mandatory fields have been completed
- 25. Click on the Sign option below, which then closes this window.

| t Patient Ne rgy: Hypochlo | w, Mn-Cms prite, Latex, I | s Two Penicillin -class of | Age:27 years DOB:19/Jan/90 | Sex:Female MRN:6432741 EDD:06/09/2017 | Loc:CUMH-Emergency Room Outpatient(Public) [23/Nov/2016 08:49] Consultant: |
|-------------------------------|------------------------------|--|-------------------------------|---|--|
| dd 🖨 Docum، | nent Medicatio | n by Hx À Check Inte | eractions 🛄 Extern | nal Rx History - No Check - | Reconciliation Status Meds History Admission |
| ers Medication | n List Docum | ent in Plan | | | |
| Orders for Signat | ure | | | | |
| & \$ | | Order Name | Status Start | Details | |
| ⊿ CUMH-Em | | n Fin#:0111933640 A | dmit: 23/Nov/2016 | 5 08:49 GMT | |
| ⊿ Laboratory | | | | | <u> </u> |
| | ~ي _ | | | | e, Coll date/time: 19/Apr/17 08:25 WEST |
| | _ <u>ب</u> | Liver function screen, | | | e, Coll date/time: 19/Apr/17 08:25 WEST |
| | _خ 🗆 | Full blood count | Order 19/A | pr/2017 08:25 Priority: Routine | e, Coll date/time: 19/Apr/17 08:25 WEST |
| | | | | | |
| ▼ Details for | | ers | | | |
| | | | | | |
| Petails | Order Comm | ients 🕅 🔯 Diagnoses 🗋 | quired details | | |
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Using the Orders Tab

1. From the options listed on the left hand side of the patients' chart, click on the + Add on the Orders tab or alternatively, click the Orders tab and then click the + Add option on the orders window which opens

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| | | | | 4 | - in meanean | Record Request 😋 Result Copy 🛼 Rel | | | |
|--|---|-----------------|-----------------|---|-------------------|--|--|--|--------------------|
| est Patient est Patient New, llergy: Hypochlorit | × Mn-Cms Two æ, Latex, Penicillin ⊣ | class of antib | iotic- | Age:27 years DOB:19/Jan/90 | | Sex:Female MRN:6432741 EDD:06/09/2017 | Loc:CUMH-Emergency Roo Outpatient(Public) [23/Nov/2010 Consultant: | | • |
| /enu | | | Orders | | | 200.00/09/2017 | consultant. | D Full screen Print | € 2 minutes a |
| aternity View | | _ | | | | | | | € 2 minutes e |
| aternity View eonate View | | dd 🖓 Docum | | ation by Hx 🚴 Check Interactions 🛅 | External Rx H | istory • No Check • | | Reconciliation Status Meds History Admissi | on Outpatier |
| rders | + Add | Document | in Plan | | | | | | |
| nical Notes cumentation | + Add | Displayed: Al A | ctive Orders | Inactive Orders Since 23/Nov/16 (All Order | rs (All Statuses) | | | S | how More Orders |
| tivities | | a 5 | 2 | Order Name | Status | Details | | | |
| | | ⊿ Laborator | , | | | | | | |
| g Chart Summary | | | | Acanthamoeba PCR, specimen | Discontin. | Priority: Routine, Spec type: Eye swab, | Coll date/time: 11/Apr/17 16:14:00 WEST | | |
| | | | | Acanthamoeba PCR, specimen | | | Coll date/time: 06/Apr/17 09:53:00 WEST | | |
| ent Information | | | II G | Acanthamoeba PCR, specimen | | Priority: Routine, Spec type: Eye swab, | | | |
| | | | v | | | Priority: Routine, Coll date/time: 08/M | | | |
| m Browser | | | ~ | Albumin level, fluid | | Priority: Routine, Coll date/time: 23/N | | | |
| | | | ~ | Alk phos level, fluid Amvlase level, fluid | | Priority: Routine, Coll date/time: 23/No Priority: Routine, Coll date/time: 23/No | | | |
| ern Report | | | iii R | | | Priority: Routine, Coll date/time: 25/14 | | | |
| | | | × * | Anti-SM level. Blood | | Priority: Routine, Coll date/time: 30/No | | | |
| rningLIVE | | | ~ | Anti-SSA level blood (Anti-Ro level, I | b Ordered (. | Priority: Routine, Coll date/time: 30/N | ov/16 14:49:00 GMT | | |
| | | | | Atypical pneumonia serology, blood | InProcess | Priority: Routine, Spec Type: Serum, Co | oll date/time: 16/Dec/16 17:47:00 GMT | | |
| | | | | Atypical pneumonia serology, blood | I InProcess | Priority: Routine, Spec Type: Serum, Co | oll date/time: 16/Dec/16 17:45:00 GMT | | |
| | | | | Atypical pneumonia serology, blood | | Priority: Routine, Spec Type: Serum, Co | | | |
| | | | | Atypical pneumonia serology, blood | | Priority: Routine, Spec Type: Serum, Co | | | |
| | | | Q ✓ <u>≯</u> | Atypical pneumonia serology, blood | | Priority: Routine, Spec Type: Serum, Co | | | |
| | | | ≝.2 | Baby blood group and baby DAT, blood CMH | Ordered | mary, 2347889, 06/Dec/10, Priority: Ro PLEASE PRINT A REQUISTION FOR TH | utine, Coll date/time: 06/Dec/16 10:33:00 GM Is OPDER | 1 | |
| | | | | Bartonella screen, blood | | Priority: Routine, Coll date/time: 16/De | | | |
| | | | | Biochemistry add-on request | | Test: CRP. Bleep/tel no.: 111. Priority: F | | | |
| | | | | Blood culture MCS | | Print 2 labels if more than one bottle r | umen, Coll date/time: 12/Apr/17 14:43:00 WE equired. Transfer to the lab ASAP, delays can | result in false negative results. | |
| | | | | Blood culture MCS | Complete | | men, Coll date/time: 30/Mar/17 14:20:00 WE equired. Transfer to the lab ASAP, delays can | | |
| | | Tetails | | | | | | | |
| | | Dx Table | Orders F | or Cosignature | | | | On | ders For Signature |

2. A new window opens, to narrow down the search area, firstly use the drop down option for the Search within field and select Laboratory, then enter the required order in the Search field

| est Patient New, Mn-Cms Two Iergy: Hypochlorite, Latex, Penicillin -class of an | Age:27 years DOB:19/Jan/90 Sex:Female MRN:6432741 EDD:06/09/2017 | Loc:CUMH-Emergency Room Outpatient(Public) [23/Nov/2016 08: Consultant: | :49] |
|--|---|---|------------------|
| Diagnoses & Problems Diagnosis (Problem) being Addressed this Visit | Search: urea Search: urea Sulphonylurea level, urine Urea and electrolytes, blood Urea level, blood Urea level, fluid Urea output, 24hr urine | ed Options • Type: • Outpatient Orde Search within: • Laboratory | из & Rx – – – |
| | • | | 432741 Done |

- 3. Find the required order and click on it
- 4. An Ordering Clinician window pops up, enter Clinician Surname and search, n select the appropriate option.
- 5. The Order Date/Time and the Communication type default Click OK
- 6. Click Done or X out of the window if all required orders are already placed
- 7. Fill in the required details for the order, the mandatory fields in this case appear in yellow. Enter the information and click Sign

9.5.4 Collection of Specimen

Any orders made can be collected using the Specimen Collection option from the tabs across the top of the patients' chart.

1. Click the Specimen Collection tab

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| | | inks Notifications Navigation | | | | | | |
|-------------------|---|----------------------------------|--------------------------|---------------------------------------|--|------------------------------|--|------------------------------------|
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| est Patient | | | | | | | | t |
| | v, Mn-Cms Two ite, Latex, Penicillir | n -clais d'antibiotic- | Age:27 yea DOB:19/Ja | | Sex:Female MRN:6432741 EDD:06/09/201 | Out | :CUMH-Emergency Room patient(Public) [23/Nov/2016 08:49] nsultant: | |
| Aenu | 9 < | * A Maternity View | | | | | | 🗇 Full screen 🛛 👼 Print 🕹 0 minut |
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| ocumentation | + Add | Pregnancy Overview | | | | | | ∂ =- |
| ctivities | | | | | | | Cancel Pregnancy Cl | ose Pregnancy Modify Pregnancy |
| rug Chart Summary | | Current Pregnancy | Contact Info Demo | ographics | | | | |
| esults Review | | | | | | | | |
| tient Information | | E | DD 06/09/17 (Authorit | tative) | Current Weight | 56kg | Blood Type | O Rh D Negative |
| | | | GA 20 Weeks, 0 Days | | Pre-Preg Weight | | | |
| | | | ity G14,P10(9,1,2,1 | 10) | Height | | | |
| | | | ses No, Singleton | | BMI | 20.57kg/m2 | | |
| iscern Report | | Feeding P | lan | | | | | |
| earningLIVE | > | New Antenatal Inter | action (4) 🗸 | Result | | Author | Date/Time | Selected visit 2 =- |
| | | ⊿ Results (1) | | | | | | |
| | | Reason for Visit, Unplanned | I | Headache, Blurre pain, Reduced fel | d vision, Hypertension, Epigastri al movement | c Barry, Ciara Mary 142467 M | I/Std 11/01/17 1 | 1:51 |
| | | ⊿ Forms (3) | | | | | | |
| | | Booking Assessment | | Auth (Verified) | | ORegan, Monica 003416 CM | | 9:19 |
| | | Unplanned Maternal Admiss | sion/Assessment | Auth (Verified) | | Barry, Ciara Mary 142467 M | | |
| | | Booking Assessment A | | Modified | | ORegan, Monica 003416 CM | 4M2 23/11/16 0 | 3:14 |
| | | Antenatal Visits | | | | | | |
| | | | | | | | | OD EILEENKEOHAN 19 April 2017 16:2 |

- 2. Scan the patient's barcode
- 3. A window opens showing all orders made but pending collection.
- 4. For each of the orders, hover the mouse to the far right hand side and left click, then click on Collected from the options
- 5. At this stage, specimen barcodes can be printed for the orders.
- 6. Click on the Print option, followed by Print Label. The name of the printer you are printing the barcodes to needs to be known.
- After clicking Collected for an order, the coloured box (on the left hand side of this window), – indicating the colour of the sample tube required for that particular test, changes to a tick mark – indicating the order has been successfully collected.
- 8. Once all orders have been signed, the window updates with the message Patient has no specimen orders for collection. Click the Close option

9.5.5 Results Viewing

- 1. Results are available in MN-CMS once all parts of the request profile are authorised by Lab
- 2. Click on the **Results Review** option on the left hand side of the patients' chart.
- 3. Laboratory can be selected from the tab on this window to show only the relevant information from a laboratory perspective.
- 4. The results displaying are those within the timeframe shown across the top of this window. The arrows to the far left and far right of this window can be used to change the timeframe of viewable results.
- 5. Double click on a result to view additional information, such as the Laboratory Accession Number under the Result tab; the Source Type under the Specimen tab; specimen comments under the Comments tab; and an audit trail under the Action List tab.

9.5.6 Contingency

Submitting Orders:

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Users should revert to manual contingency i.e. use paper forms for any requests submitted during downtimes (either MN-CMS or Laboratory Information System {LIS})

Result Viewing:

If MN-CMS is down results will be available on iLaboratory

If LIS is down only results authorised prior to downtime will be available on MN-CMS.

Laboratories can be contacted for URGENT results.

Remember

Patient identity must be confirmed before phlebotomy Samples must be labelled at all times

9.6 Instructions for using the Blood Collection System Through Lab Enquiry

Please note that the 'yellow' blood collection slip can ONLY be generated through the 'Lab Enquiry' Icon. Web Browser CANNOT be used.

If the Lab Enquiry icon is not available, Please contact the Blood Transfusion Department at 22537

Double click on Lab Enquiry icon for results

Click once ****** the "Yellow Telephone" icon **Part 1** from toolbar

- Enter Username: Press Return.
- Enter Password and press Return.
- From Ward Enquiry Menu Screen:
- Enter Option 1
- Press Enter.
- From Ward Enquiry Screen:
- At the Patient Number prompt type C for Cork PIMS registered patients followed by the patients Medical Record Number.
- Press Enter.
- If asked Type first three letters of patient's surname and press Enter.
- Go to the latest Haematology Result. This allows you to check the Haemoglobin result prior to transfusion, if applicable.
- Select the appropriate button for the product required from the upper tool bar (i.e. 'Collect BLOOD' to collect a unit of red cells or 'Col. PLATELETS' to collect a unit of platelets) and click once.
- When finished search click this button 🖅 from toolbar to exit Lab Enquiry.
- A yellow collection slip will be generated in the Laboratory, to be used as a collection identification slip by the person collecting the blood or blood product.
- Bleep the porter/person collecting the blood and inform them that a unit of blood or blood product is to be collected on the required patient.
- When the porter/person collecting the unit arrives in the laboratory to collect the unit of blood or blood product, they time-stamp the yellow collection slip.
- The yellow collection slip is then brought to the ward with the blood/ blood product, where it is again time-stamped on receipt.

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- The nurse who receives the unit of blood at the ward then signs on the appropriate line on the yellow collection slip to verify receipt of the blood/ blood product.
- When the unit of blood/ blood product is 'hung', the smaller sticky strip from the bar-coded patient identification label on the blood/blood product is stuck on the appropriate line on the yellow collection slip, and the nurse who has transfused the blood/blood product signs on the appropriate line.
- The yellow collection slips are then collected and returned to the Blood Transfusion Laboratory, where they serve as transfusion confirmation records.
- NB -When finished search click this button from toolbar to exit Lab Enquiry.

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10 ON CALL (EMERGENCY SERVICE)

The on-call service is restricted to true emergencies. The turn-around time will be adversely affected if excessive demands are made on the service.

Tests Available On-Call

| Test | Laboratory | Unrestricted | Restricted Requiring |
|---|-------------------|---------------------------------------|-------------------------|
| | | | Consultation |
| Alanine amino Transferase | Biochemistry | ✓ | |
| Albumin | Biochemistry | ✓ | |
| Alkaline phosphatase | Biochemistry | ✓ | |
| Ammonia | Biochemistry | ✓ | |
| Amylase | Biochemistry | ✓ | |
| Antibiotic Assays | Microbiology | ✓ | |
| Antibody Screen | Blood Transfusion | \checkmark | |
| APTT | Haematology | \checkmark | |
| Aspartate amino Transferase (AST) | Biochemistry | \checkmark | |
| Blood Cultures | Microbiology | \checkmark | |
| Blood gases | Biochemistry | ✓ | |
| B-HCG (Blood) ¹ | Biochemistry | ✓ | |
| Calcium | Biochemistry | \checkmark | |
| Carbamazapine (Tegretol) ² | Biochemistry | | \checkmark |
| Carboxyhaemoglobin | Biochemistry | \checkmark | |
| Chloride | Biochemistry | ✓ | |
| Cold Agglutinins | Blood Transfusion | | ✓ |
| CAPD Fluid | Microbiology | ✓ | |
| Creatine kinase (CK) | Biochemistry | \checkmark | |
| Creatinine | Biochemistry | \checkmark | |
| C R P (C-Reactive Protein) | Biochemistry | \checkmark | |
| CSF Microscopy and Culture | Microbiology | ✓ ✓ | |
| CSF Protein and Glucose | Biochemistry | ✓ ✓ | |
| Digoxin ² | Biochemistry | • | \checkmark |
| Direct Bilirubin | Biochemistry | ✓ | |
| Direct Coombs Test | Blood Transfusion | · ✓ | |
| ESR | | · · | |
| Ethanol ² | Haematology | • | ✓ |
| | Biochemistry | | ▼ ✓ |
| Epanutin (Phenytoin) ² | Biochemistry | | ▼ ✓ |
| Epilim (Sodium Valproate) ² | Biochemistry | √ | • |
| Gamma GT (GGT) | Biochemistry | ▼ ✓ | |
| Fibrinogen | Haematology | ▼ ✓ | |
| Full Blood Count (FBC) | Haematology | | |
| Glucose | Biochemistry | ✓ | |
| Group and Coombs | Blood Transfusion | , | ✓ |
| Group and Crossmatch ³ | Blood Transfusion | ✓ | |
| Group and Hold | Blood Transfusion | ✓ | |
| HIV Ag/Ab, HBsAg, HCV antibody (Needlestick | Microbiology | \checkmark | |
| Injury - Source) | | | |
| HIV Ag/Ab, HBsAg, HCV antibody, Anti-HBs (Needlestick Injury - Victim) | Microbiology | \checkmark | |
| INR | Haematology | ✓ | |
| Influenza ⁸ | Microbiology | | ✓ |
| Iron ² | Biochemistry | | ✓ |
| Kleihauer testing | Haematology | | ✓ |
| Lactate | Biochemistry | ✓ | |
| Lactate Dehydrogenase (LDH) | Biochemistry | ✓ | |
| Lithium ² | Biochemistry | | ✓ |
| Magnesium | Biochemistry | ✓ | |
| Malaria Screen | Haematology | · · · · · · · · · · · · · · · · · · · | |
| Methaemoglobin | Biochemistry | ✓ ✓ | |

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| Test | Laboratory | Unrestricted | Restricted Requiring Consultation |
|--|-------------------|--------------|---|
| Microbiology – urgent samples ⁴ | Microbiology | \checkmark | |
| Osmolality | Biochemistry | \checkmark | |
| Paracetamol | Biochemistry | \checkmark | |
| Phenotyping Red Cell Antigens | Blood Transfusion | \checkmark | |
| Phosphate | Biochemistry | \checkmark | |
| Pregnancy Test | Haematology | \checkmark | |
| Potassium | Biochemistry | \checkmark | |
| Prolactin ⁵ | Biochemistry | | \checkmark |
| Protein – Total | Biochemistry | \checkmark | |
| Reticulocytes | Haematology | ✓ | |
| Salicylate | Biochemistry | \checkmark | |
| SARS CoV 2 ⁹ | Microbiology | | ✓ |
| Sickle Cell Screen | Haematology | \checkmark | |
| Sodium | Biochemistry | ✓ | |
| Theophylline ² | Biochemistry | | \checkmark |
| Total bilirubin | Biochemistry | \checkmark | |
| Transfusion Reaction Investigation | Blood Transfusion | \checkmark | |
| Troponin I ⁶ | Biochemistry | \checkmark | |
| Urate | Biochemistry | \checkmark | |
| Urea | Biochemistry | \checkmark | |
| Urinary creatinine | Biochemistry | \checkmark | |
| Urinary electrolytes | Biochemistry | \checkmark | |
| Urinary urea | Biochemistry | \checkmark | |
| Urinary Osmolality | Biochemistry | \checkmark | |
| Urine Microscopy & Culture (urgent e.g. A/E) | Microbiology | ✓ | |

Notes:

- 1. Urgent Beta HCG requests only will be processed.
- 2. Currently analysis of these drugs (TDM) is only available in an 'over-dose' situation. Routine monitoring of the anti-epileptic drugs, digoxin and theophylline on Saturday and Sunday mornings.
- 3. Blood is crossmatched only for Emergency purposes. Requests for blood for planned transfusion will generally not be crossmatched during emergency "On Call" hours and will be processed on the next routine working day.
- Sterile body fluids marked "special attention" or "emergency". Sputa and swabs (excluding MRSA screens and HVS) marked "special attention" or "emergency" daily up to 8pm.
- 5. Prolactin requests will be processed only to exclude a prolactin-secreting tumour when emergency surgery is contemplated.
- 6. Troponin I requests which fulfil the agreed criteria.
- 7. All Coagulation Factor assays must be requested by prior approval by Consultant Haematologist On-Call.
- 8. Emergency Influenza testing provided up to 23:00 hrs during influenza season
- 9. SARS CoV 2 routine service available up to 20:00 hrs week days and emergency requests up to 20:00 must be clinically approved by Microbiology Medical team.

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11 BLOOD TRANSFUSION

| Laboratory Profile: | The Blood Transfusion Laboratory at CUH provides testing and advice to users in relation to general transfusion issues including antenatal blood group serology. Since September 2008, it operates a quality management system to ISO15189 & AML–BB standards and since then time has been accredited by the Irish National Accreditation Board (INAB) - reference 199MT (details available from www.inab.ie). The laboratory continues to actively engage in the accreditation process to ensure compliance with the EU Blood Directive 2002/98/EC and other relevant legislation and works closely with Haemovigilance personnel to ensure all aspects of best transfusion practice, Haemovigilance and Traceability requirements are maintained. |
|---------------------------------------|--|
| | In 2022: 19,875 blood group and antibody screen investigations were performed in the crossmatch section of the laboratory. 11,341 blood group and antibody screen investigations were performed in the antenatal section of the laboratory. 1,618 infant blood group and DCT specimens were processed 1,435 cffDNA Screening requests were referred 253 HLA B27 Investigations were performed 15,759 donor red cell units were crossmatched 8,140 units of red cells were transfused. 1,426 units of plasma were transfused. 2,149 units of platelets were transfused. 838 gms Fibrinogen concentrate were transfused 1,542 vials of Anti-D Ig were transfused |
| | The laboratory also plays an important role in the care and management of antenatal patients and those patients who may require transfusions with various blood components or products while in hospital. |
| Hospital Transfusion Committee: | A Hospital Transfusion Committee exists within CUH and is co-ordinated by blood transfusion laboratory personnel. This committee meets at least 4 times per year and its remit is to promote the highest standard of transfusion practice through peer review and advocate a high standard of care in Cork University Hospital (CUH) and Cork University Maternity Hospital (CUMH) for patients at risk of transfusion (i.e. those who must be transfused, and also those who, with good clinical management, may avoid the need for transfusion). The committee also monitors that the conditions and requirements of the EU Blood Directive 2002/98/EC including articles 14 and 15 in relation to Traceability and Haemovigilance are implemented at CUH and CUMH. Representatives of users of the blood transfusion laboratory service are essential and welcome on the committee. It provides a forum for information exchange and is chaired by a consultant haematologist (see list above). |
| Tests available: | The following table outlines the tests available from the Blood Transfusion Laboratory, CUH. Details of tests are contained in the A to Z section of this Handbook. |

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| Sample bottles & Request Forms | Sample bottles and request forms may be obtained from CUH Stores. |
|-----------------------------------|--|
| | It is very important that sample tubes used are within their expiry date. |
| | Please note that expired sample bottles may be rejected and repeat samples requested |
| On-call services: | The routine day in the blood transfusion laboratory 08:00-20:00 Monday-Friday and 09:00-12:30 Saturday |
| | Outside of these hours the transfusion operates an on-call schedule whereby only emergency samples are processed during on-call hours. |
| | The on-call service is provided by a single staff member and is contactable by the bleep system #199. |
| | The list of tests available during out-of-hours on-call times are listed in this handbook with specific notes as appropriate. |
| | Samples for elective procedures should be brought directly to the laboratory before 5 p.m. on the day prior to surgery. |
| | It cannot be guaranteed that blood will be ready for elective surgery the following morning if samples arrive in the laboratory after this time. |
| Consent: | Upon admission to the CUH, it is understood that consent is given by the patient by way of signature for any treatment deemed necessary by medical personnel that includes transfusion of blood and/or blood products. |

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Consent is required for HLA B27 typing (see section 12 TEST DIRECTORY for further details)

Turnaround time: Turnaround time (TAT) is defined as the time from receipt of specimen in the laboratory until the result (and/or blood is issued) is reported either in the computer or by phone. The Blood Transfusion Laboratory will attempt to meet the turnaround times outlined in the test directory A to Z section of this handbook, subject to the availability of sufficient resources.

- The laboratory operates a "zero-tolerance policy" in relation to sample labelling which is in line with internationally recognised BSH Guidelines. Inadequately labelled samples must be resampled.
- The presence of antibodies may lead to delays in the provision of blood in both emergency and non-emergency situations. It is therefore essential that samples for routine elective surgeries be sent to the laboratory to arrive no later than 5 p.m. on the previous working day to ensure blood will be ready.
- On occasion, the laboratory may request additional or repeat samples. This may be due to the investigation of unusual results, poor sample quality (e.g. haemolysis, labelling errors) or patients requiring several crossmatches etc.

Laboratory Important considerations for blood transfusion laboratory requests: Requests:

Blood transfusion samples are only valid for 72 hours.

For **<u>urgent requests</u>**, the requestor must contact the blood transfusion laboratory by phone (routine hours) or bleep (on call hours)

From the patient perspective, there are no specific requirements in terms of fasting etc. with regard to preparation prior to sample collection.

The volume of blood sample required for blood transfusion testing should be sufficient to meet the needs of testing procedures requested. The volumes required are outlined in A to Z section.

Sampling & Labelling of Blood Transfusion Samples

Blood transfusion samples may only be taken by Doctors or specially trained Nurses/Midwives at CUH/CUMH.

Request forms and samples for blood transfusion laboratory requests from all users of the service MUST be

- handwritten or
- labelled with a BloodTrack personal digital assistant (PDA) label or
- labelled using the MN_CMS system (CUMH)
- The **BloodTrack PDAs** are an IT based solution intended to prevent sample labelling errors. The PDAs work by scanning a barcode on the user's ID badge and then scanning a barcode on the patient's wristband, which encodes the patient's demographics (forename,

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surname, date of birth & medical record number). The user's details and patient's demographics are then printed on a label, which can be attached to the blood transfusion sample & Request Form.

 The CUMH uses the MN_CMS (Maternity Newborn Clinical Management System) Millennium Electronic record. Transfusion sample labels & Request Forms generated correctly through the MN_CMS EHR are accepted in the CUH Blood Transfusion Department.

Essential information required on both samples and Request Forms MUST include:

- Patient's Forename
- Patient's Surname
- MRN (in case of GP samples where no MRN available the address is to be used)
- Date of Birth
- Identity of person taking the sample (Doctor/dedicated nurse) including bleep/contact number. Ideally, Doctors should include their MCRN, Nurses/Midwives should include An Bord Altranais PIN.
- Date and time that the sample was taken.

Adequate completion of requests SHOULD include clinical information so that work may be prioritised and processed accordingly in the laboratory (e.g. obstetric history, transfusion history, reason for transfusion etc.).

Unconscious patients admitted to the emergency department should be identified using the system as agreed with the blood transfusion laboratory, CUH as detailed in local instructions (Please be familiar with current instructions in the emergency department).

In the event of a major incident when many patients may be admitted at the same time, the labelling protocols should be used as described in the local major incident policies available in the Emergency Department. Refer to PPG-CUH-CUH-215 for additional information.

Transport of Blood Transfusion Samples

Samples should be transported to the laboratory using the guidelines described in this document.

All inpatient samples should be brought directly into the laboratory and not left at Laboratory Reception.

Urgent samples sent using the pneumatic chute system must be accompanied with a telephone call or bleep to alert Laboratory personnel.

Samples should arrive in the laboratory no later than 48 hrs after sampling.

Materials used in the collection of primary samples should be disposed of in accordance with local health and safety guidelines.

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Red cell concentrates are stored from 2-6°C in temperature-controlled and monitored fridges, which can only be accessed by trained authorised personnel.

Additional red cell concentrates are ordered by contacting the CUH Blood Transfusion Laboratory (phone or bleep) and by sending a fully completed Blood Product Requisition Form (LF-C-BTR-PROREQ) to the laboratory. Addressograph labels may be used on this form however; the requestor MUST sign this form.

It is important to note that the sample used for that crossmatch is only valid for 72 hours from the time of sampling after which time a new sample is required.

For urgent requests, once labelled and prepared, the laboratory will contact the requesting location when the red cell concentrates are ready for collection

Prior to the collection of red cell concentrate for transfusion from the blood transfusion laboratory, it is recommended that the clinical area review the most recent haemoglobin result.

Ward staff generate a collection slip either:

- electronically through the Blood Track Enquiry Function (CUH) or the MN_CMs system (CUMH) (these electronic collection slips print in the laboratory for the porter to access) OR
- they complete a manual collection slip (handed directly to porter)

Red cells should transfused within 4 hours of 'spiking' the pack and/or 4 $\frac{1}{2}$ hours of removal from the blood fridge/igloo, whichever is sooner. They should be returned to the laboratory if the transfusion is unduly delayed.

For further details on the collection process and administration of red cell concentrates refer to procedure PPG-CUH-CUH-13 Platelets are stored from 20-24°C on in temperature-controlled and

monitored platelet agitator in the blood transfusion laboratory which can only be accessed by trained authorised personnel.

Platelet components are ordered by contacting the CUH Blood Transfusion Laboratory (phone or bleep) and by sending a fully completed Blood Product Requisition Form (LF-C-BTR-PROREQ) to the laboratory. Addressograph labels may be used on this form however; the requestor MUST sign this form.

Laboratory personnel may have to request a sample for blood grouping if no record of blood group is available in the laboratory. Laboratory personnel will arrange the delivery of platelets from IBTS. It may not always be possible to have ABO compatible platelets available from IBTS, so laboratory personnel may need to confirm suitability with requesting clinician.

Storage, Ordering and Collection of Platelets:

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Storage, Ordering and

Collection of

plasma (*i.e. LG-Octaplas*), For urgent requests, once labelled and prepared, the laboratory will contact the requesting location when the platelets are ready for collection

Platelets should not be stored at ward level and should be returned to the laboratory immediately if not being used immediately.

For further details on the collection process and administration of platelet components refer to procedure PPG-CUH-CUH-13

Plasma is stored at less than -18°C in temperature-controlled and monitored freezers, which can only be accessed by trained authorised personnel.

Plasma areordered by contacting the CUH Blood Transfusion Laboratory (phone or bleep) and by sending a fully completed Blood Product Requisition Form (LF-C-BTR-PROREQ) to the laboratory.

Addressograph labels may be used on this form however; the requestor MUST sign this form.

Plasma products are thawed in the laboratory upon request and requires 30-45 minutes to be prepared depending on the number of units required.

For urgent requests, once labelled and prepared, the laboratory will contact the requesting location that the plasma is ready.

Once thawed, they are stored at 2-8°C in the laboratory and once collected it is recommended that they are used within 8 hours from thawing. If the product is not being transfused the product should be returned to the laboratory immediately.

For further details on the collection process and administration of plasma components refer to procedure PPG-CUH-CUH-13

Plasma is NOT routinely necessary in the management of overanticoagulation with warfarin and the National Haemovigilance Office has issued the following guidelines:

| Coagulation Status of Patient | Corrective Action |
|---|---|
| INR result between 3.0-6.0 (target 2.5) | 1. Reduce warfarin dose or stop. |
| INR result between 4.0-6.0 (target 3.5) | 2. Restart warfarin when INR < 5.0 |
| INR result between 6.0-8.0 with no | 1. Stop Warfarin |
| bleeding or minor bleeding. | 2. Restart warfarin when INR < 5.0 |
| INR result >8.0 with no bleeding or minor | 1. Stop warfarin |
| bleeding | 2. Restart warfarin when INR < 5.0 |
| | 3. If other risk factors for bleeding exist, give |
| | 0.5-2.5 mg of oral or I.V. Vitamin K. |
| Life-threatening bleed | 1. Stop warfarin |
| | 2. Give Prothrombin complex concentrate (e.g |
| | Octaplex) (50IU/kg) or Plasma (15 mL/kg) |
| | 3. Give 5mg of oral or I.V. Vitamin K |

Note: The maximum recommended Prothrombin Complex Concentrate dose is 3000 IU

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| Storage, Ordering and Collection of other blood products e.g. Prothrombin | All blood products issued by the blood transfusion laboratory are stored according to manufacture instructions at either room temperature (monitored) in the laboratory or at 2-8°C in temperature controlled and monitored fridges, which can only be accessed by trained authorised personnel. |
|--|--|
| Complex Concentrate (<i>i.e.</i> Octaplex), Albumin, Fibrinogen Concentrate, | Products are ordered by contacting the CUH Blood Transfusion Laboratory (phone or bleep) and by sending a fully completed Blood Product Requisition Form (LF-C-BTR-PROREQ) to the laboratory. Addressograph labels may be used on this form however; the requestor MUST sign this form. |
| Clotting Factor Concentrates, etc. | The blood transfusion laboratory holds a minimum stock level of all blood products supplied by the laboratory. |
| | Should the requestor have a requirement for a substantial quantity of any particular product, the requestor where possible should contact the laboratory so that additional product may be ordered. |
| | For urgent requests, once labelled and prepared, the laboratory will contact the requesting location that the blood product is ready for collection. |
| | All blood products should be transfused a soon as possible on arrival on the ward and if there is any undue delay in the commencement of the transfusion, the blood product should be returned to the laboratory |
| | For further details on the collection process and administration of blood products refer to procedure PPG-CUH-CUH-13 |
| Storage, Ordering and Collection of Anti-D | Anti-D Immunoglobulin is stored from 2-6°C in temperature-controlled and monitored fridges, which can only be accessed by trained authorised personnel. |
| Immunoglobulin | Anti-D Immunoglobulin are ordered by contacting the CUH Blood Transfusion Laboratory (phone or bleep) and by sending a fully completed Blood Product Requisition Form (LF-C-BTR-ANTID) to the laboratory. Anti-D Immunoglobulin is primarily transfused in the CUMH and the Blood Product Requisition Form can be generated electronically through the MN_CMS system (See note below) |
| | For urgent requests, once labelled and prepared, the laboratory will contact the requesting location that the anti-D immunoglobulin is ready. |
| | Anti D immunoglobulin should be transfused a soon as possible on arrival on the ward and if there is any undue delay in the commencement of the transfusion, the blood product should be returned to the laboratory |
| | For further details on the collection process and administration of blood products refer to procedure PPG-CUH-CUH-13 and PPG-CUH-MAT-5 |

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Note: For <u>all</u> blood component and blood products requests from the CUMH, the MN_CMS system allows the user to generate an electronic Blood Product Requisition Form.

The CUMH user must still contact the CUH Blood Transfusion Laboratory (phone or bleep) and send either the electronic or manual Blood Product Requisition Form to the laboratory

| Storage of samples in the Blood | Blood transfusion samples are stored for 72 hours in controlled monitored storage 2-8°C. |
|---|---|
| Transfusion Laboratory: | After this time, samples are disposed in accordance with local policies. |
| Emergency Blood Requests | A limited number of O Rh(D) Negative Blood are available for EXTREME emergency situations. These units are stored in selected locations which include the blood transfusion laboratory issue fridge and the theatre reception fridge. The laboratory must be informed if these units are used and the accompanying form must be fully completed and returned to the laboratory. For further information refer to procedure PPG-CUH-CUH-210 |
| Pre-Hospital Transfusion: | The Blood Transfusion laboratory in conjunction with the CUH Emergency Department run a successful pre-hospital blood transfusion project whereby blood is taken from the transfusion laboratory to the scene of an incident and may be transfused at the scene. For further information refer to procedure PPG-CUH-CUH-282 |
| | This entire transfusion chain is governed by the laboratory's quality management system to the ISO15189 standards and is fully compliant with the EU Blood Directive 2002/98/EC and other relevant legislation in terms of best transfusion practice, Haemovigilance and Traceability. |
| Blood transferred with a patient from an external location: | Any blood transferred to the CUH/CUMH with a patient from an external source (e.g. another hospital) should be brought directly to the blood transfusion laboratory. It is essential that any documentation accompanying the blood is completed accordingly and given to the transfusion laboratory personnel. It is imperative that the storage conditions of blood 'in transit' are controlled. |
| | It is also necessary to obtain a fresh group and hold sample as soon as possible from such patients so that should additional blood be required, it can be used for crossmatching in the CUH blood transfusion laboratory. |
| General Haemovigilance: | Haemovigilance may be defined as: "a set of surveillance procedures, from the collection of blood and its components, to the follow up of recipients to collect and assess information on unexpected or undesirable effects resulting from the therapeutic use of labile blood products, and to prevent their occurrence or recurrence" (National Haemovigilance Office, 2004.) |
| | Since 2005 the role of the Haemovigilance staff has been greatly influenced by the transposition into Irish law of the EU Blood Directive 2002/98/EC. The directive became law in Ireland on the 8 th February 2005 and has implications for all hospital blood banks. Eight articles apply directly to all |

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| staff involved in the transfusion process throughout the hospital. The major implications involve the implementation of quality systems for all aspects of transfusion, the total traceability of every blood product, the training of |
|--|
| personnel involved in the transfusion process and the reporting of any |
| serious adverse reactions or events associated with the transfusion of blood components. Compliance with this legislation is policed by the Health Products Regulatory Authority (HPRA, formerly known as the Irish Medicines |
| Board) under the HPRA Act 1995 and in the event of directive non- compliance; the HPRA has censure authority up to and including the closure |
| of a facility |

The remit of the haemovigilance personnel includes the following:

- Promotion of safe and effective transfusion practice for those receiving blood components/products.
- Participation in local working groups and on a national basis to promote the safe and effective transfusion practice for those receiving blood components/products.
- Provision of educational programmes for staff involved in the transfusion process
- Participation in and development of audit initiatives as appropriate.
- Development and maintenance of effective channels of communication by encouraging networking, support and cross-clinical group working.
- Contribution to the shaping of policy relating to transfusion of blood components by responding to local and national developments
- Investigation of any serious adverse reactions or events associated with the transfusion of blood components.
- Maintenance of blood component traceability.

Haemovigilance Training and Policies Haemovigilance personnel have put policies and procedures in place via the Q-Pulse document management system in CUH promoting good transfusion practice in clinical areas. Scheduled Haemovigilance education sessions are provided by Haemovigilance personnel to all clinical staff. Clinical staff who are unable to attend these scheduled training sessions should make contact with the CUH/CUMH haemovigilance personnel to arrange training.

It is CUH policy that all clinicians should have completed both (*Safe Transfusion Practice (Formerly Module 1*) and *Blood Components and Indications for Use (Formerly Module 2*) of the SNBTS LearnPro e-learning program. (*www.learnbloodtransfusion.org.uk/*). Instructions on how to access the Q-Pulse system and the SNBTSe-learning program are available from haemovigilance staff.

All hospitals have a legal requirement to trace each individual blood component, whether transfused or disposed of, in accordance with the EU Blood Directive (2002/98/EC). This information must be held and available for thirty years. Therefore, full and clear documentation associated with transfusion is essential.

All serious adverse reactions and events associated with the transfusion of blood components are investigated documented and, where required, reported to the National Haemovigilance Office (NHO) through a confidential anonymous reporting system. If you suspect a transfusion reaction, you must contact the Blood Transfusion Laboratory or Haemovigilance personnel as identified in this Handbook. There is a Policy dealing with the recognition,

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investigation and management of a Suspected Transfusion Reaction on Q-Pulse. (PPG-CUH-CUH-30).

| | The decision to transfuse is the responsibility of the prescribing clinician and should be based on the best available evidence. The prescribing clinician should discuss the transfusion with the patient in accordance with hospital policy (PPG-CUH-CUH-80), document this discussion in the patient's medical notes and should give the patient the 'Having a Blood Transfusion – Information Leaflet for Patients and Guardians' (INF-CUH-CUH-9). The information leaflets are available from the Stationary Stores Department. Where clinically possible it is recommended that blood transfusions should only be given during routine working hours. There is a policy available on Q-Pulse which details the procedure required for the prescription of blood & blood components. This policy also details the correct procedure for the taking of the pre-transfusion sample by medical staff. (PPG-CUH-CUH-36). |
|--|---|
| | covered in the policy PPG-CUH-CUH-13, available on Q-Pulse. |
| Results | Results are issued in Hard Copy report format. Note: In the CUMH, transfusion results are available electronically through the MN_CMS Millennium Electronic Health Record. For any staff with access to transfusion results electronically, it is their responsibility to ensure that they satisfy themselves that the blood transfusion laboratory has a valid transfusion specimen and/or products available. |
| | It is the general policy of the laboratory not to issue results over the phone. Copy reports can be printed on request. In accordance with HSE policy, faxing of results can be facilitated in exceptional circumstances only. Users will be asked to fax a request for a faxed report, to ensure the laboratory can fax report to a secure fax number. |
| Advice and consultation: | Should clarification be sought on any issues related to the Blood Transfusion Laboratory service at CUH, queries may be directed to Blood Transfusion Laboratory or Haemovigilance personnel as identified in this Handbook. |
| Complaints /Positive Feedback | The Blood Transfusion Laboratory at CUH endeavours to produce a system of continual improvement to meet the needs and requirements of users and in the best interest of patients. To facilitate this, the Blood Transfusion Laboratory welcomes all feedback (both Negative and Positive) and users can provide feedback by way of telephone call, email or in hard copy writing to contacts provided. All feedback will be processed in accordance with the laboratory's feedback / complaints system. |
| Data Protection / Patient Information Code of Conduct: | All staff in the laboratory are made aware of their responsibilities in relation to protection of personal patient information consistent with the Data Protection Act 2018 and Freedom of Information Act 2003. All records are retained in accordance with requirements outlined in EU Blood Directive 2002/98/EC and securely managed in accordance with local laboratory instruction MI-C-BTR-RECORDM. |
| Contingency | In the event that the laboratory's computer system fails, a manual contingency plan is in place. Users may be informed that a manual back-up |

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system is in place and are requested to facilitate the laboratory by limiting requests to 'urgent requests' only, while IT systems are restored.

In the extremely unlikely event that the laboratory is unable to provide a service (e.g. Fire/Flood Damage), the IBTS may provide a back-up service. Users may be requested to facilitate the laboratory by limiting requests to 'urgent requests' only, until service is restored on site in CUH.

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12 TEST DIRECTORY (A-Z)

Acanthamoeba (amoebic keratitis)

| Laboratory: | Microbiology (Main lab | oratory) | |
|--------------------------|---|---|--|
| Specimen: | | ected onto a specific swab obtained directly from the | |
| | Microbiology Laborato | • | |
| Comment: | | rted directly to microbiology where it will be referred to | |
| Turnaround: | the UK for PCR testing. Testing performed by Micropathology Ltd, Coventry. 1 week (1 working day from receipt of swab in UK) | | |
| Report: | | etected or not detected. | |
| | (corneal scrape) | | |
| Laboratory: | Neuropathology | | |
| Specimen: | Corneal scrape – spec | ial fixative required, (CytoLyt) available from | |
| | Neuropathology Labor | | |
| Comment: | - | athology Department in advance on 4922520 | |
| Turnaround: | 3 weeks – Positive res | ults phoned | |
| ACTH | | | |
| Laboratory: | - | CUH Biochemistry to Eurofins-Biomnis Laboratories | |
| Specimen: | frozen < 30 minutes | n EDTA available from Biochemistry) on ice, must be | |
| Comment | Consultant request on | lv | |
| Turnaround: | 3 weeks | , | |
| Ref. Range: | See report form, or vis | sit internet site https://www.eurofins.ie/biomnis/ for up to | |
| _ | date referral test infor | mation. | |
| Activated Parti | al Thromboplastin Ti | me (APTT) | |
| Laboratory: | Haematology | | |
| Specimen: | | /acuette® (sodium citrate 3.2%) | |
| | | haemolysed, under filled or overfilled cannot be lation sample bottles are not expired to ensure correct | |
| | filling.) | nation sample bottles are not expired to ensure correct | |
| Comment: | - , | used to evaluate abnormalities in the Intrinsic | |
| | 2 / | and to monitor the effectiveness of heparin therapy. | |
| | | Thrombophilia and /or Lupus screen. See Main | |
| | | on Guidelines for Investigation of Thrombophilia. | |
| | sampling. | mens should arrive in the laboratory within 4 hours of | |
| | | to Friday, during routine working hours, and for | |
| | emergency reasons at | all other times. | |
| Turnaround: | | nours. Ward specimens: 8 hours | |
| Ref. Range: | Age Mear | 5 () | |
| | Day 1 43 | 31 - 55 | |
| | Day 5 43 | 25 - 60 | |
| | Day 30 41 | 26 - 55 | |
| | Day 90 37 | 24 - 50 | |
| | Day 180 36 | 28 - 43 | |
| Activated Broth | Adult 27 | See final report | |
| | Haematology | | |
| Laboratory: Specimen: | | ette [®] (sodium citrate 3.2%) | |
| opeemen | - | haemolysed, underfilled or overfilled cannot be | |
| | | ulation sample bottles are not expired to ensure | |
| | correct filling) | | |

| | Iedicine User Handbook Reference: PPG-CUH-PAT-31 Revision: 22 Active Date: 02(11/2023) Date: 02(11/2023) | | | |
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| | Author: Mr Paul Cantwell | | | |
| | | | | |
| Comment: | Test available Mon to Fri, during routine working hours. This test forms pa | | | |
| | of a Thrombophilia Screen, used as a screening test for Factor V Leiden | | | |
| | mutation, see Main Haematology Section on Guidelines for Investigation of | | | |
| | Thrombophilia (if positive an EDTA sample is confirmed by PCR analysis). | | | |
| | Samples must be received within 4 hours. | | | |
| | Thrombophilia request form FOR-CUH-PAT-1575, including documentation | | | |
| | of patient consent, must be received with all requests and is available on | | | |
| | the CUH website. | | | |
| Turnaround: | 3 – 4 weeks (Refer to the main Haematology Section on Coagulation). | | | |
| Report: | Ratio > 0.7 Negative | | | |
| | Ratio ≤ 0.70 Positive | | | |
| cyl Carnitine, | | | | |
| Laboratory: | Sample referred from Clinical Biochemistry to The Children's Hospital, | | | |
| Laboratory. | Temple Street, Dublin | | | |
| Chacimany | • • | | | |
| Specimen: | Newborn screening card. 2 full circles | | | |
| Comment: | Consultant request only | | | |
| Turnaround: | 3 weeks | | | |
| Ref. Range: | See report form. | | | |
| denovirus Mo | | | | |
| Laboratory: | Microbiology (Infectious Diseases Serology) | | | |
| Specimen: | 4mL clotted blood, 4mL EDTA blood, viral swab (eye, throat), stool, | | | |
| | nasopharyngeal aspirate, sputum, broncho-alveolar lavage, CSF, urine | | | |
| Comment: | Performed by a reference laboratory (National Virus Reference Laboratory | | | |
| | (NVRL), Dublin) | | | |
| Turnaround: | 14 working days | | | |
| Report: | Detected or not detected | | | |
| denovirus (fa | neces samples) | | | |
| See Rotavirus/A | denovirus assay | | | |
| Adrenal Antibo | odies | | | |
| Laboratory: | Sample referred from Autoimmune Serology to Eurofins-Biomnis | | | |
| | Laboratories | | | |
| | | | | |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) | | | |
| Specimen: Turnaround: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) Approx. 3 Weeks | | | |
| | Approx. 3 Weeks | | | |
| Turnaround: | | | | |
| Turnaround: Ref. Range: | Approx. 3 Weeks See report form, or visit internet site <u>https://www.eurofins.ie/biomnis/</u> for up to date referral test information. | | | |
| Turnaround: Ref. Range: | Approx. 3 Weeks See report form, or visit internet site <u>https://www.eurofins.ie/biomnis/</u> for up to date referral test information. Transferase (ALT) | | | |
| Turnaround: Ref. Range: Manine amino Laboratory: | Approx. 3 Weeks See report form, or visit internet site <u>https://www.eurofins.ie/biomnis/</u> for up to date referral test information. Transferase (ALT) Clinical Biochemistry | | | |
| Turnaround: Ref. Range: Manine amino Laboratory: Specimen: | Approx. 3 Weeks See report form, or visit internet site <u>https://www.eurofins.ie/biomnis/</u> for up to date referral test information. Transferase (ALT) Clinical Biochemistry 4.0 mL blood plain tube (clotted sample) | | | |
| Turnaround: Ref. Range: Manine amino Laboratory: | Approx. 3 Weeks See report form, or visit internet site <u>https://www.eurofins.ie/biomnis/</u> for up to date referral test information. Transferase (ALT) Clinical Biochemistry 4.0 mL blood plain tube (clotted sample) A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF | | | |
| Turnaround: Ref. Range: Manine amino Laboratory: Specimen: Turnaround: | Approx. 3 Weeks See report form, or visit internet site <u>https://www.eurofins.ie/biomnis/</u> for up to date referral test information. Transferase (ALT) Clinical Biochemistry 4.0 mL blood plain tube (clotted sample) A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF MGH: - 3 hours approx. GP or OPD- Results posted within 4 days | | | |
| Turnaround: Ref. Range: Manine amino Laboratory: Specimen: | Approx. 3 Weeks See report form, or visit internet site <u>https://www.eurofins.ie/biomnis/</u> for up to date referral test information. Transferase (ALT) Clinical Biochemistry 4.0 mL blood plain tube (clotted sample) A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF MGH: - 3 hours approx. GP or OPD- Results posted within 4 days Up-to-date reference intervals will be applied to all Biochemistry reports as | | | |
| Turnaround: Ref. Range: Manine amino Laboratory: Specimen: Turnaround: Ref. Range: | Approx. 3 Weeks See report form, or visit internet site <u>https://www.eurofins.ie/biomnis/</u> for up to date referral test information. Transferase (ALT) Clinical Biochemistry 4.0 mL blood plain tube (clotted sample) A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF MGH: - 3 hours approx. GP or OPD- Results posted within 4 days Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate | | | |
| Turnaround: Ref. Range: Manine amino Laboratory: Specimen: Turnaround: Ref. Range: | Approx. 3 Weeks See report form, or visit internet site <u>https://www.eurofins.ie/biomnis/</u> for up to date referral test information. Transferase (ALT) Clinical Biochemistry 4.0 mL blood plain tube (clotted sample) A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF MGH: - 3 hours approx. GP or OPD- Results posted within 4 days Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate d) | | | |
| Turnaround: Ref. Range: Manine amino Laboratory: Specimen: Turnaround: Ref. Range: Mbumin (Blood Laboratory: | Approx. 3 Weeks See report form, or visit internet site <u>https://www.eurofins.ie/biomnis/</u> for up to date referral test information. Transferase (ALT) Clinical Biochemistry 4.0 mL blood plain tube (clotted sample) A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF MGH: - 3 hours approx. GP or OPD- Results posted within 4 days Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate d) Clinical Biochemistry | | | |
| Turnaround: Ref. Range: Manine amino Laboratory: Specimen: Turnaround: Ref. Range: Mbumin (Blood Laboratory: Specimen: | Approx. 3 Weeks See report form, or visit internet site <u>https://www.eurofins.ie/biomnis/</u> for up to date referral test information. Transferase (ALT) Clinical Biochemistry 4.0 mL blood plain tube (clotted sample) A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF MGH: - 3 hours approx. GP or OPD- Results posted within 4 days Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate d) Clinical Biochemistry 4.0 mL in blood plain tube (clotted sample) | | | |
| Turnaround: Ref. Range: Manine amino Laboratory: Specimen: Turnaround: Ref. Range: Mbumin (Blood Laboratory: | Approx. 3 Weeks See report form, or visit internet site <u>https://www.eurofins.ie/biomnis/</u> for up to date referral test information. Transferase (ALT) Clinical Biochemistry 4.0 mL blood plain tube (clotted sample) A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF MGH: - 3 hours approx. GP or OPD- Results posted within 4 days Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate d) Clinical Biochemistry 4.0 mL in blood plain tube (clotted sample) A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF | | | |
| Turnaround: Ref. Range: Alanine amino Laboratory: Specimen: Turnaround: Ref. Range: Albumin (Blood Laboratory: Specimen: Turnaround: | Approx. 3 Weeks See report form, or visit internet site https://www.eurofins.ie/biomnis/ for up to date referral test information. Transferase (ALT) Clinical Biochemistry 4.0 mL blood plain tube (clotted sample) A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF MGH: - 3 hours approx. GP or OPD- Results posted within 4 days Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate d) Clinical Biochemistry 4.0 mL in blood plain tube (clotted sample) A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF SMOH, MGH: - 3 hours approx. GP or OPD- Results posted within 4 days | | | |
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| Turnaround: Ref. Range: Manine amino Laboratory: Specimen: Turnaround: Ref. Range: Mbumin (Blood Laboratory: Specimen: Turnaround: Ref. Range: | Approx. 3 Weeks See report form, or visit internet site https://www.eurofins.ie/biomnis/ for up to date referral test information. Transferase (ALT) Clinical Biochemistry 4.0 mL blood plain tube (clotted sample) A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF MGH: - 3 hours approx. GP or OPD- Results posted within 4 days Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate d) Clinical Biochemistry 4.0 mL in blood plain tube (clotted sample) A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF SMOH, MGH: - 3 hours approx. GP or OPD- Results posted within 4 days Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate | | | |
| Turnaround: Ref. Range: Alanine amino Laboratory: Specimen: Turnaround: Ref. Range: Albumin (Blood Laboratory: Specimen: Turnaround: | Approx. 3 Weeks See report form, or visit internet site <u>https://www.eurofins.ie/biomnis/</u> for up to date referral test information. Transferase (ALT) Clinical Biochemistry 4.0 mL blood plain tube (clotted sample) A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF MGH: - 3 hours approx. GP or OPD- Results posted within 4 days Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate d) Clinical Biochemistry 4.0 mL in blood plain tube (clotted sample) A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF SMOH, MGH: - 3 hours approx. GP or OPD- Results posted within 4 days Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate | | | |
| Turnaround: Ref. Range: Manine amino Laboratory: Specimen: Turnaround: Ref. Range: Mbumin (Blood Laboratory: Specimen: Turnaround: Ref. Range: | Approx. 3 Weeks See report form, or visit internet site https://www.eurofins.ie/biomnis/ for up to date referral test information. Transferase (ALT) Clinical Biochemistry 4.0 mL blood plain tube (clotted sample) A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF MGH: - 3 hours approx. GP or OPD- Results posted within 4 days Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate d) Clinical Biochemistry 4.0 mL in blood plain tube (clotted sample) A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF SMOH, MGH: - 3 hours approx. GP or OPD- Results posted within 4 days Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate | | | |

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| | Addion | | |
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| | m Clinical Bioch | emistry to Eurofir | s-Biomnis Laboratori |
| | | | |
| 4.0 mL blood in EDTA. State if the subject was standing (after at least 1 | | | |
| | | • | 2 (|
| - / | • | | , |
| 3 weeks | | | |
| See report form, or | visit internet si | te https://www.euroi | fins.ie/biomnis/ for up to |
| date referral test in | formation. | • | |
| natase (Alk Phos) | | | |
| Clinical Biochemistr | У | | |
| 4.0 mL blood in plai | in tube (clotted | sample) | |
| | | mins approx. CUH | wards, CUMH, SI, SF |
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| • | in tube (clotted | sampie) | |
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| | result is <1g/L, | sample is referred | to the Alpha 1 |
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| Sample referred fro College of Surgeons Hospital, Dublin 9. | | | 1 Foundation, Royal rch centre, Beaumont |
| Sample referred fro College of Surgeons | | | · · · · · |
| | 1 Day Up-to-date reference appropriate inine Ratio (urine) Clinical Biochemistr Spot urine 1 Day Up-to-date reference appropriate bi) (See also Toxice Clinical Biochemistr 4.0 mL blood in glue tube (clotted sample Do Not use alcohol for screening for alco purposes only. Sam testing 1 Day Up-to-date reference appropriate enin ratio Sample referred fro (Paediatric samples 4.0 mL blood in EDT hour of walking) or Consultant request 3 weeks See report form, or date referral test in hatase (Alk Phos) Clinical Biochemistr 4.0 mL blood in plai A/E or urgent samp SMOH, MGH: - 3 ho GP or OPD- Results Up-to-date reference appropriate (psin Clinical Biochemistr 4.0 mL blood in plai A/E or urgent samp SMOH, MGH: - 3 ho GP or OPD- Results Up-to-date reference appropriate | Active Date: Approved By: Author: 1 Day Up-to-date reference intervals will appropriate inine Ratio (urine) Clinical Biochemistry Spot urine 1 Day Up-to-date reference intervals will appropriate Obl (See also Toxicology Screen) Clinical Biochemistry 4.0 mL blood in glucose tube, (Sod tube (clotted sample) or in Lithium Do Not use alcohol swabs. For acut for screening for alcohol abuse. Alc purposes only. Samples will not be testing 1 Day Up-to-date reference intervals will appropriate enin ratio Sample referred from Clinical Bioch (Paediatric samples sent to Leeds C 4.0 mL blood in EDTA. State if the shour of walking) or recumbent (after Consultant request only 3 weeks See report form, or visit internet siddate referral test information. hatase (Alk Phos) Clinical Biochemistry 4.0 mL blood in plain tube (clotted A/E or urgent sample: - 1 hour 30 SMOH, MGH: - 3 hours approx. GP or OPD- Results posted within 4 Up-to-date reference intervals will appropriate (psin Clinical Biochemistry 4.0 mL blood in plain tube (clotted A/E or urgent sample: - 1 hour 30 SMOH, MGH: - 3 hours approx. GP or OPD- Results posted within 4 Up-to-date reference intervals wi | Active Date: 03/11/2023 Approved By: Dr Vitaliy Mykytiv, N Author: 1 Day Up-to-date reference intervals will be applied to all B appropriate Inine Ratio (urine) Clinical Biochemistry Spot urine 1 Day Up-to-date reference intervals will be applied to all B appropriate Oll (See also Toxicology Screen) Clinical Biochemistry 4.0 mL blood in glucose tube, (Sodium Fluoride, grey tube (clotted sample) or in Lithium Heparin tube. Spot for screening for alcohol abuse. Alcohol measuremen purposes only. Samples will not be accepted for med testing 1 Day Up-to-date reference intervals will be applied to all B appropriate Sample referred from Clinical Biochemistry to Eurofir (Paediatric samples sent to Leeds General Infirmary) 4.0 mL blood in EDTA. State if the subject was stand hour of walking) or recumbent (after at least 3 hours Consultant request only 3 weeks See report form, or visit internet site https://www.eurol date referral test information. httase (Alk Phos) Clinical Biochemistry 4.0 mL blood in plain tube (clotted sample) A/E or urgent sample: - 1 hour 30 mins approx. CUH SMOH, MGH: - 3 hours approx. GP or OPD- Results posted within 4 days Up-to-date reference intervals will be applied to all B appropriate Up-to-date reference intervals will be applied to all B appropria |

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| Alpha- Amino | Adipic Semialdehyde (á-AASA) |
|----------------|---|
| Laboratory: | Referred from Biochemistry to the Institure of Child Health, London |
| Specimen: | Spot Urine (5-10mls) on ice |
| Comment: | MUST BE FROZEN immediately. |
| | Used to support a diagnosis of Pyridoxal Responsive Epilepsy. |
| | Consultant request only |
| Turnaround: | 6-8 weeks |
| Alpha Fetoprot | |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Turnaround: | 4 Days |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate |
| Amikacin / Am | ikin |
| Refer to Antib | piotic Assays |
| Amoeba Antibe | odies |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Comment: | Performed by a reference laboratory (National Parasitology Reference Laboratory (NPRL), London) |
| Turnaround: | 28 working days |
| Report: | Qualitative result |
| Ammonia | |
| Laboratory: | Clinical Biochemistry |
| Specimen: | Blood sample in Li Hep |
| Comment: | Please inform laboratory in advance. Sample must be received to the laboratory within 30 minutes of collection and spun immediately. Haemolysis invalidates result. |
| Turnaround: | Once the lab is contacted in advance, results could be ready in approx. 1 hour 15mins |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate |
| Amphetamine | |
| Laboratory: | Sample referred from Clinical Biochemistry to Toxicology Laboratory BEAUMONT Hospital Dublin, posted Monday, Tuesday, Wednesday and Thursday. |
| Specimen: | Spot urine |
| Comment: | See Toxicology / Drug Screen |
| Turnaround: | 1 week |
| Ref. Range: | See report form or contact Toxicology Laboratory BEAUMONT Hospital 01- 8092673 / (01)8092675, Emergency after hours (087) 2590749, Fax (01) 8093986 |
| Amylase (Bloo | d) |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Turnaround: | A/E or urgent sample: - 1 hour 30mins approx. CUH wards, CUMH, SI, SF, SMOH, MGH: - 3 hours approx. |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate |

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| Amylase (Urii | narv) | | | |
|---------------|--|--|--|--|
| Laboratory: | Clinical Biochemistry | | | |
| Specimen: | Spot or 24 hour urine sample | | | |
| Turnaround: | 1 Day | | | |
| | Ref. Range: Up-to-date reference intervals will be applied to all Biochemistry reports | | | |
| | appropriate | | | |
| Amyloid A (Se | | | | |
| Laboratory: | Sample referred from Clinical Biochemistry to National Amyloidosis Centre | | | |
| Constitution | - Royal Free Hospital | | | |
| Specimen: | Serum (0.5 ml minimum) 9 weeks | | | |
| Turnaround: | | | | |
| Ref. Range: | Hospital, +44 (0) 207 433 2800 / 2725 (Results), +44 (0) 207 433 2844 | | | |
| | (Interpretation) | | | |
| Amvloid Subt | yping (Tissue) | | | |
| | Sample referred from Pathology to National Amyloidosis Centre – Royal | | | |
| | Free Hospital | | | |
| Specimen: | FFPE tissue block | | | |
| Turnaround: | 3 months | | | |
| | To discuss clinical advice, contact National Amyloidosis Centre – Royal | | | |
| | Free Hospital, +44 (0) 207 433 2800 / 2725 (Results), +44 (0) 207 433 | | | |
| | 2844 (Interpretation) | | | |
| Androstenedi | | | | |
| Laboratory: | Sample referred from Clinical Biochemistry to St. James's University Hospital, Leeds | | | |
| Specimen: | 3.0 mL blood in a plain tube (clotted sample) | | | |
| Comment: | Consultant request only | | | |
| Turnaround: | 3 weeks | | | |
| Ref. Range: | See report form | | | |
| Angelman Sy | ndrome (AS) | | | |
| Laboratory: | Molecular Genetics in Biochemistry referred to National Centre for Medical | | | |
| | Genetics. NCMG request form is available on website, <u>www.genetics.ie/molecular</u> | | | |
| Specimen: | Infants: 1ml EDTA blood | | | |
| Specifien. | Adults 3-5ml EDTA blood | | | |
| Turnaround: | | | | |
| Report: | Sent to referring clinician by NCMG and copy of report filed in pathology | | | |
| | converting enzyme (ACE) | | | |
| Laboratory: | Clinical Biochemistry | | | |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) | | | |
| Turnaround: | | | | |
| Ref. Range: | | | | |
| iten itenger | appropriate | | | |
| Antenatal Scr | | | | |
| Laboratory: | Microbiology (Infectious Diseases Serology) | | | |
| Specimen: | 4mL clotted blood | | | |
| Tests: | Rubella IgG, hepatitis B surface antigen, HIV Ag/Ab, syphilis antibody, varicella-zoster virus (VZV) IgG | | | |
| Turnaround: | | | | |
| ranna ound. | positive in house for HIV Ag/Ab and syphilis antibody (confirmatory testing | | | |
| Poport. | required). Qualitative results: quantitative result for rubella IgG (IU/mL) | | | |
| Report: | Qualitative results; quantitative result for rubella IgG (IU/mL) | | | |

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Antenatal Serology

(Blood Group + Antibody Screen +/- Antibody Identification +/- Titration)

the allocated batch time.

| Laboratory: | Blood Transfusion Laboratory |
|-----------------|---|
| Specimen: | 1 x 6 ml EDTA Pink Capped Tube |
| Comment: | Antenatal blood grouping and antibody screening and identification in antenatal women. (Patients may also include the male partners of pregnant women for the purposes of establishing their blood groups and red cell phenotypes in the prediction of HDNB). Blood Group, Antibody Screen and Identification, Red Cell Phenotyping are INAB accredited tests. |
| | Request Form to be completed: Antenatal Serology Request Form (LF-C- BTR-ANTENAT) |
| Turnaround: | 2 days. |
| | NOTE: Samples received on Fridays and during weekends may be processed during next routine working day. |
| Ref. Range: | Not applicable |
| Antibiotic Assa | ys |
| Laboratory: | Microbiology |
| Specimen: | 4mL clotted blood, EDTA unsuitable |
| Test method: | Photometric absorbance |
| Turnaround: | Assays are batched and performed at 7am, 11am, 3pm, 7pm and 11pm. Please ensure the sample is in the laboratory at least 30 minutes before |

| Report: | Quantitative result (mg/L) |
|----------|--|
| Comment: | Available 7 days. Specify peak (post) or trough (pre). It is very difficult to interpret random specimens. All forms should indicate the time since the last administration of the drug. Please refer to the Cork University Hospital Antibiotic Guidelines. |
| | Teicoplanin levels are rarely indicated and are not processed. Streptomycin and Cycloserine levels are performed by a reference laboratory (South Mead Hospital, Bristol). Note for Gentamicin : In very rare cases, gammopathy in particular type |

Note for Gentamicin: In very rare cases, gammopathy in particular type IgM (Waldenström's macroglobulinemia) may cause unreliable results. In very rare cases, patient samples may contain particle agglutinating proteins (e.g. heterophilic antibodies or antibodies due to abnormal immunoglobulin synthesis, such as gammopathies like MGUS0 or Waldenström's macroglobulinemia) which may lead to incorrect low or high results with this assay. Please notify the laboratory when requesting a gentamicin assay if the patient has this type of gammopathy as an alternative assay method is required.

| Antibiotic - once daily dosage | Trough |
|--|-------------------------------|
| Amikacin - once daily dosage | <5 mg/L |
| Gentamicin - once daily dosage | <1 mg/L |
| Tobramycin - once daily dosage | <1 mg/L |
| Vancomycin - once daily dosage | 10-20 mg/L * |
| *Trough levels of 15-20mg/L may be | required to treat deep seated |
| infections, please discuss this with the | e clinical Microbiology team |

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| Anti Cardiolipi | n Antibodies ACAB IgG and IgM |
|-------------------------------|---|
| Laboratory: | Haematology |
| Specimen: | Blood 3.5 mL red Vacuette [®] (Serum) |
| Comment: | Forms part of a Thrombophilia and/or Lupus screen, see Main Haematology |
| | Section on Guidelines for Investigation of Thrombophilia. Test available Mon |
| | to Fri during routine hours. |
| | This assay is only available when requested as part of Thrombophilia/Lupus investigations. |
| | Thrombophilia request form FOR-CUH-PAT-1575, including documentation |
| | of patient consent, must be received with all requests and is available on |
| | the CUH website. |
| Turnaround: | 3 - 4 weeks |
| Ref. Range: | IgG 0 - 10 GPL /mL |
| _ | IgM 0 - 7MPL /mL |
| Anti-CCP | |
| Laboratory: | Autoimmune Serology |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) |
| Comment: | Quantitative immunoassay using Phadia Immunocap 250 analyser. |
| T | Test restricted to consultant requests. |
| Turnaround: | 24 Hours |
| Ref. Range: Anti-c Quantit | 0 - 7 AU/mL |
| Laboratory: | Available by prior arrangement with Blood Transfusion Laboratory |
| Specimen: | $2 \times 6 \text{ mL EDTA Pink Capped Tube}$ |
| Comment: | Quantitations referred to: I.B.T.S., National Blood Centre, James's St., |
| Commenter | Dublin 8. |
| | Complete the Antenatal Serology request form LF-C-BTR-ANTENAT. |
| | Please note 3 forms of identification are required: Name, DOB and hospital |
| | number (address acceptable if none available) on both sample and form |
| | Please submit samples on Mondays if possible. |
| Turnaround: | 3 Weeks for Hard Copy reports. Verbal result from IBTS within 7 days. |
| Ref. Range: | Refer to IBTS report |
| Anti-D Quantit Laboratory: | Blood Transfusion Laboratory |
| Specimen: | 2 x 6 mL EDTA Pink Capped Tube |
| Comment: | Quantitations referred to: I.B.T.S., National Blood Centre, James's St., |
| | Dublin 8. |
| | Complete the Antenatal Serology request form LF-C-BTR-ANTENAT. |
| | Please note 3 forms of identification are required: Name, DOB and hospital |
| | number (address acceptable if none available) on both sample and form. |
| Turnaround: | 3 Weeks for Hard Copy reports. Verbal result from IBTS within 7 days. |
| Ref. Range: | Refer to IBTS report |
| | says (Voriconazole, Posaconazole) |
| Laboratory: | Microbiology |
| Specimen: Comment: | 4 ml Clotted serum sample, EDTA not suitable This test is performed in a reference laboratory, Mycology Reference Centre, |
| comment. | Bristol |
| Turnaround: | 5 working days |
| Report: | Numeric level in mg/L |
| | Antibody Testing (Paraneoplastic Antibodies) |
| Laboratory: | Neuropathology Department |

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| | | | | | |
| Specimen: | 4.0 ml of clotted blo | ood (red top vad | cuette) | | |
| Turnaround: | Approximately 2 weeks. | | | | |
| Anti Neutrophi | l Cytoplasmic Antil | bodies | | | |
| Laboratory: | Autoimmune Serolo | ypy | | | |
| Specimen: | Blood, 4 mL red top | Vacuette (or si | imilar container fo | or clotted blood) | |
| Comment: | Immunofluorescence assay using Ethanol + Formalin fixed human | | | | |
| | Neutrophils as Subs | | | | |
| | against Proteinase | ainst Proteinase 3 (PR3) and Myeloperoxidase (MPO) are automati | | | |
| | undertaken on sera | showing associ | ated positive imm | nunofluorescent | |
| | patterns. | | | | |
| | Anti-PR3 and Anti-N | | | | |
| | | • | unofluorescence A | NCA's on the Phadia | |
| | Immunocap 250 an | | | | |
| | For stat PR3 and MI | PO testing pleas | se contact lab dire | ectly. | |
| Turnaround: | 24 Hours | | | | |
| Ref. Range: | Not applicable | | | · · · · | |
| | | | | immune Neutropeni | |
| Laboratory: | Referred from Haer | • / | SBT Centre, Bristo | | |
| Specimen: | Clotted specimen a | | | | |
| Comment: | | | ansport within 24 | hours, complete form | |
| Turnerusundu | from referral labora | itory | | | |
| Turnaround: | 64 working days Sent to referring clinician and copy filed in laboratory | | | | |
| Report: | - | | | У | |
| Anti Nuclear Fa | | | | | |
| Laboratory: | Autoimmune Serolo | | imilar containor f | ar clatted blood) | |
| Specimen: Comment: | Blood, 4 mL red top Part of Autoantibod | | | | |
| Turnaround: | 24 Hours | y Screen. Fatter | in reported. Intre | not reported. | |
| Ref. Range: | Not applicable | | | | |
| | ntibody Investigat | ion | | | |
| | | | | | |
| Laboratory: Specimen: | Blood Transfusion L 3 mL Clotted (Red (| • | Ding) Tubo | | |
| Comment: | • | ••• | • / | e, James's St., Dublin 8 | |
| comment. | Complete the Blood | | | e, James S St., Dubin o | |
| Turnaround: | 3 Weeks | | quest form. | | |
| Ref. Range: | Not Applicable | | | | |
| | ntibodies (CAR ant | igon / Anti-roc | warin antibadia | | |
| | | | | | |
| Laboratory: | Sample referred fro | m Neuropathold | gy Department to | D Euronns-biomnis | |
| Chasiman | Laboratories Lyon | and (rad tap you | | | |
| Specimen: Turnaround: | 1.0 ml of clotted ble | • • | | a cont to referral lab) | |
| Ref. Range: | | • | - | s sent to referral lab) fins.ie/biomnis/ for up to | |
| Kel. Kaliye. | date referral test in | | te mups.//www.euro | | |
| | | TOTTIALION | | | |
| | sin-O Titre (ASOT) | | | | |
| Laboratory: | Microbiology (Infect | tious Diseases S | Serology) | | |
| Specimen: | 4mL clotted blood | | | | |
| Turnaround: | 36 hours | | | | |
| Report: | Titre provided (IU/r | mL) | | | |
| | | | | | |
| Comment: Anti Thrombin | | ndicate acute str | reptococcal infecti | ion | |

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| Specimen: | Blood 3mL blue Vacuette® (sodium citrate 3.2%) |
|-----------|--|
| | (Specimens, which are haemolysed, underfilled or overfilled, cannot be |
| | analysed, check coagulation sample bottles are not expired to ensure |
| | correct filling) |
| Comment: | Forms part of a Thrombophilia Screen. |
| | See Main Haematology Section on Guidelines for Investigation of |
| | Thrombophilia. Samples must be received within 4 hours |

Thrombophilia request form FOR-CUH-PAT-1575, including documentation of patient consent, must be received with all requests and is available on the CUH website.

Turnaround: 3 – 4 weeks

| Ref. F | Range: | Age | Range (%) |
|--------|--------|---------|-----------|
| | | Day 1 | 39- 87 |
| | | Day 5 | 41 - 93 |
| | | Day 30 | 48 - 108 |
| | | Day 90 | 73 – 121 |
| | | Day 180 | 84 - 124 |
| | | Adult | 80 - 120 |

Apixaban

See DOAC's- Direct Orla Anti-coagulants.

Ascitic Fluid

See Sterile Body Fluid – Microscopy and Culture or Cytology

Aspartate amino Transferase (AST)

| Laboratory: | Clinical Biochemistry |
|----------------|---|
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Comment: | Haemolysis invalidates result |
| Turnaround: | A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF, SMOH, MGH: - 3 hours approx. GP or OPD- Results posted within 4 days. |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate |
| Aspergillus An | tibodies |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Comment: | Performed by a reference laboratory (Mycology Reference Centre, Leeds) |
| | |

Turnaround: 28 working days

Report: Quantitative result with an interpretative comment

Aspergillus Antigen (Glactomannan)

| Laboratory: | Microbiology (Main Lab) |
|-------------|---|
| Specimen: | Bronchial lavage (Sputum samples unsuitable for testing) |
| Comment: | Performed by a reference laboratory (Mycology reference laboratory, Bristol) |
| Turnaround: | 28 working days |
| Report: | Negative or Positive with Titre |

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| Astrovirus | |
|-----------------------|---|
| Laboratory: | Microbiology (Category 3 Laboratory) |
| Specimen: | A fresh liquid faeces specimen is essential. 1-2mL is sufficient. |
| Comment: | Test not routinely available. Test seasonally available in-house, otherwise, |
| connenti | test will be referred to external laboratory. Please discuss with the |
| | Microbiology Medical team if required. |
| | A Target Not Detected result does not automatically exclude infection from |
| | the above enteric pathogen as the level of DNA present may be lower than |
| | the limit of detection of the assay. |
| | the limit of detection of the assay. |
| Turnaround: | In-house: 5 working days; External referral: 2 weeks. |
| Report: | Target Detected or Target Not Detected for Astrovirus. |
| Autoantibody S | |
| Laboratory: | Autoimmune Serology |
| | |
| Specimen: Comment: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) Includes: Anti Nuclear Factor +/- Anti-dsDNA and Extractable Nuclear |
| comment. | Antigen if ANF Positive + Anti-Mitochondrial, Anti Smooth Muscle and Anti- |
| | Gastric Parietal Cell Antibodies |
| Turnaround: | 24 Hours |
| Ref. Range: | Not applicable |
| Autopsy (CNS | |
| Laboratory: | Neuropathology |
| Laboratory. | Coroner's cases and Consent Autopsy protocols are shared with |
| | Histopathology (see HISTOPATHOLOGY section), please contact the post- |
| | mortem room on 22525. For post-mortems on CNS disease cases, please |
| | contact the consultant Neuropathologist on duty (22520/22519). |
| | Examinations on high-risk, suspected prion disease cases are conducted in |
| | the CJD surveillance centre in Beaumont Hospital, contact 01-8377755 |
| Turnaround: | 6-8 weeks |
| Avian Antibodi | es |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Comment: | Performed by a reference laboratory (Mycology Reference Centre, Leeds) |
| Turnaround: | 28 working days |
| Report: | Quantitative result with an interpretative comment |
| | e.g. fluconazole, itraconazole, isavuconazole etc) |
| Laboratory:M | |
| Laboratory.in | |
| | Blood: 4.0 mL blood in a plain tube (clotted sample) – Clotted samples with |
| | a gel plug are unsuitable. |
| | Performed by a reference laboratory (Mycology Reference Laboratory |
| Barbiturates | Southmead Hosp Bristol UK) |
| | Sample referred from Clinical Rischamistry to Tayicalagy Laboratory |
| Laboratory: | Sample referred from Clinical Biochemistry to Toxicology Laboratory |
| | BEAUMONT Hospital Dublin, posted Monday, Tuesday, Wednesday and |
| Speciment | Thursday. |
| Specimen: | Blood: 4.0 mL blood in a plain tube (clotted sample). Urine: spot urine |
| Comment: | See Toxicology / Drug Screen |
| Turnaround: | 1 week |
| Ref. Range: | See report form or contact Toxicology Laboratory BEAUMONT Hospital 01- |
| | 8092673 / (01)8092675, Emergency after hours (087) 2590749, Fax (01) 8093986 |
| | 0052500 |
| | |

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| Bartholin's Ab | scess |
|----------------|--|
| Laboratory: | Microbiology (Main laboratory) |
| Specimen: | Aspirate using a syringe (ideally a minimum of 1mL) or using a sterile swab. |
| • | Note: Do not send needle. |
| | Specimens should be taken before antimicrobial therapy where possible. |
| | The volume of specimen influences the transport time that is acceptable. |
| | Larger volumes of purulent material maintain the viability of anaerobes for |
| | longer. Transport ASAP in charcoal containing transport media. The viability |
| | of <i>N. gonorrhoeae</i> is lost over time. |
| Comment: | Test performed routinely Monday to Friday 9-5pm or by urgent request. |
| Turnaround: | Prelim: 24 hours; Final: 72 hours |
| Report: | Microscopy report (aspirates only) on the presence or absence of |
| | Intracellular Gram-negative diplococci and WBCs. |
| | Culture report: Any clinically significant isolate with the appropriate |
| | sensitivities. |
| | adelphia Chromosome) |
| Laboratory: | Haematology referred to Cancer Molecular Diagnostics, CMD, St James Hospital Dublin |
| Specimen: | 3 x 3 mL purple Vacuette (EDTA) blood or bone marrow in 10mL RPMI. |
| | Available Mon to Thurs to reach the laboratory before 12 noon on the day of |
| | sampling |
| Comment: | BCR-ABL associated with Ph+ CML, Ph+ ALL |
| Turnaround: | 60 working days |
| Report: | Sent to referring clinician and copy filed in laboratory |
| Bence - Jones | protein |
| Laboratory: | Clinical Biochemistry (Immunology Laboratory) |
| Specimen: | 20 mL urine |
| Comment: | As of June 6th requests for BJP are limited to Haematology Consultant |
| | request only |
| Turnaround: | 4 Days |
| Ref. Range: | Should be NEGATIVE |
| Benzodiazepin | |
| Laboratory: | Sample referred from Clinical Biochemistry to Toxicology Laboratory |
| | BEAUMONT Hospital Dublin, posted Monday, Tuesday, Wednesday and |
| Creative area | Thursday. |
| Specimen: | Blood: 4.0 mL blood in a plain tube (clotted sample). Urine: spot urine |
| Comment: | See Toxicology / Drug Screen 1 week |
| Turnaround: | |
| Ref. Range: | See report form or contact Toxicology Laboratory BEAUMONT Hospital 01- 8092673 / (01)8092675, Emergency after hours (087) 2590749, Fax (01) |
| | 8093986 |
| Beta-1-3-gluca | |
| Laboratory: | Microbiology |
| Specimen: | Serum/BAL/CSF |
| Comment: | Sample must be sent to the laborsatory immediately post collection, if |
| Comment | sample is delayed it will be rejected. Sputum samples are unsuitable for |
| | processing. Test performed by Mycology Reference laboratory, Bristol |
| Turnaround: | 14 days |
| Ref. Range: | Negative, Positive with Titre |
| | |

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| | Author: | Mr Paul Cantwell | |

Beta 2 Glycoprotein 1 (Anti beta 2GP1) Laboratory: Haematology

| Laboratory: | Haematology |
|--|--|
| Specimen: | Blood 3.5 mL red Vacuette [®] (Serum) |
| Comment: | Forms part of the Lupus or Thrombophilia Screen. |
| | This assay is only available when requested as part of Thrombophilia |
| | investigations. |
| | Thrombophilia request form FOR-CUH-PAT-1575, including documentation of |
| | patient consent, must be received with all requests and is available on the |
| | CUH website. |
| Turnaround: | 4-6 weeks |
| Ref. Range: | IgG Normal: < 5U/mL |
| Ren Ranger | Borderline: 5-8U/mL |
| | Elevated: >8U/mL |
| Beta-2-Microg | |
| | |
| Laboratory: | Sample referred from Clinical Biochemistry to Eurofins-Biomnis Laboratories |
| Specimen: | 4.0 mL blood in a plain tube (clotted sample) |
| Comment: | Consultant request only |
| Turnaround: | 2 weeks |
| Ref. Range: | See report form, or visit internet site https://www.eurofins.ie/biomnis/ for up to |
| | date referral test information |
| Bicarbonate (F | |
| Laboratory: | Clinical Biochemistry |
| Specimen: | Fresh 4.0 mL blood in plain tube (clotted sample) |
| Turnaround: | A/E or urgent sample: - 1 hour 30 mins approx. |
| | CUH wards, CUMH, SI, SF, SMOH, MGH: - 3 hours approx. |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as |
| | appropriate |
| | |
| Bile Acids | |
| Bile Acids Laboratory: | Clinical Biochemistry |
| | Clinical Biochemistry 4.0 mL blood in a plain tube (clotted sample) |
| Laboratory: | • |
| Laboratory: Specimen: | 4.0 mL blood in a plain tube (clotted sample) |
| Laboratory: Specimen: Turnaround: | 4.0 mL blood in a plain tube (clotted sample) 2 days |
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| BK Virus Mole | cular | | |
|---------------|---|--|--|
| Laboratory: | Microbiology (Infectious Diseases Serology) | | |
| Specimen: | 4mL clotted blood, 4mL EDTA blood, urine | | |
| Comment: | Performed by a reference laboratory (National Virus Reference Laboratory | | |
| commenter | (NVRL), Dublin) | | |
| Turnaround: | 14 working days | | |
| Report: | Detected (viral load) or not detected | | |
| Blood Culture | | | |
| Laboratory: | Microbiology (Main laboratory) | | |
| Specimen: | The blood culture vials and instrument in use are the BACTEC fluorescentsystem (Becton-Dickinson & Co. Ltd). An exception is the investigation formycobacteria (see Mycobacteriology section). Blood culture vials should bekept at a cool room temperature in the wards (2-25°C). The number of vialsstored in each ward should be limited to their general usage and excessivestocks avoided. There is an expiry date on each vial and they should not beused after this date.Adults:Preferably, a volume of 8-10mL of specimen per vial.ChildrenUse paediatric vials – preferably, a volume of 1-3mL (the volume of blood should be no more than 1% of the patients total blood volume). No need for lytic/anaerobic vial unless clinically indicated.Note: Do not exceed the manufacturer's recommended maximum volume for each bottle. | | |
| Comment: | If blood for other tests such as blood gases or ESR is to be taken at the same venepuncture, the blood culture bottles should be inoculated first to avoid contamination. It is preferable to take blood for culture separately. Disinfect the skin at the venepuncture site with isopropyl alcohol and allow to dry. Disinfect the septum of the blood culture bottle with alcohol and allow to | | |
| | dry. For diagnosis of bacteraemia withdraw blood from a peripheral vein and divide the specimen equally among blood culture vials, ensuring that the needle is changed between bottles. If the patient has a central line or other vascular access site, it is often appropriate to take both central and peripheral blood cultures. | | |
| | For neonates consider the use of a single aerobic paediatric vial appropriate for small volumes of blood. | | |
| | If necrotising enterocolitis is suspected and sufficient blood is obtained, inoculate a paediatric and a lytic/anaerobic bottle. | | |
| | Indicate if specific organisms are sought e.g. causative organisms of infective endocarditis. Consider bone marrow aspirate rather than blood sample for the diagnosis of thyphoid fever_and brucella species. | | |
| | Blood cultures should be transported to the laboratory as soon as possible after venepuncture as delays can lead to false negative results. | | |
| | NB. Do not refrigerate or place on radiators, incubators or direct sunlight. The pneumatic tube can be utilised to transport plastic blood culture vials and is preferable to avoid unnecessary delays. | | |
| Turnaround: | Most organisms will be detected within 24-48 hours and normally blood | | |

Turnaround: Most organisms will be detected within 24-48 hours and normally blood cultures are incubated for 5 days, but this time may be extended e.g. 10

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| | | | | | | Ms Sinead Creagh | |
| | | | Author | | Paul Cantwell | | |
| Report: | slow growing | organi | sms. | | | narrow up to 21 days f report at 5 days if the | |
| Report. | blood culture | | | | s anu a miai | report at 5 days if the | |
| | | - | | as soon as a | vailable to th | e requesting area or | |
| | team. | | Shoneu | 15 50011 05 0 | valiable to th | e requesting area of | |
| lood Gas | ccani. | | | | | | |
| Laboratory: | Point of Care | Tectin | a | | | | |
| Specimen: | | | - | ial Vonouc) | or Li Hep cap | villany | |
| Specifien. | • | | • | | | try Lab only) | |
| Commont | • | | | | | | |
| Comment: | | | | | | pint of Care site. Ensur | |
| | • | • | • | | ysis. Blood G | as samples must NOT | |
| Turna a wax wa du | be sent via p | neuma | lic shule | system. | | | |
| Turnaround: | 15 Minutes | | | | | | |
| | | 240 | | | | | |
| Sample | 35 µl -RL1 | | | Gas (pH, pC | | | |
| Volume: | 100µl-RP5 | | | Gas (pH, pC | | | |
| | 100µI-RP5 | | | | olytes (pH, po | CO ₂ , pO ₂ , Na ⁺ , K ⁺ , | |
| | | 00- | | a ⁺⁺ , Cl ⁻) | | | |
| | 100μι-κρ5 | | | bod Gas, Electrolytes & Metabolites (pH, pCO ₂ , pO ₂ , | | | |
| | | | | Na ⁺ , K ⁺ , Ca ⁺⁺ , Cl ^{-,} Glucose, Lactate) | | | |
| | 100µl-RP500e | | Blood Gas, Electrolytes, Metabolites & Co-Ox (pH, | | | | |
| | pCO ₂ , pO ₂ , Na ⁺ , K ⁺ , Ca ⁺⁺ , Cl ^{-,} Glucose, Lactate, tHb, sO ₂ , FO ₂ Hb, FCOHb, FMetHb, FHHb, Hct, Bilirubin) | | | | | | |
| | Parameter | Arteria | | Venous ⁽²⁾ | | I Capillary ⁽²⁾ | |
| Dof Dongo | | | | | | | |
| Ref. Range: | pH H ⁺ | 7.320- | | 7.32 -7.45 | 7.31 - 7. 48 48.98 -33 | | |
| | pCO _{2 (kPa)} | 47.9-3 | | 5.19 - 7.33 | | | |
| | pCO _{2 (kPa)} | 11.07- | | 3.99 - 7.33 | | | |
| | Na ⁺ (mmol/L) | 136 - | | 5.55 7.55 | 4.55 0.1 | | |
| | K ⁺ (mmol/L) | 3.4 - 4 | | + | | | |
| | Cl ⁻ (mmol/L) | 98 - 10 | | + | | | |
| | iCa ⁺⁺ (mmol/L) | 1.15 - | | 1 | 1.06 - 1. | 34 | |
| | Glu (mmol/L) | 3.6 - 5 | | 1 | 2.1 - 5.3 | | |
| | Lac (mmol/L) | 0.36-1 | | 0.56-1.39 | 1.4 - 4.1 | | |
| | Bicarb (mmol/L) | 19-24 | | 22-26 | | | |
| | tHb (g/dL) | 12.0-1 | L7.5 | 1 | 14.5 - 23 | 3.4 | |
| | Hct _{(c) (%)} | 35 - 51 | | 1 | | | |
| | O ₂ Hb (%) | 94.0 - | 98.0 | 1 | | | |
| | COHb | 0.5 - 1 | .5 | | | | |
| | MetHb | 0.0-1.5 | 5 | | | | |
| | HHb (%) | 0.0 - 5 | .0 | | | | |
| | sO _{2 (%)} | 95 - 98 | 3.0 | ~75 | | | |
| | HCO3 ⁻ (c) | 22.0 - | 26.0 | 21 - 30 | | | |
| | (mmol/L) | | | | | | |
| | BEecf | -2.0 - 2 | 2.5 | -3.0 - +3.0 | | | |
| | (mmol/L) | | | ļ | | | |
| | nBilirubin | | | | | $.8 (Neonate < 1 day)^{(1)}$ | |
| | (umol/L) | | | | | 205 (Neonate 1-2 days) ⁽¹ 3.6 (Neonate 3-5 days) ⁽¹ | |
| | | | | | | $\leq \mathbf{b} (\mathbf{N} \mathbf{O} \mathbf{O} \mathbf{D} \mathbf{D} \mathbf{T} \mathbf{O} \mathbf{A} \mathbf{A} \mathbf{B} \mathbf{O} \mathbf{O} \mathbf{O} \mathbf{A}$ | |
| | | | | | | 2 (adult) ⁽¹⁾ | |

D. Bruns; Elsevier Saunders, 2015.
Tietz NW, *Tietz Fundamentals of Clinical Chemistry and Molecular Diagnostics*, 4th Edition, C. Burtis and D. Bruns; Elsevier Saunders, 1995.

3. Reference range for serum sample for direct sodium measurement is as per arterial range

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| Blood Group a | nd Coombs | | | | |
|---------------|--|--|--|--|--|
| Laboratory: | Blood Transfusion Laboratory | | | | |
| Specimen: | 1 x 6 ml EDTA Pink Capped Tube | | | | |
| • | For Newborns: Cord Blood Sample in 6 ml EDTA Pink Capped Tube. | | | | |
| | For Paediatrics: 1 ml EDTA (Purple Cap/White Ring) Paediatric Bottle. | | | | |
| Comment: | Consists of Blood Group and Direct Coombs Test. Usually performed on | | | | |
| | Newborns. | | | | |
| | Complete the Blood Transfusion request form LF-C-BTR-BBCORD or | | | | |
| | LF-C-BTR-XMATCH. | | | | |
| | Blood Group and Direct Coombs Test are INAB Accredited tests. | | | | |
| Turnaround: | 24 hours. (Note: may be shortened to 1 hour in emergency) | | | | |
| Ref. Range: | Not Applicable | | | | |
| | nd Crossmatch | | | | |
| Laboratory: | Blood Transfusion Laboratory | | | | |
| Specimen: | 1 x 6 ml EDTA Pink Capped Tube | | | | |
| | For Paediatrics: 1 ml EDTA (Purple Cap/White Ring) Paediatric Bottle. | | | | |
| | Note: May require sample from mother of infant for crossmatching: 6 ml | | | | |
| | EDTA Pink Capped Tube | | | | |
| Comment: | Samples for crossmatching for elective surgery must arrive in the laboratory | | | | |
| | before 5 p.m. on day before surgery to avoid undue delay. Blood is | | | | |
| | crossmatched in batches and in accordance with the locally agreed Maximum | | | | |
| | Surgical Blood Ordering Schedule (MSBOS), except in exceptional cases. | | | | |
| | Arrangements are in place for the emergency issue of blood. In exceptional | | | | |
| | circumstances, blood may be issued uncrossmatched on request. | | | | |
| | Complete the Blood Transfusion request form LF-C-BTR-XMATCH. | | | | |
| | The laboratory accepts "Add-On" requests for additional units to be | | | | |
| | crossmatched when appropriate. These requests must be accompanied with | | | | |
| | a completed written Blood Product Requisistion Form LF-C-BTR-PROREQ. | | | | |
| | Crossmatch is an INAB accredited test. | | | | |
| Turnaround: | 3 Hours. (Note: The presence of irregular antibodies, or the need special | | | | |
| | requirements can lead to significant delays in efforts to obtain appropriate | | | | |
| | blood). | | | | |
| | Routine (non-urgent) samples will be processed during routine hours unless | | | | |
| | specified as an emergency. | | | | |
| | In emergencies the laboratory will attempt to provide crossmatched blood within 40 minutes to 1 hour (when pessible i.e. no artibadies) | | | | |
| | within 40 minutes to 1 hour (when possible i.e. no antibodies). | | | | |
| | These turnaround times apply to "Add On" requests for blood also. | | | | |
| | The Blood Transfusion Laboratory has introduced the ELECTRONIC ISSUE (EI) of red cell concetrates in Aug 2022. If a patient meets the parameters | | | | |
| | and once the Group & Hold has been processed, fully 'electronically | | | | |
| | crossmatched blood' may be issued in 5-10 minutes | | | | |
| Ref. Range: | Not Applicable | | | | |
| Blood Group a | | | | | |
| Laboratory: | Blood Transfusion Laboratory | | | | |
| Specimen: | 1 x 6 ml EDTA Pink Capped Tube | | | | |
| Specifien. | For Paediatrics: 1 ml EDTA (Purple Cap/White Ring) Paediatric Bottle. | | | | |
| Comment: | Blood is grouped and an antibody screen is performed. The sample is then | | | | |
| comment: | held in the laboratory for 72 hours @ 2-8°C. Blood may be crossmatched | | | | |
| | subsequently on that sample within 72 hours of collection. | | | | |
| | Complete the Blood Transfusion request form LF-C-BTR-XMATCH. | | | | |
| | Blood Group, Antibody Screen and Antibody Identification are INAB | | | | |
| | accredited tests. | | | | |
| | | | | | |

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| | | Author: | MI Faul Calitwell | | | |
| Turnaround: | 4 Hours. (Note: Group and hold samples are processed in batches in the laboratory. The presence of irregular antibodies can lead to significant delays in order to identify such antibodies). Routine (non-urgent) samples will be processed during routine hours un | | | | | |
| | | | | | | |
| | specified as an emer | <i>,</i> , | be processed durin | ig routine nours unless | | |
| | • | • • | attempt to comple | to the group and hold | | |
| Dof Dango: | within 40 minutes to Not applicable | | | te the group and hold tibodies). | | |
| Ref. Range: | | ligation | | | | |
| | sion Reaction Invest | | | | | |
| Laboratory: | Blood Transfusion La | • | | | | |
| Specimens: | 1 x 6 ml EDTA Pink (| • • | | | | |
| a . | 2 x 4ml clotted samp | • • • | - / | | | |
| Comment: | Complete the Blood | | | | | |
| | Tests may include Bl Crossmatch, Direct (accredited tests, | | | ng. These are all INAB | | |
| | | • | • | transfusion reaction is | | |
| | | • | | d on the last page of | | |
| | the Blood Compome | | | | | |
| | suspected Transfusio | • | | | | |
| Turnaround: | 4 Hours. | , | | | | |
| Ref. Range: | Not applicable | | | | | |
| | rain Natriuretic Pep | tide) | | | | |
| Laboratory: | Biochemistry | | | | | |
| Specimens: | 4.0 mL blood in a pl | ain tube (clotte | d sample) | | | |
| Comment: | Test performed rout | • | | by urgent request. | | |
| Turnaround: | 4 days | | | <i>b)</i> a gene equeen | | |
| Ref. Range: | See report form. | | | | | |
| | Examination (Haema | atology) | | | | |
| Laboratory: | Haematology | | | | | |
| Specimen: | Fresh bone marrow a | air-dried films | | | | |
| Specimen. | | | pencil with the nat | ient's name, MRN and | | |
| | DOB and sent to the | | | | | |
| Comment: | Examinations are un | • . | • | natients with | | |
| Commenter | leukaemia, anaemia | | 5 | • | | |
| | thrombocytopenia a | | | | | |
| Turnaround: | | | | pect a turn around time | | |
| | | | | ute leukaemia, ITP in a | | |
| | child, myeloma with renal failure. Such marrows will also have verbal results | | | | | |
| | phoned to requesting | g team the sam | e day. Other indic | ations can expect a | | |
| | TAT of up to two we | | | | | |
| | However significant | preliminary rep | orts will be phone | d by the reporting | | |
| | haematologist. | | | | | |
| Ref. Range: | Not applicable | | | | | |
| | tussis Antibodies | | | | | |
| Laboratory: | Microbiology (Infecti | ous Diseases S | erology) | | | |
| Specimen: | 4mL clotted blood | | | | | |
| Comment: | Test performed by a | | , , , , | - | | |
| _ | Preventable Bacteria | Reference Uni | t (RVPBRU), Londo | on) | | |
| Turnaround: | 28 working days | | | | | |
| | | | | | | |

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| | | | | | | | |
| Report: | Quantitative value w | • | | | | | |
| | vaccination, values > | | consistent with rec | cent infection. | | | |
| | cies Culture (Whoo | | | | | | |
| Laboratory: | Microbiology (Main la | | | | | | |
| Specimen: | Specialist collection according to local protocols. | | | | | | |
| | A Pernasal swab (Dacron [™] with flexible wire shaft) is inserted through a nostril and advanced along the floor of the nose until it reaches the | | | | | | |
| | | - | | | | | |
| | nasopharynx. It has posterior nasopharyr | | | | | | |
| | | • | | e to tolerate this for a | | | |
| | few seconds. | Rely that a path | che win only be ub | | | | |
| | Note: Cough plates a | and throat swat | os are unsatisfactor | rv and will not be | | | |
| | processed. | | | , | | | |
| | The laboratory must | be notified in a | dvance and transp | ort specimens ASAP. | | | |
| | B. pertussis is very s | usceptible to d | rying and is a very | slow grower, so | | | |
| | transport must keep | the organism r | noist and prevent o | overgrowth of normal | | | |
| | flora. Culture plates | • | | | | | |
| Comment: | Test performed routi | nely Monday to | Friday 9-5pm or b | oy urgent request. | | | |
| Turnaround: | 7 days | | | | | | |
| Report: | Bordetella pertussis | not isolated or | Bordetella pertussi | s / parapertussis | | | |
| Brain examina | isolated. tions (post mortem) |) | | | | | |
| Laboratory: | Neuropathology |) | | | | | |
| Specimen: | | / spinal cord | | | | | |
| Comment: | Formalin-fixed brain / spinal cord Post-mortem brain referrals are from Consultant Pathologists, please refer to | | | | | | |
| comment. | the protocol for brain referrals (Neuropathology Department Information | | | | | | |
| | Users). | | | | | | |
| Turnaround: | In general brain post | : mortem exam | inations are comple | eted within 3 months | | | |
| | although this does do | • | investigations perf | formed and the | | | |
| | complexity of the cas | | | | | | |
| | nolecular analysis for Pyrosequencing an | | | | | | |
| Laboratory: | | | | ry, Beaumont Hospital | | | |
| Specimen: | Brain tumour biopsy | | | ,, _ caamone noopical | | | |
| Comment: | Processed in Patholo | gy department | before referral. | | | | |
| Turnaround: | 4-6 weeks but may b | | | plexity | | | |
| BRCA gene tes | ting- Tumour | | | · | | | |
| Laboratory: R | eferred by Pathology t | o CMD, St. Jan | nes Hospital | | | | |
| Specimen: F | FPE tissue block | | | | | | |
| | est requests must be a | | | A test request and | | | |
| | onsent form available | | | • • • • • • • • | | | |
| | 3 working days (from | | | ferral institution) | | | |
| | Core Biopsy Calcifie | | | | | | |
| Laboratory: | See formalin fixed hi | | specimens | | | | |
| | al Brushings for PCI | | (Donal) (referred | to Primany Ciliany | | | |
| Laboratory: | Histopathology (Elec | | | | | | |
| Specimen: | Dyskinesia (PCD) Diagnostic Service, University Hospital Southampton) Bronchial and Nasal brushings in 3% glutaraldehyde. | | | | | | |
| Comment: | Contact the laborato | - | . , | or hy e-mail to | | | |
| comment. | arrange collection of | | | or by e mail to | | | |
| Turnaround: | 12 weeks | Ciacai ai activat | | | | | |
| . a. nar oundr | | | | | | | |

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| Bronchoalveol | ar Lavage Fluid Culture | | | |
|-------------------|---|--|--|--|
| Laboratory: | Microbiology (Main laboratory) | | | |
| Specimen: | Specialist collection according to local protocols. It is difficult to be specific on volume required; in principle as large a volume as possible is preferred (up to 30mL). | | | |
| | The specimen should be collected into a clean, sterile, leakproof container and transported to the laboratory ASAP. If processing is delayed, | | | |
| | refrigeration is preferable to storage at ambient temperature. Please include any appropriate clinical details e.g. "Cystic fibrosis patient". If an unusual pathogen is suspected, the laboratory should be informed, <i>e.g. Burkholderia</i> <i>pseudomallei</i> and <i>Nocardia</i> sp require longer incubation of cultures. Refer to Mycobacteria Testing for instructions for collection for TB. | | | |
| Comment: | Test performed routinely Monday to Friday 9-5pm or by urgent request. Traps containing a specimen should be properly sealed. Do not send tubing to the laboratory. | | | |
| Turnaround: | Prelim: 24 hours; Final: 48-72 hours | | | |
| Report: | Aerobic culture with sensitivities, if appropriate, as well as microscopy and culture for Mycobacteria. | | | |
| Brucella Antib | odies (IgG, IgM and Total) | | | |
| Laboratory: | Microbiology (Infectious Diseases Serology) | | | |
| Specimen: | 4mL clotted blood | | | |
| Turnaround: | 28 working days | | | |
| Report: | Quantitative titre provided with interpretative comment | | | |
| Comment: | Performed by a reference laboratory (Brucella Reference Unit (BRU), Liverpool). | | | |
| | Not routinely available, please contact Microbiology Medical Team. | | | |
| | A negative result generally excludes a diagnosis of brucellosis. Positive | | | |
| | Brucella agglutination reactions should be regarded as supportive evidence | | | |
| | for the diagnosis of brucellosis provided there is reasonable epidemiological | | | |
| | and clinical evidence to suggest the diagnosis. A rising or falling titre is more significant than a single titre. | | | |
| Bursa Fluid | | | | |
| See Sterile Bo | ody Fluid – Microscopy and Culture. | | | |
| C1 Esterase In | hibitor (Function) | | | |
| Laboratory: | Sample referred from Clinical Biochemistry to Eurofins-Biomnis Laboratories | | | |
| Specimen: | 4.0 mL blood in a plain tube (clotted sample) + 5 mL citrated whole blood on ice. | | | |
| Comment: | Consultant request only | | | |
| Turnaround: | 3 weeks | | | |
| Ref. Range: | See report form, or visit internet site <i>https://www.eurofins.ie/biomnis/</i> for up to date referral test information. | | | |
| C1 Esterase In | hibitor (Total) | | | |
| Laboratory; | Sample referred from Clinical Biochemistry to Eurofins-Biomnis Laboratories | | | |
| Specimen: | 4.0 mL blood in a plain tube (clotted sample | | | |
| Comment: | Consultant request only | | | |
| Turne a navina du | | | | |

Turnaround: 3 weeks

| | Jeniency |
|-------------|---|
| Laboratory: | Clinical Biochemistry (Immunology Laboratory) |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Turnaround: | 4 Days |
| | |

Ref. Range: See report form, or visit internet site https://www.eurofins.ie/biomnis/ for up to

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| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as |
|-------------|---|
| | appropriate |

| | appropriat | e | | | | | |
|----------------|---|-------------------|--------------|----------------------------|--------------|------------------|--|
| CD3 / CD4/ CD | 8 / CD19 | / CD56 Count | S | | | | |
| Laboratory: | Haematolo | gy | | | | | |
| Specimen: | Blood 3mL x 1, purple, Vacuette [®] (EDTA). | | | | | | |
| Comment: | A screening procedure to monitor the immune status of patients / clients. | | | | | | |
| | Test available Mon to Fri during routine working hours. | | | | | | |
| Turnaround: | 24 - 72 hours | | | | | | |
| Ref. Range | CD 3 Absolu | ite Counts / μL | CD4 Abso | ute Counts /µL | CD8 Abso | olute Counts /μL | |
| | Age | Low High | Age | Low High | Age | Low High | |
| | Day 6 | 900 - 5,000 | Day 6 | 500 - 3,400 | Day 6 | 300 - 1900 | |
| | Month 2 | 2,800 - 7,000 | Month 2 | 2,100 - 4,900 | Month 2 | 500 - 1600 | |
| | Year 2 | 1,600 - 6,700 | Year 2 | 1,000 - 4,600 | Year 2 | 400 - 2100 | |
| | Year 5 | 900 - 4,500 | Year 5 | 500 - 3,400 | Year 5 | 300 - 1600 | |
| | Year 10 | 700 - 4,200 | Year 10 | 400 - 2,000 | Year 10 | 300 - 1800 | |
| | Year 16 | 700 - 3,500 | Year 16 | 400 - 2,000 | Year 16 | 200 - 1200 | |
| l | Adult | 690 - 2,540 | Adult | 400 - 1,590 | Adult | 190 - 1140 | |
| r | | | | | | | |
| | | lute Counts / µL | | Absolute Counts / | μL | | |
| | Age | Low High | Age | Low High | | | |
| | Day 6 | 200 - 1,100 | Day 6 | 200 - 1,900 | | | |
| | Month 2 | 300 - 1,900 | Month 2 | 300 - 1,000 | | | |
| | Year 2 | 600 - 2,700 | Year 2 | 200 - 1,200 | | | |
| | Year 5 | 200 - 2,100 | Year 5 | 100 - 1,000 | | | |
| | Year 10 | 200 - 1,600 | Year 10 | 90 - 900 | | | |
| | Year 16 | 200 - 600 | Year 16 | 90 - 900 | | | |
| · | Adult | 90 - 660 | Adult | 90 - 590 | | | |
| C Peptide | | | | | | | |
| Laboratory: | Clinical Bio | chemistry | | | | | |
| Specimen: | | • | ibe (clotte | d sample) at 4° (| - | | |
| Comment: | | | | ailable on reque | | | |
| Turnaround: | 7 days | | ergente at | | | | |
| Ref. Range: | - | levels should b | e appropri | ate to the alucos | e level at t | he time the | |
| iter iteriger | C-peptide levels should be appropriate to the glucose level at the time the sample was taken. Glucose should always be measured at the same time as | | | | | | |
| | | ide to facilitate | | | | | |
| CA 125 | · · | | • | | | | |
| Laboratory: | Clinical Bio | chemistry | | | | | |
| Specimen: | | od in a plain tu | ibe (clotted | d sample) | | | |
| Turnaround: | 4 Days | | , | . , | | | |
| Ref. Range: | - | e reference inte | rvals will b | e applied to all I | Biochemist | rv reports as | |
| | appropriat | | | | | , | |
| CA 15-3 | | | | | | | |
| Laboratory: | Clinical Bio | chemistry | | | | | |
| , Specimen: | | od a plain tube | (clotted s | ample) | | | |
| Turnaround: | 4 days | | • | . , | | | |
| Ref. Range: | • | e reference inte | rvals will b | e applied to all I | Biochemist | ry reports as | |

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| CA 19-9 | | | | |
|----------------|---|--|--|--|
| Laboratory: | Clinical Biochemistry | | | |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) | | | |
| Turnaround: | 4 Days | | | |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate | | | |
| Calcitonin | appropriate | | | |
| Laboratory: | Sample referred from Clinical Biochemistry to Eurofins-Biomnis Laboratories | | | |
| Specimen: | 4.0 mL blood in a plain tube (clotted sample) on ice must be frozen < 4 hours. | | | |
| Comment: | Consultant request only | | | |
| Turnaround: | 3 weeks | | | |
| Ref. Range: | See report form, or visit internet site <i>https://www.eurofins.ie/biomnis/</i> for up to date referral test information. | | | |
| Calcium (Blood | | | | |
| Laboratory: | Clinical Biochemistry | | | |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) | | | |
| Comment: | Aged samples may invalidate result. | | | |
| Turnaround: | A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF, SMOH, MGH: - 3 hours approx. GP or OPD- Results posted within 4 days. | | | |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate Please contact Clinical Biochemistry lab for Paediatric and Pregnancy-related Reference ranges. | | | |
| Calcium (Urina | | | | |
| Laboratory: | Clinical Biochemistry | | | |
| Specimen: | 24 Hr acidified sample | | | |
| Turnaround: | 1 Day | | | |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as | | | |
| | appropriate | | | |
| Calcium: Creat | inine Clearance | | | |
| Laboratory: | Clinical Biochemistry | | | |
| Specimen: | Spot urine sample and clotted blood sample | | | |
| Turnaround: | 1 day | | | |
| Ref. Range: | Contact Biochemistry laboratory | | | |
| Calcium Sensin | g Receptor (CASR) Mutation analysis | | | |
| Laboratory: | Referred from Molecular Genetics Lab in Biochemistry to Oxford NHS (via NCMG) | | | |
| Specimen: | 3-5ml EDTA blood | | | |
| Comment: | Use NCMG request form with consent available from <u>www.genetics.ie</u> .Contact ext 22531 for Oxford Proforma. | | | |
| | Please note: invoices will be issued to the referring clinician for tests not performed in NCMG. | | | |
| Turnaround: | 8 weeks | | | |
| Report: | Sent to referring clinician and copy filed in pathology | | | |
| | lobulin (GLOB) | | | |
| Laboratory: | Clinical Biochemistry | | | |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) | | | |
| Comment: | Calculation involving the measurement of both Total Protein and Albumin on all patients >16 years | | | |
| Turnaround: | A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF, SMOH, MGH: - 3 hours approx. GP or OPD- Results posted within 4 days. | | | |

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| | Up-to-date reference intervals will be applied to all Biochemistry reports as |
|------------|---|
| Ref Range: | appropriate Please contact Clinical Biochemistry lab for Paediatric and |
| 2 | Pregnancy-related Reference ranges. |

Calprotectin

| Calprotectin | |
|--------------|---|
| Laboratory: | Referred from Biochemistry to City Hospital, Birmingham |
| Specimen: | 5-10mg stool |
| Comment: | Test helps distinguish IBD from IBS |
| Turnaround: | 2 weeks |
| Cannabis | |
| Laboratory: | Sample referred from Clinical Biochemistry to Toxicology Laboratory BEAUMONT Hospital Dublin, posted Monday, Tuesday, Wednesday and Thursday. |
| Specimen: | Spot urine |
| Comment: | See Toxicology / Drug Screen |
| Turnaround: | 1 week |
| Ref. Range: | See report form or contact Toxicology Laboratory BEAUMONT Hospital 01- 8092673 / (01)8092675, Emergency after hours (087) 2590749, Fax (01) 8093986 |

CAPD

See Continuous Ambulatory Peritoneal Dialysis Fluid

| occ continuo | ds Ambulatory i chtonear Dialysis i laid | | |
|---|--|--|--|
| Carbamazepin | e | | |
| Laboratory: | Clinical Biochemistry | | |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) | | |
| Comment: | Range quoted is appropriate for a trough sample. | | |
| Turnaround: | 1 Day | | |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as | | |
| | appropriate | | |
| Carbapenamas | se Producing Enterobacteriales | | |
| Laboratory: | Microbiology (Main laboratory) | | |
| Specimen: | Rectal swabs, placed in charcoal containing transport media. | | |
| Comment: | Test performed Monday to Friday 9-5pm. Label all Microbiology forms with | | |
| | CPE SCREEN. Indicate if the patient was previously CPE positive or CPE | | |
| | contact. Transport specimens ASAP. If processing of swabs is delayed, | | |
| | refrigeration is preferable to storage at ambient temperature. | | |
| Turnaround: | Prelim: 24 hours; Final: 48-72 hours. | | |
| | e Producing Enterobacteriales PCR | | |
| Laboratory: | Microbiology (Main laboratory) | | |
| Specimen: | Rectal swab, placed in PCR transport media. Contact Microbiology Laboratory | | |
| for appropriate sterile transport swabs. Specimens are only processed | | | |
| | where there is prior agreement with the Consultant Microbiologist or | | |
| a . | the Infection Control Team. | | |
| Comment: | Test performed Monday to Friday 9-5pm. Label all Microbiology forms with | | |
| | CPE SCREEN. Indicate if the patient was previously CPE positive or CPE contact. Transport specimens ASAP. If processing of swabs is delayed, | | |
| | refrigeration is preferable to storage at ambient temperature. | | |
| Turnaround: | Final Result: 24 hours. | | |
| | | | |
| Carboxyhaemo | | | |
| Laboratory: | Clinical Biochemistry | | |
| Specimen: | Li Hep syringe | | |
| Turnaround: | 1 hour 15 mins | | |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate | | |

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| Cardiothoracic | specimens |
|-----------------|---|
| Laboratory: | See formalin fixed histolopathology specimens |
| Carnitine, Free | & Total |
| Laboratory: | Sample referred from Clinical Biochemistry to Sheffield Children's NHS Trust |
| Specimen: | 1.0 mL blood in a plain tube (clotted sample) or Lithium Heparin sample on |
| | ice, must be frozen < 30 mins. |
| Comment: | Consultant request only |
| Turnaround: | 3 weeks |
| Ref. Range: | See report form |
| Catecholamine | s – Urine |
| Laboratory: | Sample referred to from Clinical Biochemistry to Beaumont hospital |
| Specimen: | 24-hour urine sample collected into a container that has acid added. |
| • | 24 hr urine containers are available from stores; acid is added in the |
| | Biochemistry lab. |
| Comment: | Diet must NOT include bananas, chocolate, tomatoes, citrus fruits, walnuts, |
| | pineapple, plums, dried fruit, tea or coffee in the 48 hours before collection |
| Turnaround: | 4 weeks |
| Ref. Range: | Contact CUH Biochemistry Laboratory |
| Catheter / Inti | avascular Cannulae |
| Laboratory: | Microbiology (Main laboratory) |
| Specimen: | Disinfect the skin around the cannula entry site, remove cannula using |
| • | aseptic technique, and cut off 4cm of the tip into a sterile container using |
| | sterile scissors. The specimen should be collected into a clean, sterile, |
| | leakproof container and should be transported ASAP to prevent drying. If |
| | processing is delayed, refrigeration is preferable to storage at ambient |
| | temperature. |
| Comment: | Not routinely processed, if required please contact the medical team. If |
| | infection considered clinically likely please take blood cultures through the |
| | cannula. |
| | The routine culture of devices removed for other reasons is unnecessary. |
| | Urine catheters are not cultured since growth represents distal urethral |
| | culture. A urine specimen is more appropriate. Skin disinfection procedures |
| Turrenaria | depend on local protocols and may vary. |
| Turnaround: | Prelim: 24 hours; |
| Def Denser | Final: 48-72 hours |
| Ref. Range: | Culture: Any clinically significant isolate with the appropriate sensitivities. |
| CEA | |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Turnaround: | 4 Days |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as |
| | appropriate |
| Centromere B | |
| Laboratory: | Autoimmune Serology |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) |
| Comment: | Qualitative Elisa assay. Specific assay undertaken following Positive Anti ENA |
| _ . | Screen. |
| Turnaround: | 72 Hours |
| Ref. Range: | Not applicable |

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Cerebrospinal Fluid (CSF) – Biomarkers (Amyloid, Tau)

| Laboratory: | Referred from the Immunology Dept CUH to Immunology Dept, St James's |
|-------------|--|
| | Hospital, Dublin 8. |
| Specimen: | 2.5 mL CSF specimen collected in to a polypropylene tube. Sample must be |
| | centrifuged within 2 hours of collection |
| Comment: | Polypropylene tubes are available from the Immunology Lab, ext 22535. |
| Turnaround: | Contact the Immunology Dept, St James's Hospital, Dublin 8, ph 01- |
| | 4162925 |
| Pof Pango | Contact St lames's for interpretation $nh 01-4162025$ |

| Ref. Range: | Contact St James's for interpretation ph 01-4162925 | | | |
|---------------|--|--|--|--|
| Cerebrospinal | Cerebrospinal Fluid (CSF) - Culture and Microscopy | | | |
| Laboratory: | Microbiology (Main laboratory) | | | |
| Specimen: | Ideally, the laboratory should receive a minimum volume of 1mL in a universal container AND SHOULD BE SAMPLE NUMBERS 1 AND 3 . The specimen should be collected into a clean, sterile, leakproof container. | | | |
| | Information regarding suspected Prion disease MUST be indicated on the request form; the CSF MUST be double-bagged and marked with a biohazard label. | | | |
| | For Mycobacteria, as large a volume as possible should be sent (given the patient's clinical circumstances). All specimens should be taken before antimicrobial therapy where possible, but therapy should not be delayed unnecessarily pending lumbar puncture. | | | |
| Comment: | Test performed as an urgent specimen. Do not refrigerate specimen. Do not send through the pneumatic tube. CSF is normally collected sequentially into separate containers. Common practice is to send the first and third specimens taken for microbiological examination and the second specimen for Biochemistry. If only one specimen of CSF is collected, it should be submitted to Microbiology first. Transport specimens ASAP directly to the laboratory. Do not refrigerate samples if delays in transportation are encountered. Cells disintegrate and a delay may produce a cell count that does not reflect the clinical situation of the patient. Prior notification to the laboratory in cases of suspected CJD /vCJD. | | | |
| | CSF, EDTA blood specimens may be sent to the Meningococcal Reference Laboratory for PCR. All isolates of <i>N. meningitidis</i> are referred for serotyping. All lymphocytic CSEs (WBCs $>5/cmm$) are routinely sent for Mycobacterial | | | |

Laboratory for PCR. All isolates of *N. meningitidis* are referred for serotyping. All lymphocytic CSFs (WBCs >5/cmm) are routinely sent for Mycobacterial testing. With lymphocytic CSFs consideration should be given to other tests such as Viral PCR (CMV, HSV and VZV). With a culture negative lymphocytic CSF, a clearly labelled stool specimen for enteroviral investigation should be considered.

CSF samples which have an elevated white cell count as detailed below with the exception of shunts and CSF samples from Haematology and Neurology patients, these are internally reflexed to the Biofire FA/ME panel where requested by clinical team using green Microbiology form. CSF samples with normal white cell count that require virology investigation refer to Section CSF Viral screen or for meningococcal investigation See *Neisseria meningitidis* PCR or meningococcal PCR sections

As the CSF specimen volume is limited, it is worth doing serology for antibodies to viral agents. The CNS Screen includes Mumps, Measles, Herpes Simplex and Varicella-zoster. Likewise serology for systemic syndromes associated with meningoencephalitis such as HIV, syphilis and Lyme Disease should be considered. If the patient is immunosuppressed Cryptococcal meningitis should be considered.

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| | F | Approved By: | Dr Vitaliy Mykyti | v, Ms Sinead Creagh | |
| | | Author: | Mr Paul Cantwell | | |
| Turnaround: | when available. | | | tive report telephoned ture may be prolonged fo | |
| | fungal investigation i | f required (up | to 14 days) | | |
| | Where Biofire is perfo | ormed (cases | where White ce | ll count is elevated) resu | |
| | | | | nay take longer. Positive | |
| | results will be phoned | | | | |
| Report: | Report on the gross a | appearance of | the CSF, the pr | esence of a clot if | |
| | applicable. | the numbers | of MDCo/omm | and DBCalamm | |
| | Microscopic report on | | or wbcs/cmm | and RBCS/cmm. | |
| | Normal CSF cell co | | | 0.20 colle/omm | |
| | Leucocytes | | 28 days old | 0-30 cells/cmm | |
| | | Infants 1-1 | | 0-15 cells/cmm | |
| | Em throattac | | lult > 1year | 0-5 cells/cmm | |
| | Erythrocytes A WBC: RBC ratio of | | | t in a normal CSF | |
| | infection | 1:500 is gen | erally regarded | as not indicative of | |
| | | ormed on all (| SE specimens y | with a white cell count | |
| | A Gram stain is performed on all CSF specimens with a white cell count above the normal range as indicated above. | | | | |
| | A differential leucotye count is reported where sufficient cells are counted \geq | | | | |
| | 20 WBC s/cmm. Cell counts <20 WBC/cmm the predominating WBC will be | | | | |
| | reported with comment insufficient WBC for accurate differential. Cell counts | | | | |
| | are not performed on specimens containing a clot, which would invalidate the | | | | |
| | cell count. | | | | |
| | Culture: Any organis | | h the appropria | te sensitivity results. | |
| | Fluid (CSF) - Cytolog | | (= | | |
| Laboratory: | Neuropathology or Hi | | | - | |
| Specimen: | | | | 3ml. and be collected in | |
| Commont | | | | e laboratory before 4pm | |
| Comment: | | - | • | re is delay in sending the | |
| | sample to the laboratory it should be stored at 4°C. Samples from patients with suspected CJD should be sent to | | | | |
| | Neuropathology and not Cytopathology. | | | | |
| | Information regarding suspected Prion disease MUST be indicated on the | | | | |
| | request form. | 5 1 | | | |
| Turnaround: | 2 days | | | | |
| Ref. Range: | Not applicable | | | | |
| CSF Pyridox | al Phosphate | | | | |
| Laboratory | Referred from Clinica | l Biochemistry | to the Nationa | l Hospital for Neural and | |
| | Neurosurgery | | | | |
| Specimen | 1.5 mL CSF specimer | า | | | |
| Turnaround | 6 weeks | | | | |
| Ref. Range: | See report | | | | |
| Cerebrospinal | Fluid (CSF) – Glucos | е | | | |
| Laboratory: | Clinical Biochemistry | | | | |
| | | _ | | | |
| Specimen: | 1.5 mL CSF specimer | | | | |
| Specimen: Comment: | 1.5 mL CSF specimer Fresh sample require glucose bottle. | | sample should | be kept in paediatric | |
| • | Fresh sample require | | sample should | be kept in paediatric | |

| Title: Laboratory Medicine User Handbook | Reference: | PPG-CUH-PAT-31 | Revision: 22 |
|--|--------------|------------------------|-----------------|
| | Active Date: | 03/11/2023 | Page: 94 of 206 |
| | Approved By: | Dr Vitaliy Mykytiv, Ms | Sinead Creagh |
| | Author: | Mr Paul Cantwell | |

Cerebrospinal Fluid (CSF) – Immunophenotyping - primary CNS lymphoma or CNS involvement by Leukaemia/ lymphoma

| Specimen: RPMI-heparin medium is stored in the haematology Dunmanway once the CSF is added the samples are to be sent directly to the haematology laboratory. Comment: Test available Monday- Friday during routine working hours CSF immunophenotyping is for diagnosis of primary CNS lymphoma on involvement by Leukaemia/ lymphoma only. Samples from patien non haematological diagnoses will not be tested. CSF samples for cytometry must be taken directly into RPMI-heparin. CSF samples for cytometry must be taken directly into RPMI-heparin. CSF samples for cytometry must be taken directly into RPMI-heparin and w processed if greater than 1 hour old irrespective of Microbiology o cell counts Turnaround: 40 working days Report: Sent to referring clinician and copy filed in laboratory Cerebrospinal Fluid (CSF) – Neurotransmitters Laboratory: Laboratory: Referred from the Immunology Dept, CUH to the Neurometabolic Queens Square, London, pet collected from Immunolgy laboratory, (ext 22535) Comment: It is essential to contact the laboratory prior to collection to ensur availability of dry ice. Turnaround: Contact Neurometabolic unit, Queens Square, London, ph 00-44-83844 Cerebrospinal Fluid (CSF) – Oligoclonal bands Eaboratory: Laboratory: Sample referred from Clinical Biochemistry to Eurofins-Biomnis La 83844 Cerebrospinal Fluid (CSF) – Oligoclonal bands Eaboratory: Laboratory: Clinical Biochemistry Specimen: | RPMI-heparin medium is stored in the haematology Dunmanway day unit, once the CSF is added the samples are to be sent directly to the haematology laboratory. Test available Monday- Friday during routine working hours CSF immunophenotyping is for diagnosis of primary CNS lymphoma or CNS involvement by Leukaemia/ lymphoma only. Samples from patients with non haematological diagnoses will not be tested. CSF samples for flow cytometry must be taken directly into RPMI-heparin. CSF samples are extremely labile and samples not received in RPMI-heparin and will not be processed if greater than 1 hour old irrespective of Microbiology or Cytology cell counts 40 working days Sent to referring clinician and copy filed in laboratory uid (CSF) – Neurotransmitters Referred from the Immunology Dept, CUH to the Neurometabolic unit, Queens Square, London, Contact laboratory prior to specimen collection. CSF specimen containers to be collected from Immunolgy laboratory, (ext 22535) It is essential to contact the laboratory prior to collection to ensure the availability of dry ice. Contact Neurometabolic unit, Queens Square, London, ph 00-44-20-344- 83844 uid (CSF) – Oligocional bands Sample referred from Clinical Biochemistry to Eurofins-Biomnis Laboratories 0.5 mL CSF and 4.0 mL blood in plain tube (clotted sample) 3 weeks Oligocional Bands should be NEGATIVE uid (CSF) – Protein Clinical Biochemistry 1.5 mL CSF specimen Presence of blood in sample will affect results 1 hour 15 mins Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate uid (CSF) – RT-QUIC Neuropathology 2-3 mL clear CSF in a universal container. CSF should be transported as soon as possible to Neuropathology for freezing. If there is delay in sending the sample to the laboratory, it should be stored at 4°C. Details of storage conditions should be recorded on the form. The information regarding suspected Prion disease MUST be indicated on the request form. | Involvement D | |
|---|--|----------------|---|
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| Title: Laboratory Me | edicine User Handbook | Reference: | PPG-CUH-PAT-31 | Revision: 22 |
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| | | Active Date: | 03/11/2023 | Page: 95 of 206 |
| | | Approved By: Author: | Dr Vitaliy Mykytiv, M Mr Paul Cantwell | s Sinead Creagh |
| | | Author: | | |
| Comment: | Specimens are refer | red to the Irish | National CJD Surv | veillance Unit, |
| | Neuropathology Dep | | | |
| | | | | ce unit in Beaumont |
| | are available from the | | | |
| | • | | | interpret the results |
| | and must accompan | | | |
| Turnaround | Approx. 14 days from | | • | |
| | • • | • | . 2, | If a result is required |
| | more urgently pleas | e contact Neuro | opathology.) | |
| Cerebrospinal S | | | | |
| Laboratory: | Microbiology (Main I | | | |
| Specimen: | | | | sent concurrently for |
| | | | | ions should be sent in |
| | separate containers | | | he specimen should b |
| | collected into a clear | | | |
| | ASAP. If processing | | | |
| | ambient temperatur | | igeration is prefere | able to storage at |
| Comment: | Test performed rout | | o Friday 9-5pm or | bv uraent request. |
| Turnaround: | Prelim: 24 hours; | ,, . | ······, · · · · · | -, |
| | Final: 48-72 hours, culture may be prolonged for fungal /anaerobic | | | |
| | investigation if required (up to 5 days). | | | |
| Ref. Range: | If pus is clearly seer | • • | | |
| 5 | | | • | ficient CSF visible in |
| | the shunt tubing or reservoir the numbers of WBCs/cmm and RBCs/cmm | | | |
| | are reported. | | | |
| | Culture: Any clinical | ly significant is | olate with the app | opriate sensitivities. |
| Cerebrospinal I | -luid (CSF) – Specti | | (Xanthochromia | |
| Laboratory: | Clinical Biochemistry | | | |
| Specimen: | 1.0 mL CSF specime | | | |
| Comment: | Sample must be ligh | nt protected. Plo | ease use the speci | fic request form. |
| Turnaround: | 24 hours (weekdays | | | |
| Ref. Range: | Ring laboratory for i | nterpretation | | |
| Ceruloplasmin | | | | |
| Laboratory: | Clinical Biochemistry | | | |
| Specimen: | 4.0 mL blood in a pl | ain tube (clotte | d sample). | |
| Turnaround: | 4 Days | | | |
| Ref. Range: | Up-to-date reference | e intervals will | be applied to all Bi | ochemistry reports as |
| | appropriate | | | |
| Cervical Swab | for Microbiology | | | |
| Refer to Genital | swab | | | |
| Children gruppen A | ntihodies | | | |
| Chikungunya A | incidoutes | | | |
| Laboratory: | Microbiology (Infecti | ious Diseases S | erology) | |
| | | ious Diseases S | erology) | |
| Laboratory: | Microbiology (Infecti 4mL clotted blood | | | Reference Laboratory |
| Laboratory: Specimen: | Microbiology (Infecti 4mL clotted blood | | | Reference Laboratory |
| Laboratory: Specimen: | Microbiology (Infecti 4mL clotted blood Performed by a refe | | | Reference Laboratory |

| Title: Laboratory Medicine User Handbook | Reference: | PPG-CUH-PAT-31 | Revision: 22 |
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| | Approved By: | Dr Vitaliy Mykytiv, Ms | Sinead Creagh |
| | Author: | Mr Paul Cantwell | |

| Chlamydia trac | chomatis |
|------------------------|--|
| Laboratory: | Microbiology |
| Specimen: | Nucleic acid amplification method. Appropriate PCR STD Specimen Collection and Transport Kits must be used. Please read the kit insert for information on specimen collection and associated limitations. |
| Comment: | Test available Monday to Friday 9-5pm. |
| | The assay is verified for use with female Endocervical swab specimens, High Vaginal Swab specimens and male/female Urine specimens. (These specimens will also be tested for <i>Neisseria gonorrhoea</i> DNA). |
| | The preferred specimen type for Chlamydia testing in female patients is urine due to increased sensitivity and fewer problems during specimen processing. |
| | Underfilled or overfilled Urine specimen containers are unsuitable for testing. |
| | Endocervical/HVS specimen tubes with no swab or with two swabs cannot be tested. |
| | Use only flocked swabs for Endocervical sampling (this is the thinner of the 2 swabs in the sample collection kit). Woven swabs from Endocervical sites are not processed. |
| | Use woven swabs provided for all other sites, other than Endocervical sites Specimens that appear bloody or have a dark brown colour are unsuitable for testing (may give false negative results). |
| | The presence of mucous may inhibit PCR and cause false negative test results. Mucous free specimens are required for optimal test performance. Do not use collection devices beyond their expiry date. |
| Turnaround: Report: | 96 - 120 hours RT: PCR <i>Chlamydia trachomatis</i> Target Not Detected or Target Detected A Target Not Detected result does not automatically exclude infection from Chlamydia trachomatis as the level of DNA present may be lower than the limit of detection of the assay. |
| | The assay is only verified for use with female Endocervical/HVS swab specimens and male/female Urine specimens. Results from other specimen types should be interpreted with caution. |
| Chloride (Bloo | d) |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Turnaround: | A/E or urgent sample: - 1 hour 30 mins approx. CUH wards, CUMH, SI, SF, SMOH, MGH: - 3 hours approx. GP or OPD- Results posted within 4 days. |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate |
| Chloride (Urina | |
| Laboratory: | Clinical Biochemistry |
| Specimen: | Spot or 24 Hr sample |
| Turnaround: | 1 Day |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate |
| Cholesterol | |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Comment: | Fasting sample required |

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| | | Author: | Mr Paul Cantwell | |
| Turnaround: | A/E or urgent sampl | e: - 1 hour 30 | mins approx. CUH | wards, CUMH, SI, SF, |
| | | | | posted within 4 days. |
| Ref. Range: | | | | ochemistry reports as |
| 2 | appropriate | | | |
| Cholinesterase | : Phenotyping And | Genotyping | | |
| Laboratory: | | | emistry to, Cholin | esterase Investigation |
| | Unit, Department of | | | |
| | Southmead Hospital | , Bristol BS10 S | 5NB,UK | |
| Specimen: | 4.0 mL EDTA whole | blood | | |
| | Sample should NO | T be taken d | uring Sux-induced | l after apnoea as the |
| | presence of the drug | g can lead to er | roneously low enz | yme activity. |
| | Test request should | be delayed for | 24 hours and for | 6 weeks if fresh frozen |
| | plasma is administe | red. | | |
| Turnaround: | 8 weeks | | | |
| Ref. Range: | Contact Biochemistr | y (ext 22531) | | |
| Chromium & Co | obalt (non-De Puy h | nips) | | |
| Laboratory: | Sample referred from | m Clinical Bioch | emistry to Trace r | netal laboratory, |
| | Guilford, Surrey | | | |
| Specimen: | 2 ml whole blood tra | ace metal free b | oottle | |
| Comment: | Fasting sample requ | ired | | |
| Turnaround: | 6 weeks | | | |
| Ref. Range: | See report or conta | ct Trace metal | laboratory, Guilfor | d, Surrey 00-44-148 |
| _ | 368 9978 (Technica | l & Clinical Que | ries) | |
| Chromosome A | nalysis / Karyotype | e <5 years old | | |
| Laboratory: | Referred from Molec | ular Genetics L | ab in Biochemistry | v to NCMG). |
| | Patients <5yr are re | ferred to NCMC | 6. Referrals Mon-T | hurs only. |
| Specimen: | DO NOT refrigerate | specimens. | | |
| | Infants: 1mL Lithiun | n Heparin blood | 1 | |
| Comment: | Copy of NCMG reque | est form with co | onsent available at | <u>www.genetics.ie</u> . |
| Turnaround: | See NCMG website (| TAT depends o | n priority/ 6weeks | |
| Report: | Sent to referring clir | nician and copy | of report filed in p | oathology |
| Chromosome A | nalysis / Karyotype | e >5 years old | l | |
| Laboratory: | Referred from Molec | ular Genetics L | ab in Biochemistry | via Med lab Path to the |
| | Doctor's Lab, Londo | n (TDL). | | |
| | Samples sent Mon-T | hurs or by spec | ial arrangement be | efore 9.30am on Fridays |
| | (contact ex 22531 to | | | |
| Specimen: | DO NOT refrigerate | specimens. | | |
| | Adults: 3mL Lithium | Heparin blood | | |
| | Infants: 1mL Lithiun | n Heparin blood | 1 | |
| Comment: | Please use consent f | ^f orm available a | at | |
| | http://www.soniche | <u>althcare.ie/test</u> | -information/reque | <u>est-forms.aspx</u> |
| | Please note: invoice | s are issued dir | ectly to referring o | clinician. |
| Turnaround: | 6 weeks | | - | |
| Report: | Report sent to refer | ring clinician ar | d copy of report fi | led in pathology |
| Citrate (Urinar | | | | |
| Laboratory: | | m Clinical Bioch | emistry to Eurofin | s-Biomnis Laboratories |
| Specimen: | 24 hour urine, must | | • | |
| | - | | • | |
| Turnaround: | 3 weeks | | | |
| Turnaround: Ref. Range: | | visit internet si | te https://www.eurof | ins.ie/biomnis/ for up to |

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| CLIFT (Crithidi | a Luciliae Immuno Fluorescence Test) |
|-----------------|--|
| Laboratory: | Autoimmune Serology |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) |
| Comment: | Qualitative immunofluorescent assay. Automatically checked following |
| | Positive Anti Nuclear Antibody assay showing a Homogenous ANA Patten of |
| | immunofluorescence. If CLIFT assay is positive a further quantitative Anti |
| | dsDNA Immunoassay is carried out. |
| Turnaround: | 72 Hours |
| Ref. Range: | Not applicable |
| Clostridioides | difficile Testing |
| Laboratory: | Microbiology (Category 3 Laboratory) |
| Specimen: | Fresh faeces specimen. 1-2g (1-2mL) is sufficient. |
| Comment: | A molecular diagnostic assay is used for the direct qualitative detection of |
| | Clostridioides difficile toxin B gene in human faeces samples. |
| | Test performed Monday to Friday. |
| | Testing on individuals from 3-16 years should be restricted but exceptions |
| | can be made where indicated by the Microbiology Medical team. Testing not |
| | recommended on children <2. |
| | Requests for C. difficile are performed on inpatients, healthcare-associated |
| | and community individuals where the specimen takes the shape of the |
| | container and also on contacts during an outbreak. |
| | Repeat testing is not routinely performed on specimens positive or negative |
| | within the last 21 days except by prior approval with the Microbiology |
| | Medical team. |
| | Test of cure is not recommended. |
| | Specimens should be sent to the laboratory as soon as possible after |
| | collection for testing. If there is a delay in transit specimens should be |
| | stored in a refrigerator at 2-8°C, and tested within 72 hours. |
| | Samples greater than 3 days old on receipt in the laboratory are unsuitable |
| | for testing. |
| Turnaround: | Within 24 hours if received between Monday and Thursday; specimens |
| | received on Friday after 11:30am should be reported before 5 pm on the |
| | following Monday. |
| | Urgent specimens may be processed at weekends following consultation with |
| | the Microbiology Consultant. |
| | Positive reports are telephoned when available to the requesting area. |
| Report: | C. difficile toxin PCR target NOT detected/TARGET DETECTED. |
| • | C. difficile Toxin testing carried out on all PCR TARGET DETECTED samples. |
| | A Target Not Detected result does not automatically exclude infection from |
| | C. difficile as the level of DNA present may be lower than the limit of |
| | detection of the assay. |
| CLL Prognostic | : Markers (TP53 and IGVH mutation status) |
| Laboratory: | Referred from Haematology Dept to Royal Marsden Hospital UK |
| Specimen: | Blood 3 mL purple Vacuette (EDTA) 5 -10 mLs required and 3 mL green |
| | Vacuette (Lithium Heparin) |
| | Available Mon – Thurs, sample to reach Haematology Lab by 12 noon on |
| | day of sampling. |
| Comment: | Prognostic markers for CLL |
| Turnaround: | 62 working days |
| Report: | Sent to referring clinician and copy filed in laboratory |

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| | Author: | Mr Paul Cantwell | |

| <u>Coayulatioli</u> Fa | ctor VIII Inhibitors – Quantitation Assay |
|--------------------------|--|
| Laboratory: | Haematology |
| Specimen: | Blood 3mL x 2, blue Vacuette (sodium citrate 3.2%). |
| | Specimens that are haemolysed, underfilled or overfilled cannot be analysed |
| | check coagulation sample bottles are not expired to ensure correct filling. |
| Comment: | Test available Monday to Friday, during routine working hours by |
| | arrangement with the Haematology dept. Quantitation of coagulation |
| | factor inhibitors reported in Bethesda Units. One Bethesda Unit is the |
| | amount of inhibitor in 1 mL of plasma that will neutralise 50% of the clotting |
| | factor activity. |
| | Samples must be received within 4 hours |
| Turnaround: | 2 – 4 weeks |
| Report: | Negative |
| | Weak Factor Inhibitor: = 10 BU/mL.</td |
| | Strong Factor Inhibitor: > 10 BU/mL. |
| Coagulation Fa | ictor Inhibitor Screen |
| Laboratory: | Haematology |
| Specimen: | Blood 3mL x 2; blue Vacuette \mathbb{R} (sodium citrate 3.2%) |
| • | Specimens that are haemolysed, underfilled or overfilled cannot be |
| | analysed, check coagulation sample bottles are not expired to ensure |
| | correct filling. |
| Comment: | Demonstrates the inhibitory effect of Coagulation Factor antibodies. Test |
| Commenter | available Monday to Friday, during routine working hours by arrangement |
| | with the Haematology dept. See also Coagulation factor VIII Inhibitors – |
| | Quantitation Assay. |
| | Samples must be received within 4 hours |
| Turnaround: | Routine specimens: 2 weeks |
| Report: | Positive / Negative |
| Cocaine | |
| Laboratory: | Sample referred from Clinical Biochemistry to Toxicology Laboratory |
| | BEAUMONT Hospital Dublin, posted Monday, Tuesday, Wednesday and |
| | Thursday. |
| Specimen: | Spot urine |
| Comment: | See Toxicology / Drug Screen |
| Turnaround: | 1 week |
| Ref. Range: | See report form or contact Toxicology Laboratory BEAUMONT Hospital 01- |
| iter iteriger | 8092673 / (01)8092675, Emergency after hours (087) 2590749, Fax (01) |
| | 8093986 |
| Coccidioides A | |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Comment: | Performed by a reference laboratory (UKHSA Mycology Reference |
| | Laboratory, Bristol) |
| Turnaround: | 28 working days |
| Report: | Qualitative result |
| Coeliac Screen | |
| | Autoimmune Serology |
| Laboratorv: | |
| Laboratory: Specimen: | |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) |
| • | Blood, 4 mL red top Vacuette (or similar container for clotted blood) Includes IgA Anti-tTG plus IgA Anti-EMA if Anti-tTG Positive. |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) Includes IgA Anti-tTG plus IgA Anti-EMA if Anti-tTG Positive. IgA deficient sera automatically detected on Anti-tTG assay. Deficient sera |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) Includes IgA Anti-tTG plus IgA Anti-EMA if Anti-tTG Positive. IgA deficient sera automatically detected on Anti-tTG assay. Deficient sera are analyzed for total serum IgA. IgA deficient sera are tested for IgG Anti- |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) Includes IgA Anti-tTG plus IgA Anti-EMA if Anti-tTG Positive. |

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| Ref. Range: | 0 - 5 AU/mL |
|----------------------|---|
| Cold Agglutini | ns |
| Laboratory: | Blood Transfusion Laboratory |
| Specimen: | For Pre-Cardiac Surgery Patients: 1 x 6 ml EDTA Pink Capped Tube |
| | For investigation of Cold Haemagglutinin Disease: 1 x 4 mL Clotted Sample |
| | (red cap/yellow ring tube) and 1 x 6 ml EDTA Pink Capped Tube BOTH |
| | brought to laboratory while still warm 37°C if possible. |
| Comment: | This test is performed to detect cold agglutinins: |
| | In Pre-Cardiac surgery patients at ambient room temperature (18-25°C). |
| | In Cold Haemagglutinin Disease (CHAD). |
| | Complete the Blood Transfusion request form LF-C-BTR-XMATCH. |
| | NOTE: This is not an accredited test. |
| Turnaround: | 8 Hours (Note: This may exceed 8 hours if positive for cold agglutinins) |
| Ref. Range: | Not applicable |
| Conjunctivitis | |
| See Eye Swał | D. |
| | NB1) Mutation analysis |
| Laboratory: | Referred from Molecular Genetics Lab in Biochemistry to Leeds NHS (via |
| , | NCMG) |
| Specimen: | 3-5ml EDTA |
| Comment: | Use NCMG request form with consent available from www.genetics.ie |
| | Please note: invoices will be issued to the referring clinician for tests not |
| | performed in NCMG. |
| Turnaround: | 40 days |
| Report: | Sent to referring clinician and copy filed in pathology |
| Continuous An | nbulatory Peritoneal Dialysis Fluid |
| Laboratory: | Microbiology (Main laboratory) |
| Specimen: | Ideally, a volume of 20mL should be collected into a clean, sterile, leakproof |
| | container. In addition, blood culture bottles should be inoculated aseptically |
| | with 5-10mL of dialysate. Transport ASAP. If processing is delayed, |
| | refrigeration of the 20mL aliquot is preferable to storage at room |
| | temperature. |
| Comment: | Test performed as an urgent specimen. If routine cultures are negative and |
| | abnormal dialysate findings persist, please discuss with the Microbiology medical staff. If mycobacterial culture is required it should be specifically |
| | requested. |
| Turnaround: | Microscopy: 2 hours. Urgent report telephoned when available. |
| Turnarouna. | Prelim: 48 hours; Final: 5 days. Clinically significant isolates are telephoned |
| | when available. |
| Report: | White cell count and aerobic culture. Where the white cell count is \geq 50/cmm |
| | a Gram stain and white cell differential is performed. |
| Copper | |
| Laboratory: | Referred from Clinical Biochemistry to SAS Laboratory for Trace Elements, |
| , | Guildford |
| Specimen: | Sod Hep trace metal free tube (navy top) |
| Turnaround: | 2 weeks |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as |
| | appropriate |
| Copper (Urina | ry) |
| Laboratory: | Referred from Clinical Biochemistry to SAS Laboratory for Trace Elements, Guildford. |
| Specimen: | 24 hr urine sample |
| | 1 |

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|--|---|--|---|---|
| | | Author: | Mr Paul Cantwell | |
| Comment: | Lico acid washod co | ntainar anly | | |
| Turnaround: | Use acid-washed co 3 weeks | Intainer only | | |
| | | shomictry labor | ton | |
| Ref. Range: | Contact Clinical Bio | chemistry labora | itory | |
| Corneal Scrapi | | craningo | | |
| | ular fluids /Corneal S | crapings | | |
| | Clinical Dischamistr | | | |
| Laboratory: | Clinical Biochemistr | • | | |
| Specimen: | 4.0 mL blood in plai | n tube (clotted | sample) | |
| Turnaround: | 3 Days | o intorvale will l | a applied to all Big | ochemistry reports as |
| Ref. Range: | appropriate | | be applied to all bit | schemistry reports as |
| Cortisol-Saliva | rv | | | |
| Laboratory: | | m Clinical Bioch | emistry to Universi | ity Hospital of Wales, |
| Laboratory | Cardiff | | | ···, ·····, |
| Specimen: | Saliva collected in S | alivette contain | er | |
| Comment: | Time of sample mus | st be recorded. | | |
| Turnaround: | 5-6 weeks | | | |
| Ref. range: | See report form | | | |
| Cortisol (Urina | ry) | | | |
| Laboratory: | | al Biochemistry | to Biochemistry La | aboratory in the Mater |
| , | Hospital, Dublin. | , | , | , |
| Specimen: | 24 Hour urine collect | tion | | |
| Turnaround: | 2 Weeks | | | |
| Ref. Range: | Up-to-date reference | e intervals will l | be applied to all Bio | ochemistry reports as |
| | appropriate | | | |
| COVID-19 (Mo | lecular) | | | |
| | | | | |
| See section: | | | | |
| SARS CoV-2 | | | | |
| SARS CoV-2 Coxiella burne | tii IgG and IgM (Q | | | |
| GARS CoV-2 Coxiella burne Laboratory: | Microbiology (Infect | | erology) | |
| GARS CoV-2 Coxiella burne Laboratory: Specimen: | Microbiology (Infect 4mL clotted blood | ious Diseases S | | |
| GARS CoV-2 Coxiella burne Laboratory: | Microbiology (Infect 4mL clotted blood Performed by a refe | ious Diseases S rence laborator | | d Pathogens Laborator |
| SARS CoV-2 Coxiella burne Laboratory: Specimen: Comment: | Microbiology (Infect 4mL clotted blood Performed by a refe (RIPL), Porton Down | ious Diseases S rence laborator | | d Pathogens Laborator |
| GARS CoV-2 Coxiella burne Laboratory: Specimen: Comment: Turnaround: | Microbiology (Infect 4mL clotted blood Performed by a refe (RIPL), Porton Down 28 working days | ious Diseases S rence laborator | | d Pathogens Laborator |
| SARS CoV-2 Coxiella burne Laboratory: Specimen: Comment: Turnaround: Report: | Microbiology (Infect 4mL clotted blood Performed by a refe (RIPL), Porton Down 28 working days Qualitative result | ious Diseases S rence laborator | | d Pathogens Laborator |
| SARS CoV-2 Coxiella burne Laboratory: Specimen: Comment: Turnaround: Report: Creatine Kinas | Microbiology (Infect 4mL clotted blood Performed by a refe (RIPL), Porton Down 28 working days Qualitative result e (CK) | ious Diseases S rrence laborator າ) | | d Pathogens Laborator |
| GARS CoV-2 Coxiella burne Laboratory: Specimen: Comment: Turnaround: Report: Creatine Kinas Laboratory: | Microbiology (Infect 4mL clotted blood Performed by a refe (RIPL), Porton Down 28 working days Qualitative result e (CK) Clinical Biochemistr | ious Diseases S rence laborator ı) y | y (Rare & Imported | d Pathogens Laborator |
| GARS CoV-2 Coxiella burne Laboratory: Specimen: Comment: Turnaround: Report: Creatine Kinas Laboratory: Specimen: | Microbiology (Infect 4mL clotted blood Performed by a refe (RIPL), Porton Down 28 working days Qualitative result e (CK) Clinical Biochemistr 4.0 mL blood in plai | ious Diseases S rence laborator ר) y n tube (clotted | y (Rare & Imported | |
| GARS CoV-2 Coxiella burne Laboratory: Specimen: Comment: Turnaround: Report: Creatine Kinas Laboratory: | Microbiology (Infect 4mL clotted blood Performed by a refe (RIPL), Porton Down 28 working days Qualitative result e (CK) Clinical Biochemistr 4.0 mL blood in plai A/E or urgent samp | ious Diseases S prence laborator n) y n tube (clotted le: - 1 hour 30 p | y (Rare & Imported sample) nins approx. CUH | wards, CUMH, SI, SF, |
| ARS CoV-2 Coxiella burne Laboratory: Specimen: Comment: Turnaround: Report: Creatine Kinas Laboratory: Specimen: Turnaround: | Microbiology (Infect 4mL clotted blood Performed by a refe (RIPL), Porton Down 28 working days Qualitative result e (CK) Clinical Biochemistr 4.0 mL blood in plai A/E or urgent samp SMOH, MGH: - 3 ho | ious Diseases S rence laborator n) y n tube (clotted le: - 1 hour 30 n urs approx. GP | y (Rare & Imported sample) nins approx. CUH or OPD- Results p | wards, CUMH, SI, SF, osted within 4 days. |
| GARS CoV-2 Coxiella burne Laboratory: Specimen: Comment: Turnaround: Report: Creatine Kinas Laboratory: Specimen: | Microbiology (Infect 4mL clotted blood Performed by a refe (RIPL), Porton Down 28 working days Qualitative result e (CK) Clinical Biochemistr 4.0 mL blood in plai A/E or urgent samp SMOH, MGH: - 3 ho Up-to-date reference | ious Diseases S rence laborator n) y n tube (clotted le: - 1 hour 30 n urs approx. GP | y (Rare & Imported sample) nins approx. CUH or OPD- Results p | wards, CUMH, SI, SF, |
| SARS CoV-2 Coxiella burne Laboratory: Specimen: Comment: Turnaround: Report: Creatine Kinas Laboratory: Specimen: Turnaround: Ref. Range: | Microbiology (Infect 4mL clotted blood Performed by a refe (RIPL), Porton Down 28 working days Qualitative result e (CK) Clinical Biochemistr 4.0 mL blood in plai A/E or urgent samp SMOH, MGH: - 3 ho Up-to-date reference appropriate | ious Diseases S rence laborator n) y n tube (clotted le: - 1 hour 30 n urs approx. GP | y (Rare & Imported sample) nins approx. CUH or OPD- Results p | wards, CUMH, SI, SF, osted within 4 days. |
| SARS CoV-2 Coxiella burne Laboratory: Specimen: Comment: Turnaround: Report: Creatine Kinas Laboratory: Specimen: Turnaround: Ref. Range: Creatinine (Blo | Microbiology (Infect 4mL clotted blood Performed by a refe (RIPL), Porton Down 28 working days Qualitative result e (CK) Clinical Biochemistr 4.0 mL blood in plai A/E or urgent samp SMOH, MGH: - 3 ho Up-to-date reference appropriate | ious Diseases S erence laborator n) y n tube (clotted le: - 1 hour 30 r urs approx. GP e intervals will l | y (Rare & Imported sample) nins approx. CUH or OPD- Results p | wards, CUMH, SI, SF, osted within 4 days. |
| ARS CoV-2 Coxiella burne Laboratory: Specimen: Comment: Turnaround: Report: Creatine Kinas Laboratory: Specimen: Turnaround: Ref. Range: Creatinine (Blo Laboratory: | Microbiology (Infect 4mL clotted blood Performed by a refe (RIPL), Porton Down 28 working days Qualitative result e (CK) Clinical Biochemistr 4.0 mL blood in plai A/E or urgent samp SMOH, MGH: - 3 ho Up-to-date reference appropriate od) Clinical Biochemistr | ious Diseases S rence laborator n) y n tube (clotted le: - 1 hour 30 urs approx. GP e intervals will l | y (Rare & Imported sample) mins approx. CUH or OPD- Results p be applied to all Bid | wards, CUMH, SI, SF, osted within 4 days. |
| SARS CoV-2 Coxiella burne Laboratory: Specimen: Comment: Turnaround: Report: Creatine Kinas Laboratory: Specimen: Turnaround: Ref. Range: Creatinine (Blo Laboratory: Specimen: | Microbiology (Infect 4mL clotted blood Performed by a refe (RIPL), Porton Down 28 working days Qualitative result e (CK) Clinical Biochemistr 4.0 mL blood in plai A/E or urgent samp SMOH, MGH: - 3 ho Up-to-date reference appropriate od) Clinical Biochemistr 4.0 mL blood in plai | ious Diseases S prence laborator n) y n tube (clotted le: - 1 hour 30 n urs approx. GP e intervals will l y n tube (clotted | y (Rare & Imported sample) mins approx. CUH or OPD- Results p be applied to all Bid sample) | wards, CUMH, SI, SF, osted within 4 days. ochemistry reports as |
| ARS CoV-2 Coxiella burne Laboratory: Specimen: Comment: Turnaround: Report: Creatine Kinas Laboratory: Specimen: Turnaround: Ref. Range: Creatinine (Blo Laboratory: | Microbiology (Infect 4mL clotted blood Performed by a refe (RIPL), Porton Down 28 working days Qualitative result e (CK) Clinical Biochemistr 4.0 mL blood in plai A/E or urgent samp SMOH, MGH: - 3 ho Up-to-date reference appropriate DOD Clinical Biochemistr 4.0 mL blood in plai A/E or urgent samp | ious Diseases S prence laborator n) y n tube (clotted le: - 1 hour 30 n urs approx. GP e intervals will l y n tube (clotted le: - 1 hour 30 n | y (Rare & Imported sample) mins approx. CUH or OPD- Results p be applied to all Bid sample) mins approx. CUH | wards, CUMH, SI, SF, osted within 4 days. ochemistry reports as wards, CUMH, SI, SF, |
| SARS CoV-2 Coxiella burne Laboratory: Specimen: Comment: Turnaround: Report: Creatine Kinas Laboratory: Specimen: Turnaround: Ref. Range: Creatinine (Blo Laboratory: Specimen: Turnaround: | Microbiology (Infect 4mL clotted blood Performed by a refe (RIPL), Porton Down 28 working days Qualitative result e (CK) Clinical Biochemistr 4.0 mL blood in plai A/E or urgent samp SMOH, MGH: - 3 ho Up-to-date reference appropriate DOD Clinical Biochemistr 4.0 mL blood in plai A/E or urgent samp SMOH, MGH: - 3 ho | ious Diseases S erence laborator n) y n tube (clotted le: - 1 hour 30 h urs approx. GP e intervals will l y n tube (clotted le: - 1 hour 30 h urs approx. GP | y (Rare & Imported sample) mins approx. CUH or OPD- Results p be applied to all Bid sample) mins approx. CUH or OPD- Results p | wards, CUMH, SI, SF, osted within 4 days. ochemistry reports as wards, CUMH, SI, SF, osted within 4 days. |
| SARS CoV-2 Coxiella burne Laboratory: Specimen: Comment: Turnaround: Report: Creatine Kinas Laboratory: Specimen: Turnaround: Ref. Range: Creatinine (Blo Laboratory: Specimen: | Microbiology (Infect 4mL clotted blood Performed by a refe (RIPL), Porton Down 28 working days Qualitative result e (CK) Clinical Biochemistr 4.0 mL blood in plai A/E or urgent samp SMOH, MGH: - 3 ho Up-to-date reference appropriate Od) Clinical Biochemistr 4.0 mL blood in plai A/E or urgent samp SMOH, MGH: - 3 ho Estimated Glomerul | ious Diseases S rence laborator n) y n tube (clotted le: - 1 hour 30 r urs approx. GP e intervals will l y n tube (clotted le: - 1 hour 30 r urs approx. GP ar filtration rate | y (Rare & Imported sample) mins approx. CUH or OPD- Results p be applied to all Bid sample) mins approx. CUH or OPD- Results p (eGFR) is availabl | wards, CUMH, SI, SF, osted within 4 days. ochemistry reports as wards, CUMH, SI, SF, osted within 4 days. e on request. Method |
| SARS CoV-2 Coxiella burne Laboratory: Specimen: Comment: Turnaround: Report: Creatine Kinas Laboratory: Specimen: Turnaround: Ref. Range: Creatinine (Blo Laboratory: Specimen: Turnaround: | Microbiology (Infect 4mL clotted blood Performed by a refe (RIPL), Porton Down 28 working days Qualitative result e (CK) Clinical Biochemistr 4.0 mL blood in plai A/E or urgent samp SMOH, MGH: - 3 ho Up-to-date reference appropriate DOD Clinical Biochemistr 4.0 mL blood in plai A/E or urgent samp SMOH, MGH: - 3 ho Estimated Glomerul adjusted 4-variable | ious Diseases S rence laborator n) y n tube (clotted le: - 1 hour 30 r urs approx. GP e intervals will l y n tube (clotted le: - 1 hour 30 r urs approx. GP ar filtration rate MDRD formula | y (Rare & Imported sample) mins approx. CUH or OPD- Results p be applied to all Bid sample) mins approx. CUH or OPD- Results p (eGFR) is availabl is used for calculat | wards, CUMH, SI, SF, osted within 4 days. ochemistry reports as wards, CUMH, SI, SF, osted within 4 days. e on request. Method |

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| Creatinine (PC | DCT) | | | | | |
|------------------------|--|--|----------------|----------------|---------------|--------------|
| | iStat Alinity for Creatinine is for use with Adult samples only . | | | | | |
| | Heparinised Arterial, Venous or Capillary Samples may be used. | | | | | |
| | Minimum sa | mple volume = 6 | 5µL | | | |
| Time to | . | | | | | |
| result: | 2 minutes | | | | | |
| Ref range: | Measured | | | | | |
| | Test | Units | Reportable | Reference | Range | |
| | | | Range | Arterial | Venous | Critical |
| | Creatinine | µmol/L | 18-1768 | 53-115* | 53-115* | ≥ 300 |
| | eGFR | ml/min/1.73m 2 | n/a | n/a | n/a | ≤ 30 |
| | Estimated G calculation. | lomerular filtratio | on rate (eGFI | R) is calcula | ated using t | he UK-MDRD |
| | NOTE: Aler | t/critical result | s must be co | onfirmed w | ith a veno | us sample. |
| Comment | | ample must be | | | | - |
| | | for determination | | • | | |
| | • | the capillary POC | | | | |
| | | 0 will require disc | | | | |
| | | a non contrast stu | | | . , | |
| <u>Cuestinine (III</u> | ······································ | | | | | |
| Creatinine (U | | chomictry (| | | | |
| Laboratory: | | ochemistry | no cloaranco | (Spot comp | la for micro | albumin / |
| Specimen: | | mple for creatini | | (Spot samp | le for microa | aidumin / |
| Turnaround: | creatinine ratio, see below) 1 Day | | | | | |
| Ref. Range: | | | | | | |
| | appropriat | | • | • | | |
| Creatinine Cle | arance | | | | | |
| Laboratory: | | ochemistry | | | | |
| Specimens: | | ood in a plain tub | e (clotted san | nple) and a | 24-hour uri | ne sample. |
| Turnaround: | | 1 Day Up-to-date reference intervals will be applied to all Biochemistry reports as | | | | |
| Ref. Range: | • | | als will be ap | plied to all I | Biochemistry | reports as |
| CRP | appropriat | e | | | | |
| Laboratory: | Clinical Big | ochemistry | | | | |
| Specimen: | | od in a plain tub | e (clotted san | nple) | | |
| Comment: | | when appropriat | • | • • | ided. | |
| | | is not suitable for | | | | disease. |
| Turnaround: | , | ent sample: - 1 h | | | | |
| - | | GH: - 3 hours app | | | , - | , |
| Ref. Range: | • | e reference interv | als will be ap | plied to all | Biochemistry | y reports as |
| Currente la la serie | appropriat | е | | | | |
| Cryoglobulin | Clinias I D' | ab a painting (Trans | unalogy Lab | notors () | | |
| Laboratory: | | chemistry (Immu | | | + 27 00 | |
| Specimen: | | t be collected into all sent to the lat | | | | |
| Comment: | | e with Laboratory | | - | | 57 C. |
| Turnaround: | 5 Days | | y LAC. 2233 | | | |
| Ref. Range: | • | in should be NEG | ATIVE | | | |
| | Cryoglobul | | | | | |

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| Cryptococcal A | Intigen –Blood sample |
|----------------|---|
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Comment: | Performed by a reference laboratory (Mycology Reference Centre, Bristol) |
| Turnaround: | 28 working days |
| Report: | Negative or Positive (Titre) |
| Cryptococcal A | Antigen –CSF sample |
| Laboratory: | Microbiology |
| Specimen: | CSF (0.3mL minimum) |
| Comment: | Performed by a reference laboratory (Mycology Reference Centre, Bristol) |
| Turnaround: | 28 working days |
| Report: | Negative or Positive (Titre) |
| Cryptosporidiu | ım Species |
| Laboratory: | Microbiology (Category 3 Laboratory) |
| Specimen: | Faeces. |
| | Performed routinely on all suitable faeces samples submitted for Routine Molecular Enteric Screening. |
| | Other types of clinical specimen such as duodenal aspirates are also stained for cryptosporidia. |
| Comment: | Test performed routinely Monday to Friday 9-5pm. Diagnosis is based upon the molecular detection of <i>Cryptosporidium parvum/hominis</i> and demonstration of oocysts in faeces samples using a modified Ziehl-Neelsen stain. |
| | A Target Not Detected result does not automatically exclude infection from the above enteric pathogen as the level of DNA present may be lower than the limit of detection of the assay. |
| Turnaround: | 36 hours. |
| Report: | PCR for Cryptosporidium parvum/hominis: Target DETECTED or target NOT detected. |
| | Oocysts of Cryptosporidium seen or not seen |

| See Cerebros | pinal Fluid |
|-----------------|--|
| CSF Oligoclona | al bands and CSF IgG Index |
| See Cerebros | pinal Fluid - Oligoclonal bands and CSF IgG Index |
| CSF Viral Scree | en |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | CSF (>0.5mL) |
| Tests: | Molecular tests for enterovirus, herpes simplex virus (HSV 1/2), varicella- zoster virus (VZV). For patients <3 years of age, human herpes virus 6 (HHV-6) and parechovirus are also included. |
| Comment: | Testing performed by a reference laboratory (National Virus Reference Laboratory (NVRL), Dublin) |
| Turnaround: | 10 working days |
| Report: | Detected or not detected |
| CSU - Catheter | [·] Urine |
| See Urine Mi | croscopy and Culture |
| Cyclosporin (N | leoral) |
| Laboratory: | Clinical Biochemistry |

| Clinical Biochemistry |
|--|
| Trough sample required, (Blood 3mL, EDTA). Analysed on Thursdays |
| 7-8 days |
| |

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| Ref. Range: | Patient specific Interpretation of Cyclosporin is dependent on time interval |
|-------------|--|
| | between sample and last dose, clinical indication for use of the drug, |
| | duration of therapy and other drug therapy and method of measurement. |

| | defation of therapy and other drug therapy and method of medsarement. |
|------------------------|--|
| Cystic Fibrosis | s (CF) |
| Laboratory: | Specimens referred from Molecular Genetics Lab in Biochemistry to NCMG. |
| Specimen: | Adults: 3-5 ml EDTA blood, |
| | Infants: 1ml EDTA blood |
| Comment: | NCMG request form available from <u>www.genetics.ie</u> . |
| | Patient Information Request (PIR) form for carrier status in CF families |
| | available from <u>www.genetics.ie</u> |
| Turnaround: | 6-8 weeks |
| Report: | Sent to referring clinician by NCMG and copy of report filed in pathology |
| Cystine (WI | BC) |
| Laboratory: | Specimen are referred to CHI Temple Street (cell preparation) and then to |
| | Wellchild Lab (analysis) |
| Specimen: | 3 ml Li-Hep whole blood. |
| Comment: | 4-5 hours post Cystagon dose |
| Ref. Range | See report form |
| Cytogenetics | (Chromosome banding) for the diagnosis of AML, CML, ALL and MDS |
| Laboratory: | Referred from Haematology to Munich Leukaemia Laboratory (MLL MVZ |
| | GmbH), Germany |
| Specimen: | 5 ml heparin bone marrow |
| Comment: | Must arrange with Haematology, transport within 24 hours, complete form from |
| | referral laboratory |
| Turnaround: | 3-7 working days |
| Report: | Sent to referring clinician and copy filed in laboratory |
| | |

Cytological Examination

| Laboratory: | Histopathology (Cytology Department) |
|-------------|--------------------------------------|
| Specimen: | Cerebrospinal Fluid (CSF) - Cytology |
| | See Cerebrospinal Fluid |

Fine Needle Aspirate (FNAs)

An immediate fine needle aspiration service is available on request for both in-patients and out-patients. Aspirations are performed by a consultant Cytopathologist for palpable lesions. This can be arranged by discussion with the Laboratory (Ext.22511) or with the consultant (Ext.20499).

An FNA clinic accepting GP referrals for patients with palpable swellings is available on Thursday afternoons. A Consultant FNA Referral form needs to be completed and faxed/sent to the laboratory to arrange an appointment. This form is available in the CUH Staff Directory under CUH Forms or alternatively, by contacting 021 4922883/4922510.

Assistance to those performing FNAs in radiology is available before 4.30pm Monday to Friday. The service must be pre-booked with the Cytopathology laboratory @ Ext.22511.

Other Diagnostic Specimens

- Sputa specimens are collected in sterile universal containers early morning on three consecutive days
- Bronchial samples, Serous fluids etc all collected according to local protocols in sterile universal containers and transported to the laboratory as soon as possible. Protocols available from the cytology laboratory.

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| | | | | |
| | - | • | n volume of 30 m | Ls. Please do not |
| | submit drain bags. Urines – specimens are collected into sterile universal containers. Joint fluid – see Joint Aspirate for Crystals. | | | |
| | | | | |
| | | • | • | - I |
| | | | amples taken out | ology and Endoscopy fo of hours where |
| Comment: | Tests are performed hours. | l routinely Mond | lay to Friday durir | ng routine working |
| Turnaround: | Non gynaecological | cvtology – FNA | - 80% of cases b | v dav 5 |
| | Non gynaecological | | | |
| | A verbal report may | be available wi | thin 2 hours for c | linically urgent samples |
| | by prior communica | | | |
| Ref. Range: | Not applicable. | | | |
| | us (CMV) IgG and I | [aM | | |
| Laboratory: | Microbiology (Infect | | erology) | |
| Specimen: | 4mL clotted blood | | 0.010977 | |
| Comment: | | aG antihodies a | re tested senarat | elv. The clinician must |
| comment. | CMV IgM and CMV IgG antibodies are tested separately. The clinician must indicate the appropriate test by full history <i>etc</i> . | | | |
| Turnaround: | 36 hours | The cost by full | | |
| Report: | Qualitative result | | | |
| | us (CMV) Molecular | r | | |
| | | | orology | |
| Laboratory: | Microbiology (Infect | | | ncho-alvoolar lavago |
| Specimen: | nasopharyngeal asp | | | ncho-alveolar lavage, |
| Comment: | | | | Reference Laboratory |
| comment. | (NVRL), Dublin) | | | Reference Laboratory |
| Turnaround: | 14 working days | | | |
| Report: | Detected (viral load |) or not detecte | d | |
| | or-specific) Antibo | / | ч | |
| | | | | |
| Laboratory: | Blood Transfusion L | • | | |
| Specimen: | 5-10ml clotted blood | · · | , | nunaganatica |
| Comment: | This test is carried of Laboratory, Beaumo | , | . , | nunogenetics |
| Turnaround: | Contact Histocompa | | | oratory Resument |
| | Hospital, Dublin 9. | icionicy and min | anogenetics Labo | Statory, Deaumont |
| -dimers | | | | |
| | Haomatology | | | |
| Laboratory: Specimen: | Haematology Blood 3mL, blue Va | cuatta® (cadius | citrato 2 20/1 | |
| specimen: | • | • | , | hotomy |
| Commont | Specimens must be | | • | |
| Comment: | | | | nostic for lysis of a fibri working hours, and for |
| | | • | | working nours, and for |
| | emergency reasons | | | 8 hours |
| Turnaround | Emergency specime | insi o nours; Ro | aune specimens: | 0 110015 |
| Turnaround: | | | | |
| Turnaround: Ref. Range: | Negative: 0 – 0.5 m Positive: > 0.5mg/L | | | |

Dengue Virus IgG and IgM

| Laboratory: | Microbiology (Infectious Diseases Serology) |
|-------------|---|
| Specimen: | 4mL clotted blood |

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| | | Author: | Mr Paul Cantwell | | | |
| Comment: | Performed by a refe | erence laborator | y (National Virus F | Reference Laboratory | | |
| | (NVRL), Dublin) | | | , | | |
| Turnaround: | 14 working days | | | | | |
| Report: | Qualitative result | | | | | |
| Dexamethas | one Suppression T | est | | | | |
| Laboratory: | Clinical Biochemistr | У | | | | |
| Specimen: | Serum sample | • | | | | |
| Comment: | Clearly indicate on f | form if patient is | s on dexamethaso | ne. | | |
| Turnaround: | 3 days | · | | | | |
| Ref. range: | | e intervals will l | pe applied to all Bi | ochemistry reports as | | |
| 9 | appropriate | | | · · | | |
| Dermatophytos | sis | | | | | |
| See Mycology | | | | | | |
| DHEA Sulphate | 1 | | | | | |
| Laboratory: | Sample referred from Clinical Biochemistry to St. James's University Hospital, Leeds | | | | | |
| Specimen: | 2.0 mL blood in a plain tube (clotted sample) | | | | | |
| Comment: | Consultant request | only | | | | |
| Turnaround: | 4 weeks | | | | | |
| Ref. Range: | See report form | | | | | |
| DHT (Dihydrot | | | | | | |
| Laboratory: | Sample referred from Clinical Biochemistry to St. James's University Hospital, Leeds | | | | | |
| Specimen: | 2.0 mL blood in a plain tube (clotted sample) | | | | | |
| Comment: | Consultant request only | | | | | |
| Turnaround: | 3 weeks | | | | | |
| Ref. Range: | See report form | | | | | |
| Digoxin | | | | | | |
| Laboratory: | Clinical Biochemistry | | | | | |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) | | | | | |
| Comment: | Samples for Digoxin must be taken at least 6 hours post dose. Range quoted | | | | | |
| T | is appropriate for a minimum 6 hours post dose sample. | | | | | |
| Turnaround: | Daily, urgent samples prioritised | | | | | |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate | | | | | |
| Diphtheria | appropriate | | | | | |
| Laboratory: | Clinical Biochemistry | | | | | |
| Specimen: | Blood 4mL red top Vacuette [®] (or similar container for clotted blood) | | | | | |
| Comment: | Test performed by reference laboratory (Respiratory Infections Laboratory, Colindale, London). | | | | | |
| Turnaround: | 2-3 weeks | | | | | |
| Report: | Reported in anti-toxin levels – see specific laboratory report. | | | | | |

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| Laboratory: | Haematology | | | | | |
|----------------------|--|--|--|--|--|--|
| Specimen: | Blood 3mL, blue Vacuette® (sodium citrate 3.2%) | | | | | |
| | Specimens which are haemolysed, underfilled or overfilled cannot be analysed, check coagulation sample bottles are not expired to ensure correct | | | | | |
| | | | | | | |
| | fill. | | | | | |
| Comment: | Used to monitor the edffectivenss of Apixaban and Rivoroxaban therapy. | | | | | |
| | It is essential to state the details of the type of Direct Oral Anticoagulant on request form. Test performed by haematology consultant request only. For accurate interpretation, it is important to know when the drug was last administered and the dose taken. A peak level should be taken 2-4 hours after | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | | | | | | |
| | the drug is taken and a trough level should be taken when the next dose of | | | | | |
| T | the drug is due. | | | | | |
| Turnaround | | | | | | |
| | Refer to report. | | | | | |
| Direct Coomb | | | | | | |
| Laboratory: | Blood Transfusion Laboratory | | | | | |
| Specimen: | 3 mL Purple Capped (FBC) Tube. | | | | | |
| | For Paediatrics: 1 mL EDTA (Purple Cap/White Ring) Paediatric Bottle. | | | | | |
| Comment: | Investigation to demonstrate whether red cells are coated in vivo with | | | | | |
| | immunoglobulins and/or complement. | | | | | |
| | Complete the Blood Transfusion request form LF-C-BTR-XMATCH. | | | | | |
| | This is an INAB accredited test. | | | | | |
| Turnaround: | 3 Hours | | | | | |
| Ref. Range: | Negative or Positive (IgG, IgA, IgM, C3c, C3d). | | | | | |
| | ofluorescence – Renal Biopsy | | | | | |
| See Renal Bi | opsy | | | | | |
| Direct Immun | ofluorescence – Skin/Oral Mucosa | | | | | |
| Laboratory: | Histopathology (E.M Dept.) | | | | | |
| Specimen: | Fresh tissue in Michel's transport medium (Tissue fixative for immunofluorescence) | | | | | |
| Comment: | Fresh specimens are accepted Mon- Fri 8am to 3:30pm only. | | | | | |
| | Where a separate specimen from the same patient is taken for routine | | | | | |
| | Histopathology, it should be delivered to the laboratory with the specimen for | | | | | |
| | Direct Immunofluorescence. | | | | | |
| Turnaround: | 80% in 12 days | | | | | |
| ds-DNA Elisa | | | | | | |
| Laboratory: | Autoimmune Serology | | | | | |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) | | | | | |
| Comment: | Quantitative Elisa. Quantitation of CLIFT Positive Anti-dsDNA sera. | | | | | |
| Turnaround: 72 Hours | | | | | | |
| Ref. Range: | 0 - 200 IU/mL | | | | | |
| Duodenal Asp | | | | | | |
| Laboratory: | Microbiology (Parasitology) | | | | | |
| Specimen: | Specimens will be obtained by specialist collection according to local protocols. The specimen volume may vary - ideally, a minimum volume of 1 mL should be sent to the lab. A screw-capped sterile universal container is practical for this purpose. Transport specimens ASAP. If processing is delayed do NOT refrigerate specimen, leave at room temperature. Delays of | | | | | |

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| | | Aution | Mi Faul Calitwell | | |
| Comment: | Test performed Monday to Friday 9-5pm. Fluid from the duodenum is examined for the presence of <i>Strongyloides stercoralis</i> larvae, <i>Giardia</i> <i>lamblia</i> trophozoites, <i>Cyclospora</i> , and <i>Isospora belli</i> . Duodenal fluid is also examined for the presence of Microsporidia where specifically requested or | | | | |
| Turnaround: | where the patient is immunocompromised. 24 hours. Microsporidia investigation referred to Reference laboratory. (turnaround time varies) | | | | |
| Report: | Report on any parasites seen. Where possible the organism is reported to species level and the stage identified (trophozoite, cyst, oocyst, <i>etc</i>). | | | | |
| Dynamic Fu | · · · | - | 、 、 、 <i>、</i> | | |
| Laboratory: | Clinical Biochemistry | 1 | | | |
| Specimen: | | | with Biochemistr | v Department | |
| Comment: | Dependent on DFT requested, liase with Biochemistry Department Prior arrangement with Biochemistry Department required | | | | |
| | | with biochemist | ry Department red | quired | |
| Turnaound: | Within 24 hours | - : | ha analiad ta all r | | |
| Report: | Up-to-date reference intervals will be applied to all Biochemistry reports as | | | | |
| | appropriate | | | | |
| Ear Swab | Mienelsis / / / / | - h - u - t \ | | | |
| Laboratory: | Microbiology (Main I | •• | | | |
| Specimen: | Swab any pus or ex | | | | |
| Comment: | Test performed routinely Monday to Friday 9-5pm. Transport specimens | | | | |
| | ASAP in charcoal containing transport media. If processing is delayed, | | | | |
| | refrigeration is preferable to storage at room temperature. Tympanocentesis | | | | |
| | (needle aspiration) and Myringotomy (surgical incision of tympanic | | | | |
| | membrane), to specimen middle ear effusion, is rarely justified. | | | | |
| Turnaround: | Prelim: 24 hours; Fi | nal: 48-72 hou | rs | | |
| Report: | Culture report: Any clinically significant isolate with the appropriate | | | | |
| | sensitivities. | | | | |
| Echinococcus (| (Hydatid cyst) Antik | | | | |
| Laboratory: | Microbiology (Infect | ious Diseases S | erology) | | |
| Specimen: | 4mL clotted blood | | | | |
| Comment: | Performed by a reference laboratory (National Parasitology Reference Laboratory (NPRL), London) | | | | |
| Turnaround: | 28 working days | | | | |
| Report: | Qualitative result | | | | |
| <i>E. coli</i> 0157 Se | rology | | | | |
| Test not avail | able. Please refer to F | aeces – Molecu | lar Analysis and | Culture. | |
| <i>E. coli</i> PCR | | | | | |
| Laboratory: | Microbiology (Infect | ious Diseases S | Serology) | | |
| Specimen: | CSF (0.5mL) | | | | |
| Comment: | Performed by Irish Meningitis & Sepsis Reference Laboratory (IMSRL), Dublir | | | | |
| Turnaround: | • | - | • | e 11am, verbal result | |
| rama oana. | between 4pm and 5pm the same day (positive only). | | | | |
| Report: | Detected or not detected | | | | |
| EGFR, ALK, BR | AF, KRAS, NRAS, PI | DL-1*, MMR/N | | | |
| Laboratory: | ROS, Oncomine**, MLH-1 Promoter Methylation studies***. Molecular Pathology/Immunocytochemistry: Molecular testing in the Pathology laboratory CUH is performed on request from Consultant Histopathologists on FFPE tissue samples from patients with Lung cancer, Colon cancer and Melanoma. The current repertoire of tests includes, EGFR with reflex ALK, BRAF, KRAS, NRAS, PDL-1*, MMR/MSI, ERBB2, MET, NTRK 1, NTRK 2, NTRK 3, RET, ROS. FFPE tissue block | | | | |
| Specimen: | | RET, ROS. | | | |

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| | | Authori | | |
| Turnaround: | 5-10 working days | | | |
| | * Some PDL-1 testi | ng is referred to | the Poundbury C | ancer Institute, Dorset |
| | if a different clone is | s required to the | e clone we use in- | house. |
| | Turnaround in this i | nstance is 6 wo | rking days (from o | late testing material is |
| | sent to referral insti | • | | |
| | ** Samples outside | | | - |
| | CMD, St James Hos | | | |
| | days (from date tes | _ | | - |
| | | | | out to the Mancheste |
| | Centre for Genomic | | | ng days (from date |
| | testing material is s | ent to referral i | istitution) | |
| GFR (cfDNA F | | | | |
| Laboratory: | 5, | | | esting in the pathology |
| | plasma samples fro | | • | ant Histopathologists |
| | The cut-off for recei | - | _ | atomy is 15,00 |
| Specimen: | 2 K2 EDTA Blood tu | • | • | |
| Specimen. | OR | | | 5) |
| | at least 1 Roche cfD | NA blood tube | | |
| Comment: | | | to taking the same | ole at Ext.22513 /2279 |
| comment. | | | | atory immediately and |
| | hand directly to th | | | |
| Turnaround: | 5-10 working days | | | |
| lectron Micro | | | | |
| Laboratory: | Pathology (E.M. Dep | nt) | | |
| Specimen: | Fresh unfixed tis | • | orushings in 3% | % glutaraldehyde a |
| opeennen | | , | - | |
| | neuropathology specimens (in-house and referral) in Karnovsky's fixative. (For | | | |
| | renal biopsies see Renal Biopsy) | | | |
| | Please contact the laboratory in advance of the procedure at Ext. 21315 to | | | |
| | organise collection of appropriate specimen container and fixative. | | | |
| | Tissue samples for EM should be brought immediately to the laboratory and | | | |
| | handed directly to | o a Medical Sci | entist. | |
| | Note: For PCD speci | imens, the clinio | cians collect the ap | propriate fixative fron |
| | the laboratory staff | | | |
| Comment: | Specimens are acce | • | 8am to 3:30pm | |
| Turnaround: | 3-5 working days re | enal biopsy | | |
| | 5-7 working days in | -house muscle | biopsy | |
| | 5-7 working days in | -house nerve b | iopsv | |
| | 14 working days samples referred from CUH Neuropathology | | | |
| | | mples referred f | rom Cun Neuropa | |
| | | • | | |
| | 12 weeks PCD samp | les (referred by | EM Dept CUH to P | rimary Ciliary Dyskines |
| | 12 weeks PCD samp (PCD) Diagnostic Se | les (referred by | EM Dept CUH to P | rimary Ciliary Dyskines |
| | 12 weeks PCD samp (PCD) Diagnostic Se sial Antibodies) | les (referred by ervice, Universit | EM Dept CUH to P | rimary Ciliary Dyskines |
| Laboratory: | 12 weeks PCD samp (PCD) Diagnostic Se sial Antibodies) Autoimmune Serolo | les (referred by ervice, Universit | EM Dept CUH to P y Hospital Southa | rimary Ciliary Dyskines mpton) |
| Laboratory: Specimen: | 12 weeks PCD samp (PCD) Diagnostic Se sial Antibodies) Autoimmune Serolo Blood, 4 mL red top | lles (referred by ervice, Universit gy Vacuette (or si | EM Dept CUH to P y Hospital Southan milar container for | rimary Ciliary Dyskines mpton) r clotted blood) |
| Laboratory: | 12 weeks PCD samp (PCD) Diagnostic Se sial Antibodies) Autoimmune Serolo Blood, 4 mL red top Immunfluorescence | gy Vacuette (or si test using Prim | EM Dept CUH to P y Hospital Southan milar container for ate Oesophagus a | rimary Ciliary Dyskines mpton) r clotted blood) s substrate. |
| Laboratory: Specimen: | 12 weeks PCD samp (PCD) Diagnostic Se sial Antibodies) Autoimmune Serolo Blood, 4 mL red top Immunfluorescence Part of Coeliac Scre | gy Vacuette (or si test using Prim | EM Dept CUH to P y Hospital Southan milar container for ate Oesophagus a | rimary Ciliary Dyskines mpton) r clotted blood) |
| Laboratory: Specimen: Comment: | 12 weeks PCD samp (PCD) Diagnostic Se sial Antibodies) Autoimmune Serolo Blood, 4 mL red top Immunfluorescence Part of Coeliac Scre screen. | gy Vacuette (or si test using Prim | EM Dept CUH to P y Hospital Southan milar container for ate Oesophagus a | rimary Ciliary Dyskines mpton) r clotted blood) s substrate. |
| Laboratory: Specimen: | 12 weeks PCD samp (PCD) Diagnostic Se sial Antibodies) Autoimmune Serolo Blood, 4 mL red top Immunfluorescence Part of Coeliac Scre | gy Vacuette (or si test using Prim | EM Dept CUH to P y Hospital Southan milar container for ate Oesophagus a | rimary Ciliary Dyskines mpton) r clotted blood) s substrate. |

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Erythrocyte Membrane Analysis EMA for Hereditary Spherocytosis

| | / / / | | | | | |
|---------------|---|--|--|--|--|--|
| Laboratory: | Specimen referred from Haematology to Haematology, Our Lady's Hospital | | | | | |
| | Crumlin, Dublin 12 | | | | | |
| Specimen: | Blood 3mL, purple, Vacuette® (EDTA) | | | | | |
| | Available Mon to Thurs only, to reach laboratory by 12 noon, Time of | | | | | |
| | phlebotomy must be stated on form. | | | | | |
| Comment: | Requested by Consultant Haematologist | | | | | |
| Turnaround: | 28 working days | | | | | |
| Report: | Sent to referring clinician and copy filed in laboratory | | | | | |
| ENA Screen (E | ENA Screen (Extractable Nuclear Antigens) | | | | | |
| Laboratory: | Autoimmune Serology | | | | | |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) | | | | | |
| Commente | Qualitative Immunoaccovy using Phadia Unicon 250 analyzer, Careening accovy | | | | | |

| Comment: | Qualitative Immunoassay using Phadia Unicap 250 analyser. Screening assay for antibodies to Ro, La, U1RNP, Sm, SCL-70 & Jo-1. Undertaken on all |
|-------------|---|
| | positive ANF sera. |
| Turnaround: | 72 Hours |
| Ref. Range: | Not applicable |

Endocervical Swab

| Refer to Geni | |
|----------------|--|
| | |
| | micularis (Sellotape slide for Pinworm) |
| Laboratory: | Microbiology (Category 3 Laboratory) |
| Specimen: | The specimen is collected first thing in the morning, before the patient has bathed or used the toilet. Apply sellotape to the perianal region, pressing the adhesive side of the tape firmly against the left and right perianal folds several times. Smooth the tape back on the slide, adhesive side down. The sellotape slide should be kept in a slide box in a sealed plastic bag. It is recommended that samples should be taken for at least 4-6 consecutive days. |
| Comment: | Test performed routinely Monday to Friday 9-5pm. Transport specimens ASAP. Do not refrigerate or incubate specimens. Occasionally, an adult worm may be collected from a patient and should be sent in saline or water in a sterile leak-proof universal container for identification. |
| Turnaround: | 24 hours |
| Report: | Enterobius vermicularis ova present or Enterobius vermicularis adult worm present |
| Enterovirus Mo | blecular |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | Faeces (2-5g), viral throat swab, CSF (>0.5mL), vesicular fluid, 4mL clotted blood, 4mL EDTA blood |
| Comment: | Performed by a reference laboratory (National Virus Reference Laboratory (NVRL), Dublin). |
| | Samples positive in enteroviral screen are further tested to determine enterovirus type, which includes echovirus and coxsackie virus. A throat swab is requested for CSF samples positive for enterovirus RNA so that characterisation can be carried out. |
| Turnaround: | 14 working days, additional time required for positive samples |
| Report: | Detected (with characterisation) or not detected |
| | irus (EBV) IgG and IgM |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| | |

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| | | Author: | Mir Paul Calitwell | |
| Comment: | EBV IgM (VCA) per | formed in-house | | |
| | EBV IgG (VCA and | | | erence laboratory |
| | (National Virus Refe | | | |
| Turnaround: | 36 hours for EBV Ig | | | , |
| Report: | Qualitative result | ,, <u> </u> | , | |
| | /irus (EBV) Molecul | ar | | |
| Laboratory: | Microbiology (Infect | | erology) | |
| Specimen: | 4mL EDTA blood | | croidgy) | |
| Comment: | | erence laborator | v (National Virus) | Reference Laboratory |
| comment. | (NVRL), Dublin) | | | |
| Turnaround: | 14 working days | | | |
| Report: | Detected (viral load | I) or not detecte | d | |
| Erythropoieti | | I) of not detecte | u | |
| Laboratory: | | m Clinical Riach | emistry to Eurofin | ns-Biomnis Laboratories |
| Specimen: | Lithium Heparin or | | | ווווטום-פוווווטום-פו |
| Comment: | Consultant request | • • | eu sample). | |
| | • | OTTY | | |
| Turnaround: | 3 weeks | | | |
| Ref. Range: | | | te <u>https://www</u> .euroi | fins.ie/biomnis/ for up to |
| | date referral test in | | | |
| | te Sedimentation R | ate | | |
| Laboratory: | Haematology | | | |
| Specimen: Adult sample: Blood 3mL purple Vacuette [®] EDTA (purple top), volume of sample required for ESR is 1.4 mL. | | | | irple top), Minimum |
| | Paediatric sample: top) | 2 x 1ml EDTA (F | Purple Cap/White I | Ring) or 2 x 1.3 ml (red |
| Comment: | ESR Measurement i | s a non-specific | test of inflammat | ion and tissue damage. |
| | Test available Mon | to Fri during rou | tine working hour | ·S. |
| | ESR is most accura | te when analyse | d within 4 hours o | of phlebotomy. |
| Turnaround: | Urgent specimens: | <2 hours (when | laboratory inforn | ned); |
| | Routine ward spec | | - | - |
| Ref. Range: | Males: 0 - 10mm/ | | 0 – 20mm/hour | |
| Eye Swab | | | · · · | |
| Laboratory: | Microbiology (Main | laboratory) | | |
| Specimen: | | | abs. Any available | e pus should be sampled |
| • | as well as the area | | | |
| | containing transpor | | | |
| | | | | indicate if testing for |
| | Neisseria gonorrhoe | ae is required. | Specific Viral or Cl | nlamydia swabs in |
| | appropriate transpo chlamydial infection | | eded for the diagr | nosis of viral and |
| Comment: | Test performed rout | | Friday 9-5pm or | by urgent request. |
| Turnaround | Prelim: 24 hours; F | • • | | ., |
| : | | | | |
| Report: | Culture report: Any sensitivities. | clinically signific | cant isolate with t | he appropriate |
| Factor I (see F | ibrinogen) | | | |
| Laboratory: | Haematology | | | |

Laboratory: Haematology

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Factor II – see also INR Prothrombin Time

| actor II - see | also TINK FIOL | | | |
|----------------|--|---------------------|------------------------|---------------------------|
| Laboratory: | Haematology | | | |
| Specimen: | Blood 3mL; blue Vacuette® (sodium citrate 3.2%). | | | |
| | Specimens wh | ich are haemolyse | ed, underfilled or ove | rfilled cannot be |
| | analysed, chec | k coagulation san | nple bottles are not e | expired to ensure correct |
| | filling). | | | |
| Comment: | Determines the | e activity of coagu | lation Factor II (Prot | hrombin). |
| | Test available | Monday to Friday | , during routine work | ing hours. |
| | Samples mus | t be received w | ithin 4 hours | |
| Turnaround: | 2 weeks | | | |
| Ref. Range: | Age | Mean (IU/mL) | Range (IU/mL) | |
| | Day 1 | 0.48 | 0.26 - 0.70 | |
| | Day 5 | 0.63 | 0.33 - 0.93 | |
| | Day 30 | 0.68 | 0.34 - 1.02 | |
| | Day 90 | 0.75 | 0.45 - 1.05 | |
| | Day 180 | 0.88 | 0.60 - 1.16 | |
| | Adult – see fin | al report | | |

Factor V (Coagulation/clotting factor)

| Laboratory: | Haematology | | | | |
|-----------------------------------|--|--------------------|----------------------------------|--------------------|--|
| Specimen: | Blood 3mL x 2; blue Vacuette® (sodium citrate 3.2%). | | | | |
| | Specimens that are haemolysed, underfilled or overfilled cannot be ana check coagulation sample bottles are not expired to ensure correct fillin | | | | |
| | | | | | |
| Comment: | Determines the activity of coagulation Factor V. Test available Monday to | | | | |
| | Friday, during | routine working h | iours, by arrangeme | nt with the | |
| | Haematology L | _aboratory. | | | |
| | Samples mus | t be received w | ithin 4 hours | | |
| Turnaround: | 2 weeks | | | | |
| Ref. Range: | Age | Mean (IU/mL) | Range (IU/mL) | | |
| | Day1 | 0.72 | 0.36 - 1.08 | | |
| | Day 5 | 0.95 | 0.45 - 1.45 | | |
| | Day 30 | 0.98 | 0.62 - 1.34 | | |
| | Day 90 | 0.90 | 0.48 - 1.32 | | |
| | Day 180 | 0.91 | 0.55 - 1.27 | | |
| | Adult | 1.06 | 0.62 - 1.50 | | |
| Factor V Leiden Mutation (G1691A) | | | | | |
| Laboratory: | Haematology N | Molecular Genetics | 5 | | |
| Specimen: | Blood 3mL x 2 | purple Vacuette | e [®] (EDTA) N.B. Separ | ate EDTA sample | |
| | necessary if FE | 3C also requested | , citrate specimen als | o required for APC | |
| | Resistance | | | | |

Comment: If the APC Resistance screening test for Factor V Leiden (which forms part of the thrombophilia screen) is positive it is confirmed by PCR analysis in the Haematology Genetics laboratory. See Main Haematology Section on Guidelines for Investigation of

Thrombophilia. Thrombophilia request form FOR-CUH-PAT-1575, including documentation of patient consent, must be received with all requests and is available on the CUH website.

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Factor VII (Coagulation/clotting factor)

| | aguiación/ ci | otting factor j | | |
|----------------|--|----------------------|------------------------|----------------------------|
| Laboratory: | Haematology | | | |
| Specimen: | Blood 3mL x 2; blue Vacuette® (sodium citrate 3.2%). | | | |
| | Specimens t | hat are haemolyse | d, underfilled or over | filled cannot be analysed, |
| | check coagu | lation sample bottl | es are not expired to | ensure correct filling. |
| Comment: | Determines | the activity of coag | ulation Factor VII. Te | est available Monday to |
| | Friday, durir | ng routine working | hours, by arrangem | ent with the |
| | Haematolog | y Laboratory. | | |
| | Samples m | ust be received v | vithin 4 hours | |
| Turnaround: | 2 weeks | | | _ |
| Ref. Range: | Age | Mean (IU/mL) | Range (IU/mL) | |
| | Day 1 | 0.66 | 0.28 - 1.04 | |
| | Day 5 | 0.89 | 0.35 - 1.43 | |
| | Day 30 | 0.90 | 0.42 - 1.38 | |
| | Day 90 | 0.91 | 0.39 - 1.43 | |
| | Day 180 | 0.87 | 0.47 - 1.27 | |
| | Adult | 1.05 | 0.67 - 1.43 | |
| actor VIII (Co | pagulation/g | lotting factor) | | |

Factor VIII (Coagulation/clotting factor)

| | | <u> </u> | | | |
|----------------|--|----------------------|----------------------|-----------------------------|--|
| Laboratory: | Haematology | Haematology | | | |
| Specimen: | Blood 3mL x 2; blue Vacuette [®] (sodium citrate 3.2%). | | | | |
| | Specimens th | nat are haemolysed | , underfilled or ove | rfilled cannot be analysed, | |
| | check coagul | ation sample bottle | s are not expired to | o ensure correct filling. | |
| Comment: | Determines t | he activity of coagu | Ilation Factor VIII. | Test available Monday to | |
| | | | - | urs, emergency requests | |
| | | , , , | ires prior Haemato | ology Consultant approval | |
| | and planning | | | | |
| | - | ist be received wi | | | |
| Turnaround: | | pecimens < 4hours | ; | | |
| | Routine spec | imens 14 days. | | 1 | |
| Ref. Range: | Age | Mean (IU/mL) | Range (IU/mL) | | |
| | Day 1 | 1.14 | 0.50 - 1.78 | | |
| | Day 5 | 1.02 | 0.50 - 1.54 | | |
| | Day 30 | 1.03 | 0.50 - 1.57 | | |
| | Day 90 | 0.87 | 0.50 - 1.25 | | |
| | Day 180 | 0.79 | 0.50 - 1.09 | | |
| | Adult | 0.99 | 0.50 - 1.49 | | |
| Factor VIII Ch | romogenic (C | Coagulation/clotti | ng factor) | | |
| Laboratory: | | | | n Laboratory, St James | |
| | | | | to Haematology Dept., | |
| | Our Lady's H | ospital, Crumlin, D | ublin 12) | | |
| Specimen: | Blood 3mL x | 2; blue Vacuette® | (sodium citrate 3.2 | 2%). | |
| | Specimens th | nat are haemolysed | , underfilled or ove | rfilled cannot be analysed, | |
| | check coagul | ation sample bottle | s are not expired to | o ensure correct filling. | |
| Comment: | By arrangem | ent with laboratory | | | |
| | Samples mu | ist be received wi | ithin 4 hours | | |
| | | | | | |

- Turnaround: 84 working days Ref. Range: Adults (>18 years) 0.55 – 1.77 IU/ml
- Report: Sent to referring clinician and copy filed in laboratory

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| hromogenic | | | | |
|--|--|--|--|--|
| Haematology | | | | |
| • / | 2: blue Vacuette® | (sodium citrate 3.2% |). | |
| Specimens that are haemolysed, underfilled or overfilled cannot be analysed, | | | | |
| | | | | |
| • | • | • | 2 | |
| | | | | |
| | | | | |
| | | p | | |
| | st be received wi | thin 4 hours | | |
| | | | | |
| . | | | | |
| • | • | | | |
| | | | | |
| | | | | |
| • | | (sodium citrate ? 20 | () | |
| | | - | - | |
| | | | | |
| - | - | - | - | |
| | | | | |
| | | | sons by an angement | |
| | • · | • | | |
| - | | | | |
| - , | • | ins (by an angement) | 1 | |
| | | Pango (ILI/mL) | 1 | |
| | | | | |
| | | | - | |
| | | | - | |
| | | | - | |
| | | | - | |
| | | | - | |
| Adult | 1.09 | 0.55 - 1.63 | | |
| gulation/clot | ting factor) | | | |
| Haematology | / | | | |
| Blood 3mL x | 2; blue Vacuette® |) (sodium citrate 3.2% | ///). | |
| Specimens t | hat are haemolysed | d, underfilled or overf | filled cannot be analysed, | |
| check coagu | lation sample bottle | es are not expired to | ensure correct filling. | |
| Determines | the activity of coag | ulation Factor X. Test | available Monday to | |
| Friday, durin | g routine working | hours, by arrangem | ent with the | |
| | • | | | |
| - | ust be received w | ithin 4 hours | | |
| 2 weeks | | | | |
| Age | Mean (IU/mL) | Range (IU/mL) | | |
| Day 1 | 0.44 | 0.21 - 0.68 | | |
| Day 5 | 0.49 | 0.19 - 0.79 | | |
| | 0.59 | | | |
| Day 90 | 0.67 | 0.35 - 0.99 | | |
| | | | | |
| Day 180 | 0.71 | 0.35 - 1.07 | | |
| | Haematology Blood 3mL x 2 Specimens the check coagula Determines the Friday by arra of routine hou planning. Samples mus Emergency sp Routine specif 0.72 – 1.61 IU agulation/cloo Haematology Blood 3mL x Specimens the check coagul Determines the during routine with the Haee Samples mus Emergency sp Routine specif Day 1 Day 5 Day 30 Day 90 Day 180 Adult gulation/clott Haematology Blood 3mL x Specimens the check coagul Day 180 Adult gulation/clott Haematology Blood 3mL x Specimens the check coagul Determines the friday, durin Haematology Samples mus 2 weeks Age Day 1 Day 5 Day 30 | HaematologyBlood 3mL x 2; blue Vacuette®Specimens that are haemolysed, check coagulation sample bottlesDetermines the activity of coaguFriday by arrangement, during r of routine hours always requires planning.Samples must be received wiEmergency specimens < 4hours Routine specimens 14 days.0.72 - 1.61 IU/mlagulation/clotting factor)HaematologyBlood 3mL x 2; blue Vacuette® Specimens that are haemolysed check coagulation sample bottleDetermines the activity of coag during routine working hours at with the Haematology LaboratorSamples must be received w Emergency specimens: 2 weeks.AgeMean (IU/mL)Day 10.53Day 300.51Day 900.67Day 1800.86Adult1.09gulation/clotting factor)HaematologyBlood 3mL x 2; blue Vacuette® Specimens that are haemolysed check coagulation sample bottleDay 10.44Day 300.51Day 1000.44Day 10.44Day 50.49Day 300.59 | Haematology Blood 3mL x 2; blue Vacuette® (sodium citrate 3.2% Specimens that are haemolysed, underfilled or overfil check coagulation sample bottles are not expired to e Determines the activity of coagulation Factor VIII. Te Friday by arrangement, during routine working hours of routine hours always requires prior Haematology C planning.Samples must be received within 4 hours Emergency specimens < 4hours; Routine specimens 14 days. $0.72 - 1.61$ IU/mlImagulation/clotting factor) Haematology Blood 3mL x 2; blue Vacuette® (sodium citrate 3.29 Specimens that are haemolysed, underfilled or overficheck coagulation sample bottles are not expired to Determines the activity of coagulation Factor IX. Test during routine working hours and for emergency real with the Haematology Laboratory.Samples must be received within 4 hours Emergency specimens < 24hours (by arrangement), Routine specimens: 2 weeks.AgeMean (IU/mL) Range (IU/mL) Day 10.510.21 - 0.81 Day 300.510.21 - 0.81 Day 30Day 900.670.21 - 1.13 Day 1800.86Day 1000.860.36 - 1.36 AdultHaematology Blood 3mL x 2; blue Vacuette® (sodium citrate 3.29 Specimens that are haemolysed, underfilled or overficheck coagulation sample bottles are not expired to Determines the activity of coagulation Factor X. Test Friday, during routine working hours, by arrangem Haematology Blood 3mL x 2; blue Vacuette® (sodium citrate 3.29 Specimens that are haemolysed, underfilled or overficheck coagulation sample bottles are not expired to Determines the activity of coagulation Factor X. Test Friday, during routine working hours, by arrangem Haematology Laboratory.Samples must be received within 4 hours | |

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Factor XI (Coagulation/clotting factor)

| Factor XI (Coa | gulation/clo | tting factor) | | | | | |
|----------------|--|----------------------|----------------------------|------------------------|--|--|--|
| Laboratory: | Haematology | | | | | | |
| Specimen: | Blood 3mL x 2; blue Vacuette® (sodium citrate 3.2%). | | | | | | |
| | Specimens t | hat are haemolysed | d, underfilled or overfill | ed cannot be analysed, | | | |
| | check coagu | lation sample bottle | es are not expired to er | sure correct filling. | | | |
| Comment: | Determines | the activity of coag | ulation Factor X1 Test | available Mon to Fri, | | | |
| | | , . | gement with the Haen | • | | | |
| | Samples must be received within 4 hours | | | | | | |
| Turnaround: | 2 weeks | | | | | | |
| Ref. Range: | Age | Mean (IU/mL) | Range (IU/mL) | | | | |
| | Day 1 | 0.38 | 0.10 - 0.66 | | | | |
| | Day 5 | 0.55 | 0.23 - 0.87 | | | | |
| | Day 30 | 0.53 | 0.27 - 0.79 | | | | |
| | Day 90 0.69 0.41 - 0.97 | | | | | | |
| | Day 180 | 0.91 | 0.49 - 1.34 | | | | |
| | Adult | 0.97 | 0.67 - 1.27 | | | | |
| Factor XII (Co | agulation/cl | otting factor) | | | | | |
| Laboratory: | Haematolog | ¥ | | | | | |
| Specimen: | Blood 3mL x 2; blue Vacuette® (sodium citrate 3.2%). | | | | | | |
| | <u> </u> | | | | | | |

Specimens that are haemolysed, underfilled or overfilled cannot be analysed, check coagulation sample bottles are not expired to ensure correct filling.
 Determines the activity of coagulation Factor X11. Test available Mon to Fri, during routine hours, by arrangement with the Haematology Laboratory.

Samples must be received within 4 hours Turnaround: 2 weeks

| Turnaround: | 2 weeks | | |
|-------------|-----------------|------|---------------|
| Ref. Range: | Ref. Range: Age | | Range (IU/mL) |
| | Day 1 | 0.53 | 0.13 - 0.93 |
| | Day 5 | 0.47 | 0.11 - 0.83 |
| | Day 30 | 0.49 | 0.17 - 0.81 |
| | Day 90 | 0.67 | 0.25 - 1.09 |
| | Day 180 | 0.77 | 0.39 - 1.15 |
| | Adult | 1.08 | 0.52 - 1.64 |

Factor XIII (Coagulation/clotting factor)

| Laboratory: | Haematology |
|----------------|---|
| Specimen: | Blood 3mL x 2; blue Vacuette [®] (sodium citrate 3.2%). |
| Comment: | Specimens that are haemolysed, underfilled or overfilled cannot be analysed, check coagulation sample bottles are not expired to ensure correct filling. A qualitative assay to diagnose congenital deficiency. Test available Mon – Thurs,(due to incubation requirements) during routine hours. Samples must be received within 4 hours |
| Turnaround: | 3 weeks |
| Ref. Range: | Normal/Abnormal clot detected, Low level detected |
| Faecal Elastas | e |
| Laboratory: | Referred from Biochemistry to City Hospital, Birmingham |
| Specimen: | Minimum 5g stool |
| Turnaround: | 2 Weeks |
| Ref. Range: | See report form |

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Faeces – Molecular Analysis and Culture

| Laboratory: | Microbiology (Category 3 Laboratory) |
|--------------|---|
| Specimen: | Faeces sample for molecular analysis of Salmonella spp., Shigella spp., |
| | Campylobacter spp. Verotoxin (VT1 and / VT2; markers of |
| | enterohaemorrhagic disease), Cryptosporidium parvum/hominis and Giardia lamblia. |
| | The specimen should be collected into a clean, sterile, leakproof container. Ideally, all specimens should be taken as soon as possible after onset of symptoms. Transport specimens ASAP. If processing is delayed, refrigeratio is preferable to storage at ambient temperature. A number of important pathogens such as <i>Shigella</i> species may not survive the pH changes that occur in faeces specimens that are not promptly delivered to the laboratory, even if refrigerated. |
| | Samples >72hrs old on receipt in the laboratory are unsuitable for testing. Hospital inpatient samples are not routinely retested for 14 days if they are |
| Comment: | continually in hospital for this period. Rectal swabs are not suitable. Full clinical information should be provided, |
| comment. | esp. presence and duration of symptoms, recent foreign travel or shellfish ingestion and previous antibiotics. |
| | Clearance samples for Salmonella, Shigella and Campylobacter not routinely |
| | processed unless clinically indicated. Please discuss with Microbiology Medic team. |
| Turnaround: | <36hours for preliminary result |
| | Clinically significant isolates are telephoned when available. |
| | Confirmatory culture results are sent to referral labs and may take up to 4 weeks |
| Report: | Report presence of specific pathogen and absence of other pathogens (Target Not Detected or Target Detected). Faeces are cultured on selective media for <i>Salmonella and Shigella</i> when positive by molecular testing. Verotoxigenic positive samples are sent to Cherry Orchard Reference laboratory for confirmation. |
| | In addition, when clinically indicated, specific media for <i>Yersinia</i> spp. And <i>Vibrio</i> sp will be inoculated. Where appropriate i.e. HUS the specimen is set to Cherry Orchard Hospital lab for detailed analysis of various enterohaemorrhagic <i>E. coli</i> |
| | A Target Not Detected result does not automatically exclude infection from the above enteric pathogens as the level of DNA present may be lower than the limit of detection of the assay. |
| | Please refer to individual sections for <i>Clostridioides difficile</i> testing, <i>Cryptosporidium Sp.</i> Parasitology and Rotavirus /Adenovirus antigens. |
| llopian Tube | Aspirate / Tubo-ovarian Fluid |
| | |

| Fanconi's Anae | Fanconi's Anaemia | | | | |
|----------------|---|--|--|--|--|
| Laboratory: | Referred from Biochemistry to Bristol Genetics Lab | | | | |
| Specimen: | 5ml Lithium Heparin blood/bone marrow in Lithium Heparin | | | | |
| | Paediatrics – at least 1ml lithium heparin (preferably 2ml) | | | | |
| Comment: | 24hrs notice required to facilitate courier arrangements (Contact ext 22531). | | | | |
| | Request form available at <u>www.nbt.nhs.uk/genetics</u> | | | | |
| Turnaround: | 28 days | | | | |
| | | | | | |

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| Farmer's Lung | Antibodies | | | |
|-----------------|--|--|--|--|
| Laboratory: | Microbiology (Infectious Diseases Serology) | | | |
| Specimen: | 4mL clotted blood | | | |
| Comment: | Performed by a reference laboratory (Mycology Reference Centre, Leeds) | | | |
| Turnaround: | 28 working days | | | |
| Report: | Quantitative result with an interpretative comment | | | |
| Ferritin | | | | |
| Laboratory: | Haematology | | | |
| Specimen: | Blood 4mL Red Vacuette [®] (clotted blood). | | | |
| Comment: | The level of serum ferritin correlates well with the body iron reserves under various physiological and pathological conditions. Ferritin is an acute phase reactant. | | | |
| | Test available Monday to Friday, during routine working hours. Exceptions to this may be available for Covid 19 screening with prior arrangement. Ferritin should be requested for investigation of abnormal FBC results and relevant clinical syndromes. | | | |
| | Use of haematinics for screening of well patients is not recommended. Requests should be accompanied by clinical details. | | | |
| | See BCSH guidelines. | | | |
| | Laboratory Diagnosis of Functional Iron Deficiency http://onlinelibrary.wiley.com/doi/10.1111/bjh.12311/pdf | | | |
| Turnaround: | 7 working days | | | |
| Ref. Range: | Females 11 – 307 ng/ml, Males 23.9 – 336.2 ng/ml | | | |
| Ken Kunge. | These are ADULT ranges – for guidance only | | | |
| Fertility Scree | | | | |
| Laboratory: | Microbiology (Infectious Diseases Serology) | | | |
| Specimen: | 4mL clotted blood | | | |
| Tests: | Hepatitis B surface antigen, anti-HBcore, HIV Ag/Ab, anti-HCV | | | |
| Turnaround: | Negative samples: 36 hours. Please allow extra time for samples testing positive in house for HIV Ag/Ab and anti-HCV (external confirmatory testing required). | | | |
| Report: | Qualitative result | | | |
| Fibrinogen (Fa | ctor 1) | | | |
| Laboratory: | Haematology | | | |
| Specimen: | Blood 3mL; blue Vacuette [®] (sodium citrate 3.2%). | | | |
| | Specimens which are haemolysed, underfilled or overfilled cannot be analysed, check coagulation sample bottles are not expired to ensure correct filling) | | | |
| | Specimens must be received within 12 hours of phlebotomy. | | | |
| Comment: | Determines the concentration of plasma fibrinogen. Forms part of a Thrombophilia and/ or Lupus screen, see Main Haematology Section on Guidelines for Investigation of Thrombophilia. Test available Monday to Friday, during routine working hours, and for emergency reasons at all other times. | | | |
| Turnaround: | Emergency specimens: 2 hours by arrangement with the laboratory; Routine specimens: 8 hours, if part of Thrombophilia 3 – 4 weeks | | | |
| Ref. Range: | Age Mean(g/L) Range g/L | | | |
| | Day 1 2.9 1.7 – 4.0 | | | |
| | Day 5 3.2 1.6 – 4.7 | | | |
| | Day 30 2.7 1.6 – 3.8 | | | |
| | Day 90 2.5 1.1 – 3.8 | | | |
| | Day 180 2.6 1.2 – 3.9 | | | |
| | | | | |

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| | Adult 2.9 1.7 – 4.1 |
|-----------------|---|
| Fibrinogen Ph | enotyping and Genetic Analysis |
| Laboratory: | Sample referred from Haematology to the DNA Laboratory, St., Thomas's Hospital, London |
| Specimen: | Blood 3 mL purple Vacuette [®] (EDTA) and Blood 3ml; blue Vacuette [®] (sodium citrate 3.2%), fill to mark on tube. |
| Comment: | Request must be booked in advance with the Haematology Laboratory CUH, performed in the investigation of Dysfibrinogenanaemia |
| Turnaround: | 80 working days |
| Report: | Sent to referring clinician and copy filed in laboratory |
| Filaria Antiboo | lies |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Comment: | Performed by a reference laboratory (National Parasitology Reference Laboratory (NPRL), London) |
| Turnaround: | 28 working days |
| Report: | Qualitative result |
| Fluroesence/C | Chromogenic In-Situ Hybridisation (FISH/CISH) (Tissue) |
| Laboratory: | Referred by Pathology to CMD, St James Hospital. |
| Specimen: | FFPE tissue block |
| Comment: | See St.James Lab User Handbook (available online) for available targets. |
| Turnaround: | 14 working days (from date testing material is sent for referral) |
| | n-Situ Hybridisation FISH, for the diagnosis of AML, CML, ALL, MDS, oma, Plasmocytoma. |
| Laboratory: | Referred from Haematology to Munich Leukaemia Laboratory (MLL MVZ |
| | GmbH), Germany |
| Specimen: | 2-3 ml bone marrow aspirate or peripheral blood are sufficient in case of normal cellularity |
| Comment: | Must arrange with Haematology, transport within 24 hours, complete form from referral laboratory |
| Turnaround: | 1-3 working days (excluding transport time) |
| Report: | Sent to referring clinician and copy filed in laboratory |
| | n-Situ Hybridisation (FISH) for Microdeletions Syndromes (eg. Di |
| George, Willia | |
| Laboratory: | Specimen referred from Molecular Genetics Lab in Biochemistry to NCMG. |
| Specimen: | Adults: 2ml Lithium Heparin blood. Infants: 1ml min Lithium Heparin blood) DO NOT refrigerate specimens. |
| Comment: | NCMG request form available from www.genetics.ie |
| Turnaround: | See NCMG website |
| Report: | Sent to referring clinician from NCMG and copy of report filed in pathology |
| Flow Cytomet | ry |
| Laboratory: | Haematology |
| Specimen: | Fresh Blood or Bone Marrow – 3mL, purple Vacuette (EDTA). Samples may be refrigerated overnight. Optimal sample age less than 24 hours. |

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| Comment: | | amples must b | e transported dired | ctly to the laboratory |
| | immediately. | tool in identify | ving louksemise T | est available Mon to |
| | during routine hours | | - | |
| | Please state specime | | | |
| | essential clinical info | | | |
| | consultant haematol | | | |
| | | 5 | ng out of hours an | d at weekends, wher |
| | | | | y rely on a diagnosti |
| | flow report, the Con | | | |
| | staff to facilitate suc | | 5 | , , |
| Turnaround: | Routine specimens: | 72 hours | | |
| | Urgent specimens: 2 | 24 hours | | |
| Ref. range: | Refer to final report | | | |
| Foetal Genotyp | e | | | |
| Laboratory: | Available by prior ar | rangement wit | h Blood Transfusio | n Laboratory |
| Specimen: | 16mL EDTA materna | al | | - |
| | 3mL EDTA paternal | | | |
| Comment: | If possible, 24 hours | notice to Bloo | d Transfusion Labo | oratory, CUH required |
| | (Contact Ext 22537) | | | |
| | | | | ting clinician (Availa |
| | from Blood Transfusion Laboratory). | | | |
| | Samples referred to: IBGRL, Bristol, United Kingdom via IBTS. | | | |
| | NOTE: Foetal Sex Typing is NOT referred by the Blood Transfusion | | | |
| | Laboratory, CUH. | | | |
| Turnaround: | 21 Working Days | | | |
| Foetal DNA Rh | | | | |
| Laboratory | Blood Transfusion | | | |
| Specimen: | 1 x 6ml EDTA | | | 6 |
| Comment: | This test available si | | | |
| | (International Blood | | rence Laboratory, | Bristol, UK) |
| | Minimum gestation | 11 weeks + 2. | | |
| Turnersende | | | | |
| Turnaround: | 3 weeks | | | |
| Ref. Range: | 3 weeks Rh D Positive; Rh D | |) Inconclusive | |
| Ref. Range: Flecanide | Rh D Positive; Rh D | Negative; Rh [| | |
| Ref. Range: | Rh D Positive; Rh D | Negative; Rh [| | 's University Hospita |
| Ref. Range: Flecanide | Rh D Positive; Rh D Referred from Clinic | Negative; Rh [al Biochemistry | | 's University Hospita |
| Ref. Range: Flecanide Laboratory: | Rh D Positive; Rh D Referred from Clinic London. Serum (Trough sam Toxicity may occur a | <u>Negative; Rh [</u> al Biochemistry ple) | to ASI, St George | |
| Ref. Range: Flecanide Laboratory: Specimen: | Rh D Positive; Rh D Referred from Clinic London. Serum (Trough sam Toxicity may occur a trough sample. | <u>Negative; Rh [</u> al Biochemistry ple) | to ASI, St George | |
| Ref. Range: Flecanide Laboratory: Specimen: Comment: Turnaround: | Rh D Positive; Rh D Referred from Clinic London. Serum (Trough sam Toxicity may occur a | <u>Negative; Rh [</u> al Biochemistry ple) | to ASI, St George | 's University Hospita d is appropriate for a |
| Ref. Range: Flecanide Laboratory: Specimen: Comment: | Rh D Positive; Rh D Referred from Clinic London. Serum (Trough sam Toxicity may occur a trough sample. | <u>Negative; Rh [</u> al Biochemistry ple) | to ASI, St George | |

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Foetal Maternal Haemorrhage FMH by Flow Cytometry ≥4 mls bleed

| Laboratory: | Referred by Haematology to the Rotunda Hospital, Parnell St, Dublin 1 |
|-------------|--|
| Specimen: | 3ml EDTA specimen |
| Comment: | All postnatal samples with bleeds \geq 4mls are referred to the Rotunda for flow |
| | cytometry. Antenatal patients with bleeds \geq 4mls are NOT referred. Flow |
| | cytometry in Rotunda is currently not validated for antenatal patients. |
| | Kleihauer on a rhesus D Neg mother of a baby with a weak D Ag are NOT |
| | referred. |
| Turnaround: | 14 working days for the hard copy report: It is practice of the referral |
| Report: | laboratory to give a verbal report as soon as possible. |
| | Sent to clinician and copy filed in laboratory |

Foetal Sex Typing

| | ····· 9 |
|----------------|--|
| Laboratory: | Referred from Biochemistry to IBGRL, Bristol. Prior notice required to |
| | facilitate courier arrangements (Contact ext 22531) |
| Specimen: | 16mL EDTA maternal |
| | 3mL EDTA paternal |
| Comment: | Pregnancy must be at least 7 weeks |
| | IBGRL request form (FM4739) to be completed by referring clinician |
| Turnaround: | 5 working days from recipt of specimen in Bristol |
| Foetus – First | Trimester |

Foetus – First Trimester

| Laboratory: | Histopathology (Diagnostic Laboratory) |
|-------------|---|
| Comment: | If pre-viable foetal tissue (however small) is identified following delivery, the <i>Consent to Pathological Examination of a pre- 16 week foetus</i> form (form 453) must be completed in full by the doctor or midwife, signed by the parent , and submitted to the Histopathology laboratory with a completed Histopathology Request Form. For full details of the protocol contact the Histopathology laboratory at (021) 4922792 |
| | |

Foetus – Post First Trimester

See Autopsies/Post-Mortems under HISTOPATHOLOGY

Folate (serum)

| Laboratory: | Haematology |
|-------------|--|
| Specimen: | Blood 4mL Red, Vacuette [®] (clotted blood). |
| Comment: | Forms part of the investigation of Megaloblastic Anaemia. |
| | Please note that international studies have indicated that folic concentrations < 4 ng/mL may be associated with deficiency. Therefore results < 4 ng/mL should be subject to clinical as well as laboratory interpretation. |
| | Test available Monday to Friday, during routine working hours. |
| | B12 and Folate should be requested for investigation of abnormal FBC results |
| | and relevant clinical syndromes. |
| | Use of haematinics for screening of well patients is not recommended. |
| | Requests should be accompanied by clinical details. |
| | See BCSH guidelines. |
| | The diagnosis of B12 and folate deficiency |
| | http://onlinelibrary.wiley.com/doi/10.1111/bjh.12959/pdf |
| | 7 working days |
| Ref. Range: | 3.1 – 20 ng /mL |

These are ADULT ranges – for guidance only

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Formalin Fixed Histopathology Specimens

| Laboratory: | Histopathology |
|-------------|---|
| Specimen: | Formalin fixed Tissues for Histopathology |
| | See separate entries for |
| | Direct Immunofluorescence |
| | Skin/Oral mucosa, |
| | Electron Microscopy, |
| | Frozen Sections, |
| | Liver Biopsy for Copper/Iron Estimation, |
| | Renal Biopsy |
| Comment: | Specimens should be placed in a container, large enough to contain adequate |
| | Buffered Formalin for fixation (recommend ratio of at least 2:1 for Buffered |
| | Formalin volume: specimen size). Ideally all specimens should be submitted |
| | intact to allow accurate gross examination. <i>Tissue should not be removed</i> |
| | from the specimen, for research purposes or otherwise, without prior |
| | consultation with a Pathologist as this may compromise accurate |
| | diagnosis. Where specimens are orientated by/with sutures etc, their |
| | designation should be clearly detailed on the accompanying Request |
| | Form. |
| | Pathologists are available for discussion of Histopathology cases, both pre |
| | and post receipt within the laboratory. |
| | Urgant Chasimana, Whare special deemed urgant by the clinician this must |

Urgent Specimens: Where case is deemed urgent by the clinician, this must be clearly indicated on the Request Form.

The Histopathology laboratory does not operate an out-of-hours service. However a consultant pathologist is on-call and may be contacted through the main hospital switchboard, Ph. 021-4922424/4922100

For special Consideration

• Breast Needle Core Biopsy Calcified and Non-Calcified.

Immediately place in Buffered Formal Saline and please state date and time specimen taken. To facilitate subsequent microscopic location of calcified deposits, breast needle core biopsies should be divided into calcified and non-calcified cores when the biopsies are taken.

Note: A separate form is required for biopsies taken from the right and left side. Non-calcified cores should be placed in yellow mesh cassettes which are then placed into a labelled Formalin pot. Calcified cores should be placed in orange mesh biopsy cassettes which are which are then placed into a labelled Formalin pot.

- **Cardiothoracic specimens** must be delivered directly to the Histopathology Laboratory reception without delay. Prolonged fixation may adversely affect subsequent laboratory test results. Optimal fixation times:
 - Small biopsy samples 6 12 hours
 - Larger surgical specimens 8-18 hours.

Lung resection specimens are inflated upon receipt to assist penetration of fixative; delay in delivery adversely affects inflation and fixation.

- **Placenta:** With complicated monochorionic twins where injection studies might be required please discuss with the Histopathology Laboratory before putting placenta into Formalin.
- **Products of Conception:** The 'Fetal Tissue in early pregnancy loss' information leaflet (EXT-CUH-PATH-1201) should be provided to the patient when products of conception tissue is sent to pathology.
- **Renal Biopsy:** See separate entry for Renal biopsy

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Turnaround: The Histopathology <u>NQI</u> Programme divides Histopathology specimens into 4 categories within which TATs are analysed.

| | | NQI Target TAT |
|-----|-----------------------|-------------------------------------|
| P01 | small biopsies | 80% in 5 working days |
| P02 | GI biopsies | 80% in 7 working days or 100% in 10 |
| | | working days |
| P03 | Cancer resections | 80% in 7 working days |
| P04 | Non-Cancer resections | 80% in 7 working days |
| | | |

Our aim is to meet the NQI target TATs for all **urgent** cases. For routine cases, we have subcategorised specimens according to speciality.

Presently the Histopathology Department are not meeting the NQI Target TAT for some routine cases. A realistic CUH Target TAT has been published in the table below. A process is in place to address the staffing and resource deficits in the laboratory as we work towards achieving NQI Target TAT for all sample types.

| Туре | P Code | NQI TAT (working | CUH TAT |
|---|--------|------------------|-----------------|
| | | days)* | (working days)* |
| Breast | P01 | 5 | 5 |
| | P03 | 7 | 7 |
| | P04 | 7 | 7 |
| GIT biopsies | P02 | 7 | 13 |
| | | 10 | 10 |
| Upper GIT | P01 | 5 | 7 |
| | P03 | 7 | 12 |
| | P04 | 7 | 13 |
| Lower GIT | P03 | 7 | 12 |
| | P04 | 7 | 12 |
| Skin | P01 | 5 | 14 |
| | P03 | 7 | 12 |
| | P04 | 7 | 12 |
| Cardiothoracic | P01 | 5 | 5 |
| | P03 | 7 | 10 |
| | P04 | 7 | 10 |
| ENT | P01 | 5 | 10 |
| | P03 | 7 | 13 |
| | P04 | 7 | 11 |
| Cervical cases | P01 | 5 | 10 |
| | P04 | 7 | 14 |
| Gynae cases | P01 | 5 | 12 |
| | P03 | 7 | 10 |
| | P04 | 7 | 12 |
| PNP - POC and Ectopic Pregnancy related specimens | P04 | 7 | 8 |
| PNP - Placenta | P04 | 7 | 40 |
| Bone Marrow biopsy | P04 | 7 | 10 |
| GU | P01 | 5 | 8 |
| | P03 | 7 | 10 |
| | P04 | 7 | 8 |
| Prostate needle biopsy | P01 | 5 | 10 |

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*Please Note:

The following factors may impact stated TAT

- Requirement for ancillary testing to include levels, IHC, Special stains and Molecular Pathology
- Intradepartmental consultation
- Requirement for decalcification
- Large number of blocks required on case.
- Some larger specimens requiring longer fixation (48-72hrs)

Fragile X Syndrome (FRAX)

| Referred from Molecular Genetics Lab in Biochemistry to NCMG. |
|---|
| Infant: 1ml EDTA & 1ml Lithium Heparin bloods |
| Adults: 3-5mls EDTA & 2mls Lithium Heparin bloods |
| Both blood types required as both DNA analysis and karyotype peformed. |
| NCMG request form available from website, <u>www.genetics.ie</u> |
| Up to 6 months |
| Sent to referring clinician and copy of report filed in pathology |
| rensis Antibodies |
| Microbiology (Infectious Diseases Serology) |
| 4mL clotted blood |
| Performed by a reference laboratory (Rare & Imported Pathogens Laboratory |
| (RIPL), Porton Down) |
| 28 working days |
| Qualitative result |
| nt Chains (SFLC)-Kappa and Lambda |
| Clinical Biochemistry |
| 4.0 mL blood in plain tube |
| Contact Immunology on Ext. 22535 if Urgent Free Light Chain required. |
| 10 days |
| Up-to-date reference intervals will be applied to all Biochemistry reports as |
| appropriate |
| ids (FFA) |
| Sample referred from Clinical Biochemistry to the Department of Clinical |
| Chemistry and Newborn Screening, Sheffield |
| 1.2 ml Fluoride oxalate plasma |
| 4 weeks |
| See report form |
| kine) |
| Clinical Biochemistry |
| 4.0 mL blood in plain tube (clotted sample) |
| 4 Days |
| Up-to-date reference intervals will be applied to all Biochemistry reports as |
| appropriate |
| thyronine) |
| Clinical Biochemistry |
| 4.0 mL blood in plain tube |
| 4 Days |
| |
| |

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Frozen Sections (Intraoperative Consultation-Urgent), Neurosurgery

| Laboratory: | Neuropathology |
|------------------------------|--|
| Specimen: | Fresh tissue (universal precautions) |
| Comment | Routine service is available 9:00am to 5:00pm Monday - Friday. Please refe |
| | to the protocol for frozen section (Neuropathology Department information |
| | for Users). Cases to be arranged between the Neurosurgeon and |
| | Neuropathologist. Please contact extension 22520. Theatre rings |
| | Neuropathology Department (ext 22519/22520) at the time the specimen is |
| | being sent. Theatre Nurse brings the specimen to Theatre Reception Area. |
| | Specimen is given to the Porter on Call, who signs the Specimen Book. The Porter brings the specimen in the appropriate container <u>directly</u> to a staff |
| | member in the Neuropathology Department. |
| | |
| | Universal safety precautions must apply. Fresh nervous system tissue |
| | requires special precautions in high risk cases. These include suspected prion |
| | diseases, and other transmissible diseases e.g. tuberculosis, HIV. Label |
| | specimen container and request form with Biohazard sticker. Please contact |
| | the Neuropathologist on duty in advance. |
| | Neuropathology Department logs receipt of the specimen and returns the box |
| | to the Porter. |
| | An urgent on-call service is available outside of these hours on weekdays and |
| | a limited on-call at certain weekends only. Cases should be arranged in |
| | advance between the Neurosurgeon and the Neuropathologist on call |
| | (contact switch). |
| Turnaround: | 20 minutes. Result is telephoned back to theatre. |
| rozen Section Laboratory: | Histopathology (Diagnostic Laboratory) |
| Specimen: | Fresh tissue |
| Comment: | The Frozen Section service is available Mon –Fri 8am to 4pm |
| comment. | Outside of these hours if a frozen section is anticipated, the case must be |
| | discussed with a pathologist (after 5pm the case must be discussed with the |
| | pathologist on-call who may be contacted through the hospital switchboard). |
| | If the fresh specimen poses a health risk to laboratory personnel (e.g. TB, |
| | HIV), <u>frozen analysis should not be undertaken.</u> Alternative approaches to |
| | rapid diagnosis may be discussed with Pathologist/Senior Medical Scientist. |
| | Booking: |
| | Frozen sections Monday – Friday, should be booked in advance where |
| | possible (preferably 24hrs before elective surgery). The Histopathology |
| | laboratory should be contacted at ext. 22792 with the following details. |
| | Date and Time schedule / Patient name /Theatre /Surgeon / Specimen type. |
| | Note: if the frozen section is delayed or cancelled please notify the |
| | Histopathology laboratory at ext. 22792. |
| - · | Unfixed tissue for frozen section must be transported directly to the |
| Transport: | |
| Transport: | laboratory immediately in a correctly labelled dry container, accompanied by |
| Transport: | laboratory immediately in a correctly labelled dry container, accompanied by a completed Request Form and handed to a Medical Scientist, NCHD or |
| Transport: | laboratory immediately in a correctly labelled dry container, accompanied by a completed Request Form and handed to a Medical Scientist, NCHD or Consultant Histopathologist in the Histopathology laboratory. The form must |
| Transport: | laboratory immediately in a correctly labelled dry container, accompanied by a completed Request Form and handed to a Medical Scientist, NCHD or Consultant Histopathologist in the Histopathology laboratory. The form must have a red Frozen sticker attached. Specimens from external hospitals must |
| Transport: Turnaround: | laboratory immediately in a correctly labelled dry container, accompanied by a completed Request Form and handed to a Medical Scientist, NCHD or Consultant Histopathologist in the Histopathology laboratory. The form must |

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| Laboratory: | Clinical Biochemistry |
|-------------|---|
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Turnaround: | 4 Days |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as |
| | appropriate |

Full Blood Count including automated WBC DifferentialBlood Films for Manual White Cell Differentials, Slide Platelets and Red Cell Morphology (peripheral blood smear)

| Laboratory: | Haematology |
|-------------|---|
| Specimen: | Blood 3mL purple Vacuette [®] (EDTA) |
| | Paediatric (1mL purple (EDTA) or 1.3 mL red) |
| | Note: 6ml purple EDTA Vacuette or any other sample type is unsuitable for |
| | FBC. |
| | Blood Films are made in the laboratory as required. |
| Comment: | Full Blood Counts: Impedence /Fluorescence Flow Cytometry Technology. |
| | Test such that Manufact to Evidence during a ventile succession because and for |

Test available Monday to Friday, during routine working hours and for emergency reasons at all other times. FBC performed in the investigation of anaemias, infections, leukeamias, platelet disorders and myeloproliferative disorders and also for the monitoring of therapies, e.g. nutritional, chemotherapy.

> **Manual differentials, slide platelets and red cell morphology** available when deemed necessary or when the laboratory is contacted by clinician. **Note:** NRBCs occur in peripheral blood in neonates and premature babies in low numbers as a normal finding. In healthy adults and older children, NRBCs are only found in bone marrow where they mature. Their appearance in peripheral blood points to extramedullary erythropoiesis or marrow stress with disruption of the blood-bone marrow barrier.¹ Results of NRBC count must be interpreted in conjunction with the full clinical picture. The requesting clinician is responsible for evaluating the reported NRBC count and evaluating the presence of any NRBCs reported in the FBC in the light of the patients age and clinical details. (¹Sysmex Xtra Online | March 2012 | The clinical relevance of measuring NRBC in the XN-CBC)

Storage: If delays are unavoidable, samples can be preserved by refrigeration at 2-8°C in a designated specimen fridge.

| Stability: | | Ambient Temperature | Refrigerated |
|------------|------|---------------------|--------------|
| | WBC | 36 hrs | 56 hrs |
| | RBC | 48 hrs | 72 hrs |
| | HB | 72 hrs | 72 hrs |
| | MCV | 8 hrs | 24 hrs |
| | PLTS | 48 hrs | 48 hrs |

Transport: Transport specimen to the laboratory at ambient temperature.

Turnaround: Full Blood Counts:

Emergency specimens < 2 hours. Urgent specimens, i.e. received from wards with urgent label: 4 hours. Routine in-hospital specimens: 8 hours GP specimens: 2 days

Manual differentials, slide platelets and red cell morphology

Clinically significant: 4 hours Routine specimens 48 hours

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Ref. Range: Age and sex Related Reference Ranges

| Age and sex Related Referen | | | |
|---|--|------------------------------|--|
| Analyte & units | Age | Sex | Range |
| Haemoglobin g/dl | 0 minutes – 24 hours | Male | 14.9-23.7 |
| Haemoglobin | 1 day – 14 days | Male | 13.4 - 19.8 |
| Haemoglobin | 14 days – 2 months | Male | 9.4-13.0 |
| Haemoglobin | 2 months – 6 months | Male | 10.0-13.0 |
| Haemoglobin | 6 months – 12 months | Male | 10.1 - 13.0 |
| Haemoglobin | 12 months – 6 years | Male | 11.0 - 13.8 |
| Haemoglobin | 6 years – 12 years | Male | 11.1 - 14.7 |
| Haemoglobin | 12 years – 18 years | Male | 12.1 - 16.6 |
| Haemoglobin | >18 years | Male | 13.0 - 17.0 |
| Haemoglobin | 0 minutes – 24 hours | Female | 14.9 - 23.7 |
| | | | |
| Haemoglobin | 1 day – 14 days | Female | 13.4 - 19.8 |
| Haemoglobin | 14 days – 2 months | Female | 9.4 - 13.0 |
| Haemoglobin | 2 months – 6 months | Female | 10.0 - 13.0 |
| Haemoglobin | 6 months – 12 months | Female | 10.1 - 13.0 |
| Haemoglobin | 12 months – 6 years | Female | 11.0 - 13.8 |
| Haemoglobin | 6 years – 12 years | Female | 11.1 - 14.7 |
| Haemoglobin | 12 years – 18 years | Female | 12.1 - 15.1 |
| Haemoglobin | >18 years | Female | 11.7 - 15.9 |
| | | | |
| Red cell count x 10 ¹² /l | 0 minutes – 24 hours | Male | 3.7-6.5 |
| Red cell count | 1 day - 14 days | Male | 3.9-5.9 |
| Red cell count | 14 days – 2 months | Male | 3.1-4.3 |
| Red cell count | 2 months – 6 months | Male | 3.8 - 4.9 |
| Red cell count | 6 months – 12 months | Male | 3.9-5.1 |
| Red cell count | 12 months – 6 years | Male | 3.9 - 5.0 |
| Red cell count | 6 years – 12 years | Male | 3.9 - 5.2 |
| Red cell count | 12 years – 18 years | Male | 4.2 - 5.6 |
| Red cell count | >18 years | Male | 4.2 - 5.6 |
| Red cell count | 0 minutes – 24 hours | Female | 3.7-6.5 |
| Red cell count | 1 day – 14 days | Female | 3.9-5.9 |
| Red cell count | | Female | 3.1-4.3 |
| | 14 days – 2 months | | |
| Red cell count | 2 months – 6 months | Female | 3.8 - 4.9 |
| Red cell count | 6 months – 12 months | Female | 3.9 - 5.1 |
| Red cell count | 12 months – 6 years | Female | 3.9 - 5.0 |
| Red cell count | 6 years – 12 years | Female | 3.9 - 5.2 |
| Red cell count | 12 years – 18 years | Female | 4.1 - 5.1 |
| Red cell count | >18 years | Female | 3.9 - 5.3 |
| | | | |
| White blood cell count x 10 ⁹ /l | 0 minutes – 24 hours | All | 10.0 - 26.0 |
| WBCC | 1 day – 14 days | All | 6.0 - 21.0 |
| WBCC | 14 days – 2 months | All | 5.0 - 15.0 |
| WBCC | 2 months – 6 months | All | 6.0 - 17.0 |
| WBCC | 6 months – 12 months | All | 6.0 - 16.0 |
| WBCC | 12 months – 6 years | All | 6.0 - 17.0 |
| WBCC | 6 years – 12 years | All | 4.5 - 14.5 |
| WBCC | 12 years – 18 years | All | 4.5 - 13.0 |
| WBCC | >18 years | All | 4.4 - 11.3 |
| | | 1 | |
| Haematocrit I/I | 0 minutes – 24 hours | Male | 0.47 - 0.75 |
| Haematocrit | | Male | 0.41 - 0.65 |
| | 1 nav = 14 navs | Huic | |
| | 1 day – 14 days | Malo | 1 11 78 - 11 77 |
| Haematocrit | 14 days – 2 months | Male | 0.28 - 0.42 |
| Haematocrit Haematocrit | 14 days – 2 months 2 months – 6 months | Male | 0.30 - 0.38 |
| Haematocrit Haematocrit Haematocrit | 14 days – 2 months 2 months – 6 months 6 months – 12 months | Male Male | 0.30 - 0.38 0.30 - 0.38 |
| Haematocrit Haematocrit Haematocrit Haematocrit | 14 days – 2 months 2 months – 6 months 6 months – 12 months 12 months – 6 years | Male Male Male | 0.30 - 0.38 0.30 - 0.38 0.32 - 0.40 |
| Haematocrit Haematocrit Haematocrit Haematocrit Haematocrit | 14 days – 2 months 2 months – 6 months 6 months – 12 months 12 months – 6 years 6 years – 12 years | Male Male Male Male | 0.30 - 0.38 0.30 - 0.38 0.32 - 0.40 0.32 - 0.43 |
| Haematocrit Haematocrit Haematocrit Haematocrit | 14 days – 2 months 2 months – 6 months 6 months – 12 months 12 months – 6 years | Male Male Male | 0.30 - 0.38 0.30 - 0.38 0.32 - 0.40 |

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| Haematocrit | 0 minutes – 24 hours | Female | 0.47 - 0.75 |
|----------------------------|---------------------------------------|--------|-------------|
| Haematocrit | 1 day – 14 days | Female | 0.41 - 0.65 |
| Haematocrit | 14 days – 2 months | Female | 0.28 - 0.42 |
| Haematocrit | 2 months – 6 months | Female | 0.30 - 0.38 |
| Haematocrit | 6 months – 12 months | Female | 0.30 - 0.38 |
| Haematocrit | 12 months – 6 years | Female | 0.32 - 0.40 |
| Haematocrit | 6 years – 12 years | Female | 0.32 - 0.43 |
| Haematocrit | 12 years – 18 years | Female | 0.35 - 0.44 |
| Haematocrit | >18 years | Female | 0.35 - 0.46 |
| | , , , , , , , , , , , , , , , , , , , | | |
| Mean Cell Haemoglobin pg | 0 minutes – 24 hours | Male | 30.0 - 37.2 |
| MCH | 1 day – 14 days | Male | 30.0 - 37.2 |
| MCH | 14 days – 2 months | Male | 27.0 - 36.0 |
| МСН | 2 months – 6 months | Male | 24.0 - 32.2 |
| MCH | 6 months – 12 months | Male | 24.0 - 29.6 |
| МСН | 12 months – 6 years | Male | 25.6 - 30.7 |
| МСН | 6 years – 12 years | Male | 26.3 - 30.9 |
| MCH | 12 years – 18 years | Male | 26.9 - 31.9 |
| MCH | >18 years | Male | 26.0 - 34.0 |
| MCH | 0 minutes – 24 hours | Female | 30.0 - 37.2 |
| МСН | 1 day - 14 days | Female | 30.0 - 37.2 |
| МСН | 14 days – 2 months | Female | 27.0 - 36.0 |
| МСН | 2 months – 6 months | Female | 24.0 - 32.2 |
| МСН | 6 months – 12 months | Female | 24.0 - 29.6 |
| МСН | 12 months – 6 years | Female | 25.6 - 30.7 |
| МСН | 6 years – 12 years | Female | 26.3 - 30.9 |
| МСН | 12 years – 18 years | Female | 26.7 - 32.5 |
| МСН | >18 years | Female | 26.0 - 34.0 |
| | | | |
| Mean Cell Haemoglobin | | | |
| Concentration g/dL MCHC | 0 minutes – 24 hours | Male | 28.1 - 34.7 |
| MCHC | 1 day – 14 days | Male | 28.1 - 34.7 |
| MCHC | 14 days – 2 months | Male | 28.1 - 35.5 |
| MCHC | 2 months – 6 months | Male | 28.8 - 37.3 |
| MCHC | 6 months – 12 months | Male | 32.1 - 37.4 |
| МСНС | 12 months – 6 years | Male | 32.9 - 35.6 |
| MCHC | 6 years – 12 years | Male | 32.7 - 35.7 |
| МСНС | 12 years – 18 years | Male | 33.5 - 35.2 |
| МСНС | >18 years | Male | 31.0 - 37.0 |
| MCHC | 0 minutes – 24 hours | Female | 28.1 - 34.7 |
| MCHC | 1 day – 14 days | Female | 28.1 - 34.7 |
| MCHC | 14 days – 2 months | Female | 28.1 - 35.5 |
| МСНС | 2 months – 6 months | Female | 28.8 - 37.3 |
| МСНС | 6 months – 12 months | Female | 32.1 - 37.4 |
| MCHC | 12 months – 6 years | Female | 32.9 - 35.6 |
| МСНС | 6 years – 12 years | Female | 32.7 - 35.7 |
| MCHC | 12 years – 18 years | Female | 33.0 - 35.5 |
| MCHC | >18 years | Female | 31.0 - 37.0 |
| | | | |
| Mean Cell Volume fl | 0 minutes – 24 hours | Male | 100-125 |
| MCV | 1 day – 14 days | Male | 88 - 110 |
| MCV | 14 days – 2 months | Male | 84 - 98 |
| MCV | 2 months – 6 months | Male | 73 - 84 |
| MCV | 6 months – 12 months | Male | 70 – 82 |
| MCV | 12 months – 6 years | Male | 72 – 87 |
| MCV | 6 years – 12 years | Male | 76 – 90 |
| MCV | 12 years – 18 years | Male | 77 – 92 |
| | | | |
| MCV | >18 years | Male | 80 - 96 |

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| MCV 0 minutes - 24 hours Female 100-125 MCV 1 day - 14 days Female 88 - 11 MCV 14 days - 2 months Female 88 - 11 MCV 2 months - 6 months Female 84 - 98 MCV 2 months - 6 months Female 73 - 84 MCV 6 months - 12 months Female 70 - 82 MCV 12 months - 6 years Female 70 - 82 MCV 12 years - 12 years Female 76 - 90 MCV 12 years - 18 years Female 77 - 94 MCV 12 years - 18 years Female 80 - 96 Basophil count x 10 ⁹ /I 0 minutes - 24 hours All 0.0 - 0. Basophil count 1 day - 14 days All 0.02 - 0 Basophil count 12 months - 6 months All 0.02 - 0 Basophil count 12 months - 6 years All 0.02 - 0 Basophil count 12 months - 6 years All 0.02 - 0 Basophil count 12 years - 18 years All | 0 1 1 0.13 0.20 0.13 0.12 0.12 0.12 |
|--|--|
| MCV14 days - 2 monthsFemale $84 - 98$ MCV2 months - 6 monthsFemale $73 - 84$ MCV6 months - 12 monthsFemale $70 - 82$ MCV12 months - 6 yearsFemale $72 - 87$ MCV6 years - 12 yearsFemale $76 - 90$ MCV12 years - 18 yearsFemale $77 - 94$ MCV>18 yearsFemale $80 - 96$ MCV>18 yearsFemale $80 - 96$ Basophil count x $10^9/I$ 0 minutes - 24 hoursAll $0.0 - 0.$ Basophil count1 day - 14 daysAll $0.02 - 0.$ Basophil count2 months - 6 monthsAll $0.02 - 0.$ Basophil count6 months - 12 monthsAll $0.02 - 0.$ Basophil count6 years - 12 yearsAll $0.02 - 0.$ Basophil count12 months - 6 yearsAll $0.02 - 0.$ Basophil count6 years - 12 wearsAll $0.02 - 0.$ Basophil count12 months - 6 yearsAll $0.02 - 0.$ Basophil count12 wears - 18 yearsAll $0.02 - 0.$ Basophil count12 years - 18 yearsAll $0.02 - 0.$ Basophil count1 day - 14 daysAll $0.02 - 0.$ Basophil count12 years - 18 yearsAll $0.02 - 0.$ Basophil count12 wearsAll $0.02 - 0.$ Basophil count12 wearsAll $0.02 - 0.$ Basophil count1 day - 14 daysAll $0.02 - 0.$ Basophil count1 day - 14 days | 1 1 0.13 0.20 0.13 0.12 0.12 0.12 |
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| Basophil count12 months - 6 yearsAll $0.02 - 0$ Basophil count6 years - 12 yearsAll $0.02 - 0$ Basophil count12 years - 18 yearsAll $0.02 - 0$ Basophil count>18 yearsAll $0.02 - 0$ Basophil count>18 yearsAll $0.0 - 0$ Eosinophil count x 10 ⁹ /I0 minutes - 24 hoursAll $0.0 - 0$ Eosinophil count1 day - 14 daysAll $0.0 - 0$ Eosinophil count2 months - 6 monthsAll $0.05 - 0$ Eosinophil count12 months - 6 yearsAll $0.05 - 0$ |).12).12).12 |
| Basophil count6 years - 12 yearsAll $0.02 - 0$ Basophil count12 years - 18 yearsAll $0.02 - 0$ Basophil count>18 yearsAll $0.0 - 0$ Eosinophil count x 10 ⁹ /I0 minutes - 24 hoursAll $0.0 - 0$ Eosinophil count1 day - 14 daysAll $0.0 - 0$ Eosinophil count14 days - 2 monthsAll $0.05 - 0$ Eosinophil count6 months - 6 monthsAll $0.1 - 1$ Eosinophil count12 months - 6 yearsAll $0.05 - 0$ |).12).12 |
| Basophil count12 years – 18 yearsAll $0.02 - 0$ Basophil count>18 yearsAll $0.0 - 0$ Eosinophil count x 10 ⁹ /I0 minutes – 24 hoursAll $0.0 - 0$ Eosinophil count1 day – 14 daysAll $0.0 - 0$ Eosinophil count14 days – 2 monthsAll $0.05 - 0$ Eosinophil count2 months – 6 monthsAll $0.1 - 1$ Eosinophil count12 months – 6 yearsAll $0.05 - 0$ |).12 |
| Basophil count>18 yearsAll $0.0 - 0.$ Eosinophil count x 10°/I0 minutes - 24 hoursAll $0.0 - 0.$ Eosinophil count1 day - 14 daysAll $0.0 - 0.$ Eosinophil count14 days - 2 monthsAll $0.05 - 0.$ Eosinophil count2 months - 6 monthsAll $0.1 - 1.$ Eosinophil count6 months - 12 monthsAll $0.05 - 0.$ Eosinophil count12 months - 6 yearsAll $0.05 - 1.$ | |
| Eosinophil count x $10^9/I$ 0 minutes - 24 hoursAll $0.0 - 0.$ Eosinophil count1 day - 14 daysAll $0.0 - 0.$ Eosinophil count14 days - 2 monthsAll $0.05 - 0.$ Eosinophil count2 months - 6 monthsAll $0.1 - 1.$ Eosinophil count6 months - 12 monthsAll $0.05 - 0.$ Eosinophil count12 months - 6 yearsAll $0.05 - 1.000$ | - |
| Eosinophil count1 day - 14 daysAll0.0 - 0.Eosinophil count14 days - 2 monthsAll0.05 - 0.Eosinophil count2 months - 6 monthsAll0.1 - 1.Eosinophil count6 months - 12 monthsAll0.05 - 0.Eosinophil count12 months - 6 yearsAll0.05 - 0. | |
| Eosinophil count 1 day - 14 days All 0.0 - 0. Eosinophil count 14 days - 2 months All 0.05 - 0. Eosinophil count 2 months - 6 months All 0.1 - 1. Eosinophil count 6 months - 12 months All 0.05 - 0. Eosinophil count 12 months - 6 years All 0.05 - 0. | 85 |
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| Eosinophil count6 months – 12 monthsAll0.05 – 0Eosinophil count12 months – 6 yearsAll0.05 – 1 | |
| Eosinophil count 12 months – 6 years All 0.05 – 1 | |
| | |
| EOSINODNII COUNT TO VEALS - 12 VEALS TAIL TU.U.S - 1 | |
| | |
| Eosinophil count 12 years - 18 years All 0.05 - 0 | |
| Eosinophil count >18 years All 0.04 - 0 |).4 |
| Lymphocyte count x $10^{9}/l$ 0 minutes – 24 hours All 2.0 – 7. | 3 |
| Lymphocyte count 1 day – 14 days All 2.8 – 9. | |
| Lymphocyte count 14 days – 2 months All 3.3 – 10 | |
| Lymphocyte count 2 months – 6 months All 3.3 – 11 | |
| | |
| | |
| | |
| Lymphocyte count 6 years – 12 years All 1.5 – 5. | |
| Lymphocyte count 12 years – 18 years All 1.5 – 4. | |
| Lymphocyte count >18 years All 0.9 – 3. | 2 |
| Monocyte count x $10^9/l$ 0 minutes – 24 hours All 0.0 – 1. | 9 |
| Monocyte count1 day - 14 daysAll0.00.1 - 1. | |
| Monocyte count1 day = 14 daysAll $0.1 = 1.$ Monocyte count14 days - 2 monthsAll $0.4 - 1.$ | |
| Monocyte count14 days - 2 monthsAll $0.4 - 1.$ Monocyte count2 months - 6 monthsAll $0.2 - 1.$ | |
| | |
| Monocyte count 6 months – 12 months All 0.2 – 0. | |
| Monocyte count 12 months - 6 years All 0.15 - 1 | |
| Monocyte count 6 years - 12 years All 0.15 - 1 | |
| Monocyte count 12 years – 18 years All 0.15 – 1 | |
| Monocyte count >18 years All 0.15 - 1 | 1.3 |
| Neutrophil count x 10^9 /l 0 minutes – 24 hours All 2.7 – 14 | 1 4 |
| Neutrophil count1 day - 14 daysAll $2.7 - 12$ Neutrophil count1 day - 14 daysAll $1.5 - 5.$ | |
| | |
| | |
| Neutrophil count2 months – 6 monthsAll $1.0 - 6.$ Neutrophil count6 months 12 monthsAll | |
| Neutrophil count 6 months – 12 months All 1.0 – 8. | |
| Neutrophil count 12 months – 6 years All 1.5 – 8. | |
| Neutrophil count6 years – 12 yearsAll1.5 – 8. | <u>0</u> |
| Neutrophil count12 years - 18 yearsAll1.5 - 6.Neutrophil count>18 yearsAll1.4 - 6. | |
| Neutrophil count >18 years All 1.4 – 6. | |

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| Platelet count x 10 ⁹ /l | 0 minutes – 24 hours | All | 150 - 450 |
|---|----------------------|----------|-----------|
| Platelet count | 1 day - 14 days | All | 170 - 500 |
| Platelet count | 14 days – 2 months | All | 210 - 650 |
| Platelet count | 2 months – 6 months | All | 210 - 560 |
| Platelet count | 6 months – 12 months | All | 200 - 550 |
| Platelet count | 12 months – 6 years | All | 210 - 490 |
| Platelet count | 6 years – 12 years | All | 170 - 450 |
| Platelet count | 12 years – 18 years | All | 180 - 430 |
| Platelet count | >18 years | All | 140 - 440 |
| | | | |
| Reticulocyte count x 10 ⁹ /l | 0 minutes – 24 hours | All | 110 - 450 |
| Reticulocyte count | 1 day – 7 days | All | 10 - 80 |
| Reticulocyte count | 7 days – 1 month | All | 10 - 65 |
| Reticulocyte count | 1 month – 2 months | All | 35 - 200 |
| Reticulocyte count | 2 months – 5 months | All | 15 - 110 |
| Reticulocyte count | 5 months – 12 months | All | 30 - 130 |
| Reticulocyte count | >12 months | All | 50 - 100 |
| | | | |
| Erythrocyte Sedimentation | All | Male | 0 - 10 |
| Rate mm/hour | | <u> </u> | |
| Erythrocyte Sedimentation Rate | All | Female | 0 – 20 |

Full Blood Counts in Pregnancy

Anaemia is defined by Hb <11g/dl in first trimester, <105g/dl in second and third trimesters, and <10g/dl in postpartum period*.

*BSCH UK Guidelines for the management of iron deficiency in Pregnancy, 2019.

| Period of Gestation | First trimester | Second Trimester | Third Trimester |
|--|--------------------|---------------------|--------------------|
| RBC x 10 ⁻¹² /l | 3.52-4.52 | 3.2-4.41 | 3.1-4.44 |
| HB g/dl | 11-14.3 | 10-13.7 | 9.8-13.7 |
| HCT L/L | 0.31-0.41 | 0.30-0.38 | 0.28-0.39 |
| MCV fl | 81-96 | 82-97 | 81-99 |
| WBC x 10 ⁻⁹ /l | 5.7-13.6 | 6.2-14.8 | 5.9-16.9 |
| Neutrophil count x 10 ⁻⁹ /l | 3.6-10.1 | 3.8-12.3 | 3.9-13.1 |
| Lymphocyte count x 10 ⁻⁹ /l | 1.1-3.5 | 0.9-3.9 | 1-3.6 |
| Monocyte count x 10 ⁻⁹ /l | 0-1 | 0.1-1.1 | 0.1-1.1 |
| Eosinophil count x 10 ⁻⁹ /l | 0-0.6 | 0-0.6 | 0-0.6 |
| Basophil count x 10 ⁻⁹ /l | 0-0.1 | 0-0.1 | 0-0.1 |
| Platelet count x 10 ⁻⁹ /l | 174-391 | 171-409 | 155-429 |

Table above: 95% ranges for haematological variables during pregnancy Taken from 'Blood Cells. A practical Guide.' Barbara J. Bain, 5th Edition, 2015.

Fungal Microscopy and Culture

See Mycology

| GATA Mutation | al analysis |
|----------------------|--|
| Laboratory: | Referred from Haematology to North Bristol NHS Trust, |
| | Bristol Genetics Lab, Pathology Sciences, Southmead Hospital, Westbury-On- |
| | Trym, Bristol, BS10 5NB |
| Specimen: | 3 mL EDTA |
| Comment: | By arrangement only with laboratory |
| Turnaround: | 64 working days |
| Report: | Sent to referring clinician and copy filed in laboratory |

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| | Author. Pil Paul Cantwen |
|----------------------------|--|
| Gastrin | |
| Laboratory: | Referred from Biochemistry to SAS Centre, Charing Cross Hospital |
| Specimen: | 10 ml EDTA blood (overnight fast) |
| | 3 ml non-hemolysed plasma for full screen. |
| Turnaround: | 10 weeks |
| Ref. range: | See report form |
| G6PD Assay | |
| Laboratory: | Referred from Haematology t to Viapath Analytics, The Red Cell Centre, Reference Haematology, King's College Hospital (Kingspath Hospital) |
| Specimen: | Blood 3mL purple Vacuette [®] (EDTA) |
| Comment: | Used in the investigation of Hereditary Haemolytic Anaemias. It is recommended that assays not be performed after severe haemolytic crisis, since G6PD levels may be falsely elevated. Test available Monday to Friday, during routine working hours. Unsuitable for analysis if Reticulocyte count is >150 x 10 ⁹ /L, may be referre |
| Turnaround: | 60 working days |
| Ref. Range: | 4.6 – 13.5 U/g Hb. |
| Ref. Range. | Note: Values for new-borns may range somewhat higher, see final report |
| | Note. Values for new-borns may range somewhat night, see final report |
| G6PD Screen | |
| Laboratory: | Haematology |
| Specimen: | Blood 3mL purple Vacuette [®] (EDTA) |
| Comment: | Used in the investigation of Hereditary Haemolytic Anaemias. Samples which have been determined deficient or intermediate by this qualitative method are referred. It is recommended that assays not be performed after severe haemolytic crisis, since G6PD levels may be falsely elevated. Test available Monday to Friday, during routine working hours. Unsuitable for analysis if Reticulocyte count is >150 x 10^9 /L |
| | |
| | Sulfasalazine or its metabolites may interfere with this test, so results are no valid for individuals taking these medications |
| Turnaround: | 1 week |
| Ref. Range: | Normal/Decreased/Inconclusive |
| | nyltransferase (γ-GT) |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Turnaround: Ref. Range: | A/E or urgent sample: - 1 hour 30 mins CUH wards, CUMH, SI, SF, SMOH, MGH: - 3 hours 130egain130. GP or OPD- Results posted within 4 days. Up-to-date reference intervals will be applied to all Biochemistry reports as |
| | appropriate |
| Ganglioside An | |
| Laboratory: | Sample referred from Autoimmune Serology to Eurofins-Biomnis Laboratories. |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) |
| Turnaround: | Approx. 3 Weeks |
| Ref. Range: | See report form, or visit internet site <u>https://www</u> .eurofins.ie/biomnis/ for up to date referral test information. |
| Gastric Parieta | l Cell Ab |
| | |
| Laboratory: | Autoimmune Serology |
| Laboratory: Specimen: | Autoimmune Serology Blood, 4 mL red top Vacuette (or similar container for clotted blood) |
| • | |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) |

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| | pecimen referred from Histopathology to Dr. Cathal O'Brien, Cancer Iolecular Diagnostics, St James' Hospital Dublin |
|-----------------------------|--|
| • | listopathology Tissue block |
| | 0 working days |
| • | lar Basement Membrane Antibodies) |
| Laboratory: | Autoimmune Serology |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) |
| Comment: | Quantitative Immunoassay using Phadia Unicap 250 analyser. Restricted to |
| | CUH patients. |
| Turnaround: | 72 Hours |
| Ref. Range: | 0 - 10 AU/mL |
| GBMQ (GBM Q | |
| Laboratory: | Autoimmune Serology |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) |
| Comment: | Qualitative Quick Card Test (5 Minutes) |
| Turnaround: | On Request. |
| Ref. Range: Genital Swab | Not applicable |
| | dia trachematic DCD and N. ganarrhaga DCD |
| Laboratory: | dia trachomatis PCR and N. gonorrhoea PCR Microbiology (Main laboratory) |
| Specimen: | Specimens for culture and sensitivity testing should be taken in the following |
| Specimen. | situations: |
| | • The patient is clearly symptomatic of gonoccal infection. |
| | • The patient has tested positive for <i>N. gonorrhoea</i> on the urine |
| | cobas assay but has not yet commenced treatment. |
| | There is evidence of treatment failure. |
| | The patient is a known contact, and immediate epidemiologica |
| | treatment is to be given. |
| | Because genital specimens are often taken from sites harbouring large numbers of commensal (normal) flora, attention to specimen selection and collection methods is critical. |
| | Specimens should be collected using a sterile swab and transported ASAP in charcoal containing transport media. |
| | The viability of <i>N. gonorrhoeae</i> is lost over time. |
| | If processing is delayed, storage at ambient temperature is preferred. |
| Comment: | Test performed routinely Monday to Friday 9-5pm or by urgent request. |
| Turnaround: | Prelim: 24 hours; Final: 72 hours. |
| Report: | Culture report on any clinically significant isolate with the appropriate sensitivities. |
| Genitourinary | - TFE3/TFEB 131egain + Renal tumour Cytogenetics |
| Laboratory: | Specimen referred from Histopathology to Dr. Jonathan Shanks, The Christie Foundation Manchester, UK |
| Specimen: | Histopathology Tissue block |
| Turnaround: | 21 days |
| | Genticin |
| | |

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| Glucocorticoid | Remedical Aldosteronism (GRA) |
|---|--|
| Laboratory: | Referred from Molecular Genetics Lab in Biochemistry to Addenbrookes NHS (|
| | via NCMG) |
| Specimen: | 3-5ml EDTA blood |
| Comment: | Use NCMG request form, available at <u>www.genetics.ie</u> |
| | Please note: invoices will be issued to the referring clinician for tests not |
| | performed in NCMG. |
| Turnaround: | 2 weeks |
| Report: | Sent to referring clinician and copy filed in pathology |
| Glucose | |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL Sodium fluoride EDTA |
| Comment: | Grey-capped specimen tube. Fluid Glucose should also be taken into a Grey- capped specimen. |
| Turnaround: | A/E or urgent sample: - 1 hour 30 mins CUH wards, CUMH, SI, SF, SMOH, |
| | MGH: - 3 hours 132egain132. |
| | GP or OPD- Results posted within 4 days. |
| Ref. Range: | WHO Guidelines. See report form |
| Glucose (Urina | |
| Laboratory: | Clinical Biochemistry or ward / GP surgery |
| Specimen: | Fresh spot urine sample |
| Comment: | Measured using dipstick. Aged sample invalidates result. |
| Turnaround: | 1 Day |
| Ref. Range: | Should be NEGATIVE |
| Glutamic Acid | Decarboxylase Antibodies |
| Laboratory: | Sample for GAD and IA2 are referred from Autoimmune Serology to |
| | Immunology lab, Exeter. |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) |
| Turnaround: | Approx. 3 Weeks |
| Ref. Range: | See report form. |
| Group B Strept | |
| | |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Laboratory: Specimen: | Microbiology (Infectious Diseases Serology) 1mL EDTA blood, CSF (0.5mL) |
| Laboratory: Specimen: Comment: | Microbiology (Infectious Diseases Serology) 1mL EDTA blood, CSF (0.5mL) Performed by Irish Meningitis & Sepsis Reference Laboratory (IMSRL), Dublin |
| Laboratory: Specimen: | Microbiology (Infectious Diseases Serology) 1mL EDTA blood, CSF (0.5mL) Performed by Irish Meningitis & Sepsis Reference Laboratory (IMSRL), Dublin 10 working days. Samples received by IMSRL before 11am, verbal result |
| Laboratory: Specimen: Comment: Turnaround: | Microbiology (Infectious Diseases Serology) 1mL EDTA blood, CSF (0.5mL) Performed by Irish Meningitis & Sepsis Reference Laboratory (IMSRL), Dublin 10 working days. Samples received by IMSRL before 11am, verbal result between 4pm and 5pm the same day (positive only). |
| Laboratory: Specimen: Comment: Turnaround: Report: | Microbiology (Infectious Diseases Serology) 1mL EDTA blood, CSF (0.5mL) Performed by Irish Meningitis & Sepsis Reference Laboratory (IMSRL), Dublin 10 working days. Samples received by IMSRL before 11am, verbal result between 4pm and 5pm the same day (positive only). Detected or not detected |
| Laboratory: Specimen: Comment: Turnaround: Report: Growth hormc | Microbiology (Infectious Diseases Serology) 1mL EDTA blood, CSF (0.5mL) Performed by Irish Meningitis & Sepsis Reference Laboratory (IMSRL), Dublin 10 working days. Samples received by IMSRL before 11am, verbal result between 4pm and 5pm the same day (positive only). Detected or not detected Detected One (GH) |
| Laboratory: Specimen: Comment: Turnaround: Report: Growth hormo Laboratory: | Microbiology (Infectious Diseases Serology) 1mL EDTA blood, CSF (0.5mL) Performed by Irish Meningitis & Sepsis Reference Laboratory (IMSRL), Dublin 10 working days. Samples received by IMSRL before 11am, verbal result between 4pm and 5pm the same day (positive only). Detected or not detected Detected or not detected Clinical Biochemistry |
| Laboratory: Specimen: Comment: Turnaround: Report: Growth hormo Laboratory: Specimen: | Microbiology (Infectious Diseases Serology) 1mL EDTA blood, CSF (0.5mL) Performed by Irish Meningitis & Sepsis Reference Laboratory (IMSRL), Dublin 10 working days. Samples received by IMSRL before 11am, verbal result between 4pm and 5pm the same day (positive only). Detected or not detected Detected or not detected Detected Biochemistry 4.0 mL blood in plain tube (clotted sample) |
| Laboratory: Specimen: Comment: Turnaround: Report: Growth hormo Laboratory: Specimen: Turnaround: | Microbiology (Infectious Diseases Serology) 1mL EDTA blood, CSF (0.5mL) Performed by Irish Meningitis & Sepsis Reference Laboratory (IMSRL), Dublin 10 working days. Samples received by IMSRL before 11am, verbal result between 4pm and 5pm the same day (positive only). Detected or not detected Detected or not detected Clinical Biochemistry 4.0 mL blood in plain tube (clotted sample) 2 Weeks |
| Laboratory: Specimen: Comment: Turnaround: Report: Growth hormo Laboratory: Specimen: | Microbiology (Infectious Diseases Serology) 1mL EDTA blood, CSF (0.5mL) Performed by Irish Meningitis & Sepsis Reference Laboratory (IMSRL), Dublin 10 working days. Samples received by IMSRL before 11am, verbal result between 4pm and 5pm the same day (positive only). Detected or not detected Detected or not detected Detected in plain tube (clotted sample) 2 Weeks Haemolysed samples should be interpreted with care |
| Laboratory: Specimen: Comment: Turnaround: Report: Growth hormo Laboratory: Specimen: Turnaround: | Microbiology (Infectious Diseases Serology) 1mL EDTA blood, CSF (0.5mL) Performed by Irish Meningitis & Sepsis Reference Laboratory (IMSRL), Dublin 10 working days. Samples received by IMSRL before 11am, verbal result between 4pm and 5pm the same day (positive only). Detected or not detected Detected or not detected Clinical Biochemistry 4.0 mL blood in plain tube (clotted sample) 2 Weeks Haemolysed samples should be interpreted with care Samples should be transported to the laboratory as soon as possible and |
| Laboratory: Specimen: Comment: Turnaround: Report: Growth hormo Laboratory: Specimen: Turnaround: Comment: | Microbiology (Infectious Diseases Serology) 1mL EDTA blood, CSF (0.5mL) Performed by Irish Meningitis & Sepsis Reference Laboratory (IMSRL), Dublin 10 working days. Samples received by IMSRL before 11am, verbal result between 4pm and 5pm the same day (positive only). Detected or not detected Detected or not detected Detected or not detected Clinical Biochemistry 4.0 mL blood in plain tube (clotted sample) 2 Weeks Haemolysed samples should be interpreted with care Samples should be transported to the laboratory as soon as possible and must be frozen within 24hours |
| Laboratory: Specimen: Comment: Turnaround: Report: Growth hormo Laboratory: Specimen: Turnaround: | Microbiology (Infectious Diseases Serology) 1mL EDTA blood, CSF (0.5mL) Performed by Irish Meningitis & Sepsis Reference Laboratory (IMSRL), Dublin 10 working days. Samples received by IMSRL before 11am, verbal result between 4pm and 5pm the same day (positive only). Detected or not detected One (GH) Clinical Biochemistry 4.0 mL blood in plain tube (clotted sample) 2 Weeks Haemolysed samples should be interpreted with care Samples should be transported to the laboratory as soon as possible and must be frozen within 24hours It is not possible to quote a reference range for random Growth Hormone due |
| Laboratory: Specimen: Comment: Turnaround: Report: Growth hormo Laboratory: Specimen: Turnaround: Comment: | Microbiology (Infectious Diseases Serology) 1mL EDTA blood, CSF (0.5mL) Performed by Irish Meningitis & Sepsis Reference Laboratory (IMSRL), Dublin 10 working days. Samples received by IMSRL before 11am, verbal result between 4pm and 5pm the same day (positive only). Detected or not detected Detected or not detected Detected or not detected Clinical Biochemistry 4.0 mL blood in plain tube (clotted sample) 2 Weeks Haemolysed samples should be interpreted with care Samples should be transported to the laboratory as soon as possible and must be frozen within 24hours |

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| Gut Hormone p | profile |
|----------------|--|
| Laboratory: | Sample referred from Clinical Biochemistry to SAS Laboratory, Charing Cross Hospital |
| Specimen: | Blood, 10mL fasting in EDTA bottle sent to the laboratory on ice. |
| Comment: | Consultant request only |
| Turnaround: | 3 weeks |
| Ref. Range: | See report form. |
| Guanidinoaceta | ate/creatine |
| Laboratory: | Sample referred from Clinical Biochemistry to Biochemical Genetics Unit, Addenbrookes |
| Specimen: | MSU + 0.5ml Li-Hep Plasma |
| Turnaround: | 5 weeks |
| Ref. Range: | See report form |
| Haemochromat | tosis |
| Laboratory: | Performed in the Molecular Genetics lab in Biochemistry |
| Specimen: | 3.0 mL EDTA blood |
| | Please see investigation guidelines and specific request form on CUH website, |
| | www.cuh.hse.ie |
| Turnaround: | 4-6 Weeks |
| Report: | Sent to referring clinician. Restricted access to genetic reports on laboratory |
| | database. |
| <u> </u> | Contact Biochemistry ext 22531/22361 to discuss results. |
| | y Molecular Genetics (Haematology) |
| Laboratory: | Specimen referred from Haematology to Cancer Molecular Diagnostics |
| Caraciana | laboratory, St. James Hospital, Dublin 8 |
| Specimen: | Blood 3mL purple Vacuette [®] (EDTA). |
| Comment: | Leukaemia: PML-RARa, MRD and Chimaerism, TCR (T cell receptor), gene rearrangements, should be requested on the advice of a consultant |
| | haematologist. |
| Turnaround: | 60 working days |
| Report: | Sent to referring clinician and copy filed in laboratory |
| | IbA1c Glycosylated Haemoglobin |
| Laboratory: | Haematology |
| Specimen: | Blood 3mL purple Vacuette® (EDTA) |
| Specimen. | Paediatric EDTA containers available from the paediatric diabetic Dept CUH, |
| | NB Primary paediatric tubes must be clearly labelled. |
| Comment: | Test available Monday to Friday, during routine working hours. As blood |
| | glucose rises, the increase in non – enzymatic glycation of proteins is |
| | proportional to both the level of glucose and the life span of the proteins in |
| | the circulation or tissues, therefore the measurement of HB A_{1c} reflects the |
| | effectiveness of treatment in diabetes mellitus. |
| | Due to elevated HbF levels this test is unsuitable for neonates and patients < 6 months |
| | Interfering haemaglobins which are not detected by the Tosoh G8 include Hb |
| | Petah Tikva. This is frequently seen in Israel. The Tosoh G8 results the HbA1c as higher. |
| Turnaround: | 24 – 48 hours |
| Ref. Range: | 20 – 42 m mol/mol (IFCC) |
| | |

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Haemoglobin A₂ Electrophoresis

| Haemoglobin A | |
|---------------|---|
| Laboratory: | Haematology |
| Specimen: | Blood 3mL purple Vacuette [®] (EDTA) |
| Comment: | Haemoglobin A_2 percentage is useful for the diagnosis of the beta |
| | thalassemias and related disorders. |
| | Test available Monday to Friday, during routine working hours. |
| Turnaround: | 1 – 2 weeks. |
| Ref. Range: | >2yrs old 2 – 3.5% |
| | at birth 0.2 – 0.3% |
| Haemoglobin F | |
| Laboratory: | Haematology |
| Specimen: | Blood 3mL purple Vacuette [®] (EDTA) |
| Comment: | Determined using HPLC / Electrophoresis Technologies. Test available |
| | Monday to Friday, during routine working hours. |
| Turnaround: | 1 – 2 weeks |
| Ref. Range: | < 2% in adults. |
| Haemoglobins | S, C, D and E Electrophoresis |
| Laboratory: | Haematology |
| Specimen: | Blood 3mL purple Vacuette [®] (EDTA). |
| Comment: | Determines the percentage of Hb S, C, D and E, that may be present in |
| | variant haemoglobins. Test available Monday to Friday, during routine |
| | working hours. |
| Turnaround: | 1 – 2 weeks |
| Ref. Range: | Normal: <1.0% |
| | Sickle Cell Screen |
| Laboratory: | Haematology |
| Specimen: | Blood 3mL purple Vacuette [®] (EDTA). |
| Comment: | Test available Monday to Friday during routine working hours. The laboratory |
| | must be contacted for all emergencies and out of hour requests. Used in |
| | screening for sickle cell disease and sickle cell trait. In the neonatal period HB |
| | F will be present in large amounts and so may mask the presence of HB S, if necessary the test should be repeated when the infant > 6 months. |
| Turnaround: | Emergency specimens: 2 hours |
| | Routine specimens: 24 hours |
| Dof Donas | |
| Ref. Range: | Positive / Negative |
| | |

Haemoglobinopathies – Haemoglobinopathy

| Laboratory: | Sample referred from Haematology to the National Haemoglobin Reference Laboratory, Oxford Haemophilia Centre, Churchill Hospital, Oxford OX3 7LJ |
|-------------|---|
| Specimen: | Example: HbE, Thalassaemias and high affinity haemoglobins |
| | Blood 3mL purple Vacuette [®] (EDTA) |
| | Due to elevated HbF levels Thalassaemia screening is unsuitable for neonates |
| | and patients < 6 months |
| Comment: | A consent form is required to perform this test. |
| | www.oxfordradcliffe.nhs.uk/molhaem (Haemoglobinopathies website) |
| | Test available Monday – Wednesday before 12.00 noon |
| Turnaround: | 12 weeks (84 working days) but may vary depending on complexity of |
| | analysis |
| Report: | Sent to referring clinician and copy filed in laboratory |

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| Laboratory: | st Blood Transfusion Laboratory | | | |
|---|--|--|--|--|
| Specimen: | 1×4 mL Clotted Sample (red cap with yellow ring) | | | |
| Comment: | Usually performed on mothers of new-born babies in the investigation of | | | |
| comment. | incompatibilities. | | | |
| | Complete the Blood Transfusion request form LF-C-BTR-XMATCH. | | | |
| | This is not an accredited test. | | | |
| Turnaround: | 3 hours | | | |
| Ref. Range: | Positive or Negative | | | |
| Haemophilia M | | | | |
| | | | | |
| Laboratory: | Referred from Haematology to HMD Laboratory, St James' Hospital | | | |
| Specimen: Comment: | 3 ml EDTA, minimum x 2 EDTA, 6 – 20 mls | | | |
| Turnaround: | By arrangement only with Haematology | | | |
| Ref. Range: | 120 working days | | | |
| - | Not applicable | | | |
| | Influenzae B Antibodies (IgG) | | | |
| Laboratory: | Clinical Biochemistry | | | |
| Specimen: | Blood 4mL red top Vacuette [®] (or similar container for clotted blood) | | | |
| Comment: | Test performed by reference laboratory (HPA Laboratory, Manchester). | | | |
| Turnaround: | 7 weeks | | | |
| Report: | Positive or negative | | | |
| - | influenzae PCR | | | |
| Laboratory: | Microbiology (Infectious Diseases Serology) | | | |
| Specimen: | 1mL EDTA blood, CSF (0.5mL) | | | |
| Comment: | Performed by Irish Meningitis & Sepsis Reference Laboratory (IMSRL), Dublin | | | |
| Turnaround: | 10 working days. Samples received by IMSRL before 11am, verbal result | | | |
| | between 4pm and 5pm the same day (positive only). | | | |
| Report: | Detected or not detected | | | |
| Hantavirus An | tibodies | | | |
| Laboratory: | Microbiology (Infectious Diseases Serology) | | | |
| Specimen: | 4mL clotted blood | | | |
| Comment: | Performed by a reference laboratory (National Virus Reference Laboratory (NVRL), Dublin) | | | |
| Turnaround: | By arrangement | | | |
| Report: | Qualitative result | | | |
| Haptoglobin | | | | |
| Laboratory: | Clinical Biochemistry | | | |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) | | | |
| Turnaround: | 4 Days | | | |
| Ref. Range: Up-to-date reference intervals will be applied to all Biochemistry reports as | | | | |
| Ken Kanye. | appropriate | | | |

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| HbA1c (POCT) | |
|-----------------|---|
| Specimen: | Whole blood sample (fingerprick) |
| | Minimum sample volume = 1 uL of whole blood |
| Time to result: | 6 minutes |
| Time to result. | o minutes |
| Ref range: | 20-42 mmol/mol (IFCC) |
| Comments: | Ideally, patients should have an individual target, balancing long-term risk of complications with quality of life and risk of hypoglycaemic events. Sending a Venous Sample to Haematology |
| | Laboratory Measurements (EDTA Sample) should be considered where: |
| | 1. Unable to obtain an adequate fingerprick blood sample. |
| | 2. Patient with severe anaemia (Hb <7 g/dL) or polycythaemia (Hb >24 g/dL) – these are unlikely in a child. |
| | 3. Patients with Haemolytic Anaemia. |
| | 4. Patients with substantial amounts of foetal heamoglobin. |
| | 5. Patients with heamoglobinopathies |
| | Result appears questionable or if the clinical signs and symptoms appear inconsistent with the result. |
| βHCG | |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Turnaround: | 1 Day (In-patients/Urgent GP samples) 4 Days (non-Urgent samples) |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as |
| | appropriate Please contact the duty biochemist (ext 22870) if requesting BHCG on |
| | patients with suspected Gestational Trophoblastic Disease. |
| Heavy Metal | Screen |
| Laboratory: | SAS Trace Element Unit, Southhampton University Hospitals |
| Specimen: | 1 ml Sod Hep Trace metal free bottle whole blood. |
| | Urine sample required for Mercury analysis |
| Turnaround: | 1-2 weeks |
| Ref. Range: | See report form. |
| | |

Helicobacter pylori Antibodies

This test is not available at the CUH laboratories.

Helicobacter pylori Culture and Sensitivity

| Microbiology (Main laboratory) |
|--|
| Specimens will only be processed by prior arrangement with the laboratory. |
| As media must be freshly prepared a minimum of 48 hours notice is required for preparation of media, reagents <i>etc</i> . Two gastric biopsy |
| specimens, one from the antrum and one from the body of the stomach, are |
| taken during endoscopy, for culture. The biopsies are immediately introduced into transport medium, supplied by the laboratory, and sent directly to the |
| Microbiology laboratory where they are processed immediately. Preferably patients should have ceased antimicrobial therapy and PPI therapy two weeks prior to endoscopy. |
| |

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| Comment: | transport cannot be hospitals), the spec looses viability at ro Please include any a | avoided (speci imens must be oom temperatu appropriate clin cs previously a | mens being trans packed on ice. No re and when expo ical details, e.g. p dministered. Pleas | previous therapy failure, se state if the patient wa |
| Turnaround: | Turnaround: Prelim report: 7 days, Final report: 14 days in cases where patients were taking antimicrobial agents at the time the biopsies were obtained. | | | |
| Report: | Culture with the app | propriate sensit | ivities | |
| leparin Assay | | • | | |
| Laboratory: | Haematology | | | |
| Specimen: | Blood 3mL, blue Va | cuette® (sodiu | m citrate 3.2%) | |
| | Specimens which ar analysed, check coa filling. | e haemolysed, igulation sampl | underfilled or ove e bottles are not | expired to ensure correc |
| Comment: | Used to monitor the effectiveness of low molecular weight heparin therapy. It is essential to state the details of the type of low molecular weight heparin (LMWH) on the request form. Test performed once weekly (presently Wednesdays) Specimen must be taken: 4 hours post administration. | | | |
| Turnaround: | 1 week. | | pose duministrati | |
| Ref. Range: | Refer to report | | | |
| | Antibody Test (HIT; | Henarin Induc | ed Thrombocyton | enia screening test) |
| Laboratory: | | | | gy laboratory staff durin |
| | Specimens are reference Coagulation Laborat | ory, St., Jame | s Hospital, Dublin | 8 |
| Specimen: | | • | • | ainer for clotted blood) |
| Comment: | | - | | of the anticoagulation |
| | history of the patier | | | |
| | 4T Score MUST be s | | • | |
| | HIT request form m | | | Deferral/DeferralForme/ |
| | · · · · · · · · · · · · · · · · · · · | | | <u>Referral/ReferralForms/</u> 20August%202015.pdf |
| Turnaround: | ELISA Test (referral | | | <u>20August /0202015.pur</u> |
| Report: | Sent to referring cli | | | |
| lepatitis A IgN | | | | У |
| | | | Corology | |
| Laboratory: | Microbiology (Infect 4mL clotted blood | ious Diseases s | Servicy) | |
| Specimen: Comment: | A qualitative test fo be used as an aid in Hepatitis A IgM test <14yrs or on sampl Otherwise request v | the diagnosis ing is only rout es from people vith a full patie vity should be | of acute or recent inely performed of recently returned nt history or in ou correlated with pa | Itbreak situations. Atient history and other |
| Turnaround: Report: | 36 hours Qualitative result | | | |

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Hepatitis A IgG Antibody

| перация А 190 | 5 Antibody |
|-----------------|--|
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Comment: | Test is used to determine the immune status to hepatitis A and is often used to monitor the success of hepatitis A vaccination. It is often performed prior to vaccination in certain risk groups, e.g., army personnel going on overseas duty. |
| Turnaround: | 36 hours |
| Report: | Qualitative result |
| Hepatitis B Aus | stralia Antibody (Anti-HBs) |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Turnaround: | Routine: 36 hours. Urgent: within 2 hours of receipt. |
| Report: | Quantitative value (mIU/mL) |
| Comment: | This test is used to check the immune status to hepatitis B and is often used to monitor the success of hepatitis B vaccination. Please indicate patient vaccination history on the request form. |

Management Following Post-Vaccination Testing:

| Anti-HBs Level | Action Required |
|----------------|---|
| ≥10 mIU/mL | Good response. No further action required. |
| | Non-responder. Test for anti-HBc and HBsAg. If anti-HBc and HBsAg negative, repeat course of hepatitis B vaccine (use a different brand). |
| <10 mIU/mL | Recheck anti-HBs 2 months later and if anti-HBs remains <10 mIU/mL, consider further vaccination as per national guidelines. |
| | Recheck anti-HBs 2 months later and if anti-HBs remains <10 mIU/mL, person is susceptible to HBV. |

Source: National Immunisation Guidelines (June 2020)

| Hepatitis B Core Antibody (Anti-HBc) | | |
|--------------------------------------|---|--|
| Laboratory: | Microbiology (Infectious Diseases Serology) | |
| Specimen: | 4mL clotted blood | |
| Comment: | Test will detect total antibody to hepatitis B core antigen, i.e., IgM and/or IgG. A positive result indicates present or past infection with the hepatitis B virus. This test should be interpreted in conjunction with other hepatitis B markers. | |
| Turnaround: | 36 hours | |
| Report: | Qualitative result | |

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| Hepatitis B Sur | face Antigen |
|--------------------------|---|
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Comment: | A positive result may indicate acute or chronic carriage of the hepatitis B virus. Positive specimens are considered presumptive positive only and a repeat specimen is requested. Positive specimens are tested with a full hepatitis B virus marker profile, which includes anti-HBc, HBeAg, anti-HBe and anti-HBs. |
| Turnaround: | Routine: 36 hours. Urgent: within 2 hours of receipt. |
| Report: | Qualitative result |
| Hepatitis C Ant | |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Comment: | Positive specimens are considered presumptive positive only and a repeat specimen is requested. All new positives are referred to National Virus Reference Laboratory (NVRL) in Dublin for confirmation. |
| Turnaround: | Routine: 36 hours. Urgent: within 2 hours of receipt. Please allow more time for samples testing positive in house. |
| Report: | Qualitative result |
| Hepatitis C Ant | |
| Laboratory: Specimen: | Microbiology (Infectious Diseases Serology) 4mL clotted blood |
| Comment: | Test performed weekly. This test is restricted to dialysis patients. A repeat |
| comment. | sample is requested for all new positives. |
| Turnaround: | 8 working days |
| Report: | Qualitative result |
| Hepatitis D Ant | |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Comment: | Hepatitis delta virus (HDV) is in fact a sub-viral particle that relies on hepatit |
| | B virus (HBV) to cause infection in humans. |
| | Performed by a reference laboratory (National Virus Reference Laboratory (NVRL), Dublin). |
| Turnaround: | 14 working days |
| Report: | Qualitative result |
| Hepatitis E IgG | |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Comment: | Performed by a reference laboratory (National Virus Reference Laboratory (NVRL), Dublin) |
| Turnaround: | 14 working days |
| Report: | Qualitative result |
| Hepatitis E IgM | |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| • | |
| Comment: | Performed by a reference laboratory (National Virus Reference Laboratory (NVRL), Dublin) |
| • | , |

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| Hepatitis Scree | en |
|---------------------------------------|--|
| | B Surface Antigen and Hepatitis C Antibody |
| · · · · · · · · · · · · · · · · · · · | ver Syndromes (FMF, TRAPS) |
| Laboratory: | Referred from Molecular Genetics Lab in Biochemistry to National Amyloidosis |
| | Centre at UCL |
| Specimen: | 3ml EDTA blood + 3ml Serum |
| Comment: | Special request form available from ext 22531 |
| | Please note: invoices will be issued to the referring clinician for tests not performed in NCMG. |
| Turnaround: | 4-6 weeks |
| Report: | Sent to referring clinician and copy filed in pathology |
| Herpes Simple | |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Comment: | Performed by a reference laboratory (National Virus Reference Laboratory (NVRL), Dublin) |
| Turnaround: | 14 working days |
| Report: | Qualitative result |
| Herpes Simple | x Virus 1/2 Molecular |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | Viral swab, CSF, nasopharyngeal aspirate, sputum, broncho-alveolar lavage, urine, 4mL EDTA blood |
| Comment: | Performed by a reference laboratory (National Virus Reference Laboratory (NVRL), Dublin) |
| Turnaround: | 14 working days |
| Report: | Detected or not detected |
| 5-HIAA | |
| Laboratory: | Sample referred from Clinical Biochemistry to Beaumont hospital. |
| Specimen: | 24-hour urine sample collected into a container, which has acid, added. |
| | 24 hr urine containers are available from stores; acid is added in the |
| | Biochemistry lab. Avoid following foods for 48h before collection: bananas, |
| | chocolate, tomatoes, grapefruit, walnuts, avocado, pineapple, plums, dried |
| - · | fruit, citrus fruit, tea and coffee |
| Turnaround: | 3 weeks |
| Ref. Range: | See report form. |
| | ipoprotein (HDL) |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Turnaround: | A/E or urgent sample: - 1 hour 30 mins. CUH wards, CUMH, SI, SF, SMOH, |
| Ref. Range: | MGH: - 3 hours 140egain140. GP or OPD- Results posted within 4 days. Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate. Target values apply to pts at low or moderate risk CVD |
| | |

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| High Vaginal C | |
|------------------------|--|
| High Vaginal S | |
| Laboratory: | Microbiology (Main laboratory) |
| Specimen: | It is important to avoid vulval contamination of the swab. The posterior fornix, including any obvious 141egain141 plaques should be swabbed. Low vaginal swabs are discouraged because the presence of high numbers of commensal flora makes them difficult to interpret (see Low Vaginal Swab for investigation of vulvo-vaginitis in paediatric patients). Only swabs sent in suitable transport medium will be processed – swabs that are sent without transport medium may be dry and may not yield the targeted organisms. Transport specimens ASAP. If processing is delayed, refrigeration is preferable to storage at ambient temperature. |
| Comment: | Specimens are generally examined for the presence of Candida or Group B Streptococci. Specimens will be processed for Trichomonas vaginalis if requested. Bacterial Vaginosis (BV) slide for women aged 12-55 years. Not routinely processed on Antenatal patients unless BV specifically requested or such clinical details given as spontaneous rupture of membranes (SROM), premature rupture of membranes (PPROM) or miscarriage. Please indicate on the request form if the specimen is post-operative /post delivery so that supplementary testing can be performed. Vaginal swabs are not recommended for gonococcal culture on adults; an endocervical specimen is more appropriate. A separate specimen of urine or specific swabs and transport medium should be collected for the detection of <i>C. trachomatis</i>. |
| Turnaround: Report: | Prelim: 24 hours; Final: 48-72 hours Microscopy: WBCs, yeasts, trichomonads and clue cells if present. Excess pus cells suggest infection; motile trichomonads indicate trichomoniasis, yeasts and hyphae suggest Candidiasis; clue cells in the absence of normal flora is suggestive of anaerobic vaginosis. |
| Listone Antika | Culture: Any clinically significant isolate with the appropriate sensitivities. |
| Histone Antibo | Sample referred from Autoimmune Serology to Eurofins-Biomnis |
| Laboratory: | Laboratories. |
| Specimen: | Blood, 4mL red top Vacuette (or similar container for clotted blood) |
| Turnaround: | Approx. 3 Weeks |
| Ref. Range: | See report form, or visit internet site <u>https://www</u> .eurofins.ie/biomnis/ for up to |
| | date referral test information. |
| Histoplasma A | |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: Comment: | 4mL clotted blood Performed by a reference laboratory (UKHSA Mycology Reference Laboratory, Bristol) |
| Turnaround: | 28 working days |
| Report: | Qualitative result |

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| HLA B27 Typin | a |
|----------------------------|---|
| Laboratory: | Blood Transfusion Laboratory |
| Specimen: | 1x 3 ml EDTA purple cap (FBC) tube. |
| Comment: | Complete the Blood Transfusion request form clearly indicating that consent |
| | for the test has been obtained from the patient. Samples received without |
| | confirmation of consent cannot be processed. |
| | A specific consent form is available from the Blood Transfusion Laboratory or |
| | available on the CUH website |
| | http://www.cuh.hse.ie/Our-Services/Our-Specialities-A-Z-/Laboratory- |
| | Medicine/Services-Provided/Downloads/Molecular-Genetics-Request-for-HLA- |
| | <u>B27.pdf</u> |
| Turne e ne une du | This is an INAB accredited test. |
| Turnaround: | 3 weeks |
| Ref. Range: Limitations | Not applicable. The primers used in the test kit used by the laboratory are expected to miss |
| LIIIItations | the following HLA B27 alleles: B*27:04:03, B*27:07:01, B*27:07:02, |
| | B*27:07:03, B*27:07:04, B*27:102, B*27:11, B*27:125, B*27:14, |
| | B*27:19, B*27:20, B*27:21, B*27:24, B*27:30, B*27:32, B*27:33, |
| | B*27:34, B*27:36, B*27:43, B*27:70, B*27:81, B*27:90:01, B*27:90:02. |
| HLA Typing Cla | iss I and Class II (pre-Bone Marrow Transplant) |
| Laboratory: | Blood Transfusion Laboratory |
| Specimen: | 3×4 ml EDTA purple cap (FBC) tube. Arrange for samples to be delivered to |
| _ | laboratory between Monday to Thursday. |
| Comment: | HLA typing referred to: HLA Department, I.B.T.S., National Blood Centre, |
| | James's St., Dublin 8. Mon. to Thurs. |
| | Complete the Blood Transfusion request forms LF-C-BTR-ANTENAT or LF-C-BTR-XMATCH |
| | This is not an INAB accredited test. |
| Turnaround: | 3 weeks |
| Ref. Range: | Not applicable. |
| | isease Association e.g. HLA DQ2, HLA DQ8) |
| Laboratory: | Blood Transfusion Laboratory |
| Specimen: | 3×4 ml EDTA purple cap (FBC) tube. Arrange for samples to be delivered to |
| | laboratory between Monday to Thursday. |
| Comment: | HLA typing referred to: HLA Department, I.B.T.S., National Blood Centre, |
| | James's St., Dublin 8. Mon. to Thurs. |
| | Complete the Blood Transfusion request forms LF-C-BTR-ANTENAT or |
| | LF-C-BTR-XMATCH |
| - . | This is not an INAB accredited test. |
| Turnaround: | 3 Weeks |
| Ref. Range: | Not Applicable |

HLA Typing (re: Solid Organ Transplant)

| Laboratory: | Blood Transfusion Laboratory |
|-------------|--|
| Specimen: | 10 ml Citrate (blue cap bottle). 7.5 ml EDTA (purple cap bottle), 10 ml clotted sample (red cap bottle). |
| Comment: | This test is carried out by Histocompatibility and Immunogenetics Laboratory, Beaumont Hospital, Dublin 9. |
| | Complete the Blood Transfusion request forms LF-C-BTR-ANTENAT or LF-C-BTR-XMATCH or equivalent. |
| | · |
| | This is not an INAB accredited test. |

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| | | Author: | Mr Paul Cantwell | | |
| Turnaround: | Contact Histocompa Hospital, Dublin 9. | tibility and Imn | nunogenetics Labo | oratory, Beaumont | |
| Ref. Range: | Not Applicable | | | | |
| HLH Granule | e release assay (Hae | emophagocyti | c Lympho Histoc | cytosis) | |
| Laboratory: | Referred from Haematology to Great Ormond Street Hospital | | | | |
| Specimen: | EDTA x 5mls | | | | |
| Comment: | Consultant sending sample for these assays needs to contact Great Ormond | | | | |
| | • | | | equest form must be | |
| | completed, available on Great Ormond street website | | | | |
| Turnaround: | 20 working days | | | | |
| Report: | Sent to referring clinician and copy filed in laboratory | | | | |
| | · Free and Total (Pa | | | | |
| Laboratory: | Sample referred from Clinical Biochemistry to The Children's Hospital, Temple Street, Dublin | | | | |
| Specimen: | Lithium Heparin sample which must be separated within 10 minutes of collection. Time must be stated on bottle and on form | | | | |
| Comment: | Please advise the lab in advance | | | | |
| Turnaround: | 1 week | | | | |
| Ref. Range: | See report or contact | ct Biochemistry | Laboratory, Temp | ole Street Hospital | |
| HMGCoAR Anti | bodies | | | | |
| | Sample referred from munology | Clinical Biochen | nistry to Oxford D | epartment of Clinical | |
| Specimen: 1 | l ml serum FROZEN | | | | |
| Turnaround: 3 | 3 weeks from receipt in Referral Laboratory | | | | |
| | See report form or cor 0) 1865225995 | ntact Oxford De | partment of Clinic | al Immunology, ph: +44 | |
| HMMA (VMA) | | | | | |
| Laboratory: | Sample referred fro | m Clinical Bioch | nemistry to BEAUM | 10NT Hospital Dublin | |
| Specimen: | Spot urine sample. immediately to have | | e brought to Bioch | nemistry laboratory | |
| Turnaround: | 2 weeks | | | | |
| Ref. Range: | See report form or o | contact Biochen | nistry Laboratory I | BEAUMONT Hospital | |
| HPA (Human P | Platelet Antigen + A | ntibody Inves | tigation for NAI | TP) | |
| Laboratory: | Blood Transfusion L | aboratory | | | |
| Specimen: | Baby: 1 mL EDTA | | | | |
| | Mother: 5 mL EDTA | and 20 mL Clo | otted | | |
| | Father: 20 mL EDT | A | | | |
| Comment: | Only by prior arrang Complete Form NBC CUH) | • | | boratory, CUH Transfusion Laboratory, | |
| | Referred to: I.B.T.S | National Bloo | d Centre James's | St., Dublin 8 | |
| | This is not an accred | | | | |
| Turnaround: | Refer to IBTS, Dubli | | | | |
| Ref. Range: | Refer to IBTS, Dubli | | | | |
| iten itunger | | | | | |

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| HTLV-I/II Ant | ibodies | | | |
|--------------------------|--|--|--|--|
| Laboratory: | Microbiology (Infectious Diseases Serology) | | | |
| Specimen: | 4mL clotted blood | | | |
| Comment: | Performed by a reference laboratory (National Virus Reference Laboratory (NVRL), Dubln) | | | |
| Turnaround: | 14 working days | | | |
| Report: | Qualitative result | | | |
| Human Herpes | S Virus 6 (HHV-6) Molecular | | | |
| Laboratory: | Microbiology (Infectious Diseases Serology) | | | |
| Specimen: | 4mL clotted blood, 4mL EDTA blood, CSF, saliva | | | |
| Comment: | Performed by a reference laboratory (National Virus Reference Laboratory (NVRL), Dublin) | | | |
| Turnaround: | 14 working days | | | |
| Report: | Detected or not detected | | | |
| | SVirus 8 (HHV-8) Molecular | | | |
| Laboratory: | Microbiology (Infectious Diseases Serology) | | | |
| Specimen: | 4mL EDTA blood | | | |
| Comment: | Test performed by a reference laboratory (Virus Reference Department, London) | | | |
| Turnaround: | 28 working days | | | |
| Report: | Detected or not detected | | | |
| | nodeficiency Virus (HIV) Serology | | | |
| Laboratory: | Microbiology (Infectious Diseases Serology) | | | |
| Specimen: | 4mL clotted blood | | | |
| Comment: | Detects HIV antigen and antibody to HIV1 and HIV2. | | | |
| | Positive specimens are referred to the National Virus Reference Laboratory, University College Dublin, for confirmation. | | | |
| Turnaround: | Negative samples: 36 hours | | | |
| . . | Samples positive in house: 14 working days (confirmation required) | | | |
| Report: | Qualitative result | | | |
| HVA | | | | |
| Laboratory: Specimen: | Sample referred from Clinical Biochemistry to BEAUMONT Hospital Dublin Spot urine sample. Sample must be brought to Biochemistry laboratory immediately to have acid added | | | |
| Turnaround: | 2 weeks | | | |
| Ref. Range: | See report form or contact Biochemistry Laboratory BEAUMONT Hospital | | | |
| Hydatid Cyst | | | | |
| See Echinoco | ccus Antibodies | | | |
| 3 Hydroxybuty | vrate (3HB/Blood Ketones) | | | |
| Laboratory: | Sample referred from Clinical Biochemistry to Sheffield Children's NHS | | | |
| Specimen: | 1.2 ml Fluoride oxalate plasma | | | |
| Comment: | 0.5ml min | | | |
| Turnaround: | 4 weeks | | | |
| Ref. Range: | See report form | | | |
| | sterone (Alpha 17-Hydroxyprogesterone) | | | |
| Laboratory: | Sample referred from Clinical Biochemistry to St James Hospital, Dublin | | | |
| Specimen: | 2.0 mL blood in a plain tube (clotted sample) | | | |
| Comment: | Consultant request only | | | |
| Turnaround: | 4 weeks | | | |
| Ref. Range: | See report form | | | |

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| Hydroxyproges | Hydroxyprogesterone (Alpha 17-Hydroxyprogesterone) Blood Spots | | | | |
|-----------------|---|--|--|--|--|
| Laboratory: | Sample referred from Clinical Biochemistry to University Hospital of Wales. | | | | |
| Specimen: | Blood spots taken at 4 points through the day. See comment. | | | | |
| Comment: | Consultant request only | | | | |
| Turnaround: | 3 – 4 weeks | | | | |
| Ref. Range: | Contact laboratory | | | | |
| - | | | | | |
| IgD | | | | | |
| Laboratory: | Sample referred to Sheffield Protein Reference Unit. | | | | |
| Specimen: | 4.0 mL blood in a plain tube (clotted sample) | | | | |
| Comment: | Consultant request only | | | | |
| Turnaround: | 6 weeks | | | | |
| Ref. Range: | See report form | | | | |
| IgE Total and S | IgE Total and Specific | | | | |
| | | | | | |

| Laboratory: | Clinical Biochemistry | |
|----------------|---|--|
| Specimen: | 4.0 mL blood in plain tube (clotted sample) | |
| Turnaround: | Up to 14 Days | |
| Ref. Range: | Contact CUH Biochemistry Laboratory | |
| IgG Subclasses | | |

| Laboratory: | Sample referred to Eurofins-Biomnis Laboratories |
|-------------|---|
| Specimen: | 4.0 mL blood in a plain tube (clotted sample) |
| Comment: | Consultant request only |
| Turnaround: | 3 weeks |
| Ref. Range: | See report form, or visit internet site https://www.eurofins.ie/biomnis/ for up to |
| - | date referral test information. |
| | |

Immunoglobulin gene rearrangements (Clonality studies)

| Labor | atory: | Sample referred from Pathology to CMD, St. James Hospital |
|--------|----------|--|
| Speci | men: | FFPE tissue block |
| Turna | round: | 19 working days (from date testing material is sent to referral lab) |
| Ref. R | Range: | Not applicable |
| Imm | unoglob | ulins / Electrophoreisis |
| Lab | oratory: | Clinical Biochemistry |

Specimen:4.0 mL blood a plain tube (clotted sample)Comment:Age related reference values are available from Laboratory on requestTurnaround:5 Days * Note additional testing such as Immunofixation and/or serum free light chain
analysis may increase the turnaround timeRef. Range:Up-to-date reference intervals will be applied to all Biochemistry reports as
appropriate.Infectious Mononucleosis Screening test

Laboratory: Haematology Specimen: EDTA specimen Comment: This test is only performed if the results of the Full Blood Count and/or manual differential suggest Infectious Mononucleosis, clinicians are requested to send a confirmatory test to Clinical Microbiology for EBV status on all positive screens. Comment added to all Negative results: A negative Monospot screen does not preclude IM infection. Result must be interpreted in conjunction with clinical details. Turnaround: Not applicable Report: Positive or Negative

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| ional Normalised Ratio) | | |
|---|--|--|
| Haematology: See Prothrombin Time (PT) | | |
| ation for Her2:Chromosome 17 ratio | | |
| Histopathology | | |
| Formalin Fixed Paraffin Embedded Tissue. | | |
| This test is performed on a subset of breast and gastric cancer cases and | | |
| other cases as required. | | |
| 10 working days | | |
| Report is expressed as a ratio of Her 2 gene copy number divided by Chromosome 17 copy number. | | |
| Antibodies | | |
| Haematology | | |
| Blood 4mL Red Vacuette [®] (clotted blood). | | |
| Test available Monday to Friday, during routine working hours. | | |
| Tests for IF antibodies are carried out on patients with suspected | | |
| megaloblastic anaemia and a depressed serum vitamin B ₁₂ to aid in the | | |
| diagnosis of pernicious anaemia. | | |
| Free B12 levels of >444 ng/L can give false positive results. | | |
| 7 working days | | |
| Negative / Indeterminate / Positive | | |
| | | |
| Clinical Biochemistry | | |
| 2 mL blood in a plain tube (clotted sample) | | |
| Consultant request only | | |
| 7 days | | |
| Insulin levels should be appropriate for the glucose level at the time the sample was taken. Glucose should always be measured at the same time as the insulin to facilitate interpretation of results. | | |
| Haemolysed sample unsuitable. Urgents available on request | | |
| ies | | |
| Sample referred from Autoimmune Serology to Eurofins-Biomnis Laboratories. | | |
| Blood, 4 mL red top Vacuette (or similar container for clotted blood) | | |
| Approximately 3 Weeks | | |
| See report form, or visit internet site <u>https://www</u> .eurofins.ie/biomnis/ for up to date referral test information. | | |
| wth Factor 1 | | |
| Clinical Biochemistry | | |
| 4.0 mL blood in a plain tube (clotted sample), fresh sample. | | |
| Haemolysed samples should be interpreted with care. | | |
| Samples should be transported to the laboratory as soon as possible and | | |
| must be frozen within 24hours | | |
| 2 weeks | | |
| Age and gender based. See report. | | |
| rowth Factor BP3 (IGBP3) | | |
| | | |
| Sample referred from Biochemistry to SAS Peptide Hormones Section, Royal Surrey County Hospital, Guildford, Surrey. | | |
| Sample referred from Biochemistry to SAS Peptide Hormones Section, Royal | | |
| Sample referred from Biochemistry to SAS Peptide Hormones Section, Royal Surrey County Hospital, Guildford, Surrey. | | |
| | | |

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Intraocular Fluids / Corneal Scrapings

Laboratory: Microbiology (Main laboratory)

Specimen: Specialist collection according to local protocols – An ophthalmic surgeon will collect corneal scrapings and intraocular fluids. Because of the small amounts of material involved, initial inoculation of culture media and preparation of slides may need to be done at the patient's side.

The laboratory, in conjunction with local ophthalmologists, has agreed the following protocol for the collection of specimens, inoculation of media, and transport to the laboratory: Corneal scrapings:

Scrapings should be taken aseptically (e.g. sterile scalpel blade)

Aseptically remove the cap of the nutrient broth.

Carefully, dip the tip of the scalpel, which contains the scrapings, into the broth and agitate gently.

Ensure that the scraping has been removed and discard the scalpel into a sharps bin. Close the lid on the nutrient broth, label as appropriate, and send to the laboratory immediately.

If Acanthamoeba keratitis is considered, please supplement the above by an additional scraping taken in the same fashion but placed on PCR swab (obtained from Microbiology laboratory, refer to Acanthaoemba above). Send to the laboratory with the appropriately completed form – the laboratory must be notified in advance. The contact lens case and rinse fluids should also be sent to the laboratory.

Intraocular fluids:

Intraocular fluids which have been taken aseptically should be injected directly into an **equal volume** of nutrient broth, labelled as appropriate and sent to the laboratory as soon as possible with an appropriately labelled form.

| P | | | |
|---------------|---|--|--|
| Comment: | Test performed routinely Monday to Friday 9-5pm or by urgent request. | | |
| Turnaround: | Prelim: 24 hours; Final: 48-72 hours | | |
| Report: | Culture: Any clinically significant isolate with the appropriate sensitivities. | | |
| Intra-Uterine | Contraceptive Device (IUCD) | | |
| Laboratory: | Microbiology (Main laboratory) | | |
| Specimen: | IUCDs should only be sent if clinical suspicion of infection exists. | | |
| | Place the entire IUCD, including any exudate, in a clean, sterile, leakproof container and transport ASAP. Specimen should be delivered to the laboratory as soon as possible to protect the viability of fragile organisms such as <i>Neisseria</i> spp. | | |
| Comment: | Test performed Monday to Friday 9-5pm. | | |
| Turnaround: | Prelim: 24 hours; Final: 48 – 72 hours. <i>Note:</i> Culture for Actinomycosis takes up to 17 days. | | |
| Report: | Any clinically significant isolate with the appropriate sensitivities. Culture for <i>Actinomyces</i> spp. Proceeding which will be reported if positive. | | |
| Intra-Uterine | Infection Screen / TORCH Screen | | |
| Laboratory: | Microbiology (Infectious Diseases Serology) | | |
| Specimen: | 4mL clotted blood (Minimum volume for baby specimens: 1mL) | | |
| Tests: | Toxoplasma gondii IgM, rubella IgM, CMV IgM and parvovirus B19 IgM | | |
| Turnaround: | 36 hours. | | |
| | Positive Toxoplasma IgM result must be confirmed by a reference laboratory – 28 working days. | | |
| Report: | Qualitative result | | |
| Intravascular | Cannulae – Culture | | |
| See Catheter | / Intravascular Cannulae | | |

See Catheter / Intravascular Cannulae

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| Iron | | | |
|--------------------------|--|--|--|
| Laboratory: | Clinical Biochemistry | | |
| Specimen: | | | |
| Comment: | Marked haemolysis invalidates the result | | |
| Turnaround: | 4 Days | | |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate. | | |
| JC Virus Moleo | | | |
| Laboratory: | Microbiology (Infectious Diseases Serology) | | |
| Specimen: | 4mL clotted blood, 4mL EDTA blood, CSF, urine, brain tissue | | |
| Comment: | Performed by a reference laboratory (National Virus Reference Laboratory (NVRL), Dublin) | | |
| Turnaround: | 14 working days | | |
| Report: | Detected or not detected | | |
| JAK2 in MPD (| and CALR) | | |
| Laboratory: | Referred from Haematology Dept. to CMD in St James Hospital, Mon to Thurs to reach haematology lab by 12 noon, | | |
| Specimen: | Blood 9mLs, $3mL \ge 3$ purple (may also use $6mL$ Purple), Vacuette [®] (EDTA) of Bone Marrow in 10mls in RPMI | | |
| Comment: | Mutation analysis in MPD | | |
| Turnaround: | 60 working days | | |
| Report: | Sent to referring clinician and copy filed in laboratory | | |
| JAK2 Exon 12 | mutation | | |
| Laboratory: Specimen: | Referred from Haematology Dept. to Oncology Cytogenetics, 5 th Floor Tower Wing, Guy's Hospital, Great Maze Pond, London SE1 9RT, Mon to Thurs to reach haematology lab by 12 noon. Whole blood 3mL, purple, Vacuette [®] (EDTA) or Bone Marrow in 10mls in | | |
| | RPMI | | |
| Turnaround: | 64 days | | |
| Report: | Sent to referring clinician and copy filed in laboratory | | |
| loint Aspirate | for Crystals | | |
| Laboratory: | Histopathology (Cytology Department) | | |
| Specimen: | Joint Fluid | | |
| Comment: | Tests are performed routinely Monday to Friday during routine working hours | | |
| Turnaround: | Can be immediate if urgently requested by prior communication, routine 1-2 days | | |
| Ref. Range: | Not applicable | | |
| Joint Fluid – M | | | |
| | ody Fluid – Microscopy and Culture. | | |
| | see Chromosome analysis) | | |
| Keppra (Leveti | | | |
| Laboratory: Specimen: | Sample referred from Clinical Biochemistry to Birmingham City Hospital EDTA plasma | | |
| Comment: | Keppra is indicated as monotherapy in the treatment of partial onset seizures with or without secondary generalisation in adults and adolescents from 16 years of age with newly diagnosed epilepsy | | |
| Turnaround: | 5 days from receipt in referral laboratory | | |
| Ref. Range: | Not applicable | | |

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| Kleihauer Test | for Foetal Cells FMH |
|----------------|--|
| Laboratory: | Haematology, and bleeds of \geq 4mls in postnatal patients are referred to |
| - | Rotunda Hospital for flow Cytometry |
| Specimen: | Blood 3mL purple Vacuette [®] (EDTA) |
| Comment: | Test available Monday – Friday during routine working hours, and Sunday of bank Holiday weekends. For all other emergencies a Consultant to Haematology Consultant request is required. It is a procedure that identifies individual cells containing HB F. It has proved |
| | useful in determining the extent of foetal bleed into the maternal circulation, and can be used to calculate the dose of Anti-D to be administered to the patient. Kleihauer test is only validated for the administration of Anti-D to Rh Neg mothers. Kleihauer test is not performed on Rhesus Positive women except in cases of Women who have had a late intrauterine foetal death (IUFD) after 18 completed weeks of pregnancy. |
| | All postnatal samples with Bleeds ≥4mls in postnatal patients are referred to the Rotunda Hospital for flow cytometry. Antenatal patients with bleeds ≥4mls are NOT referred. Flow cytometry in Rotunda is currently not validated for antenatal patients. Kleihauer on a rhesus D Neg mother of a baby with a weak D Ag are NOT referred. |
| - · | >12ml bleeds are phoned to requesting ward |
| Turnaround: | Emergency specimens: <2 hours |
| | Routine specimens: 24 – 72 hours. |
| Ref. Range: | To calculate dosage of Anti-D required refer to CUMH Anti-D dosage Policy. |
| Lacrimal (Tear | - |
| Laboratory: | Microbiology (Main laboratory) |
| Specimen: | Stones / secretions should be collected into a clean, sterile, leakproof container and immediately transported to the laboratory. |
| Comment: | Test performed routinely Monday to Friday 9-5pm or by urgent request. |
| Turnaround: | Prelim: 24 hours; Final: 48-72 hours |
| Report: | Culture report: Any clinically significant isolate with the appropriate sensitivities. |
| Lactate | |
| Laboratory: | Clinical Biochemistry |
| Specimen: | Blood in Fluoride Oxalate tube, on ice |
| Turnaround | 2 hours |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate. |
| Lactate dehydi | rogenase (LDH) |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Comment: | Haemolysis invalidates result |
| Turnaround: | A/E or urgent sample: - 1 hour 30 mins .CUH wards, CUMH, SI, SF, SMOH, MGH: - 3 hours 149egain149. GP or OPD- Results posted within 4 days. |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate. |
| La (SS-B) | |
| Laboratory: | Autoimmune Serology |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) |
| Comment: | Qualitative Elisa assay; automatically undertaken on all Anti-ENA positive sera. |
| Turnaround: | 72 Hours |
| Ref. Range: | Not applicable |

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| Lamotrigine | (Lamictal) |
|---------------|---|
| Laboratory: | Referred from Clinical Biochemistry to Birmingham City Hospital |
| Specimen: | EDTA plasma |
| Comment: | Monitoring levels of Lamotrigine, antiepileptic drug which can induce allergic |
| | reactions, especially when taken at the same time as sodium valproate. |
| Turnaround: | I week from receipt in Referral Laboratory |
| Ref. Range: | See report or contact Referral laboratory Birmingham City Hospital, ph: +44 (0) 121 |
| | 507 4271, +44 (0) 121 507 4138 |
| Lead | |
| Laboratory: | Referred from Clinical Biochemistry to SAS Laboratory for Trace Elements, |
| | Guildford |
| Specimen: | Sod Hep trace metal free tube (navy top) |
| Turnaround: | |
| Ref. Range: | |
| Leishmania A | appropriate |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Comment: | Performed by a reference laboratory (National Parasitology Reference |
| comment. | Laboratory (NPRL), London) |
| Turnaround: | |
| Report: | Qualitative result |
| Leptospira Ig | |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Comment: | Performed by a reference laboratory (National Virus Reference Laboratory |
| | (NVRL), Dublin) |
| Turnaround: | 5 1 7 |
| | Samples requiring confirmatory testing: 28 working days |
| Report: | Qualitative result |
| | Vhite Cell) Antibody Investigation |
| Laboratory: | Blood Transfusion Laboratory |
| Specimen: | 1 x 4 mL Clotted (Red Capped/Yellow Ring) Tube |
| Comment: | Samples referred to: I.B.T.S., National Blood Centre, James's St., Dublin 8. |
| | Complete the Blood Transfusion request form LF-C-BTR-XMATCH or LF-C- |
| | BTR-ANTENAT. |
| | This is not an INAB accredited test. |
| Turnaround: | |
| Ref. Range: | Not Applicable |
| LH | |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Turnaround: | |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate. |
| Lithium | / |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in a plain tube (clotted sample) |
| Comment: | Sample 12 hours post dose (trough sample) |
| Turnaround: | |
| | , |

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| Ref. Range: | levels up to 1.2 mm Up-to-date reference appropriate. | nol/L ce intervals will | | e therapy may require Nochemistry reports as |
| | r Copper /Iron Est | | | |
| Laboratory: | College Hospital, Lo | | | ements Laboratory, King |
| Specimen: | Liver Biopsy unfixed | t | | |
| Comment: | label a sterile univertype and date samp filter paper, (larger distilled water only, or leaching out of context histology, use a new piece for histology i | may be invalid rsal container w ble is taken. Play pieces do not n as the use of f ertain elements w scalpel blade s placed in a se | due to liver non-h vith Patients name ce the biopsy betw need to be on filter ormalin or saline of . If the specimen and divide the sar cond clearly labell | a, he least i climb nomogeneity). Clearly e, date of birth, specime veen two pieces of 2.5cm paper), moistened <u>with</u> can lead to contaminatio is to be divided eg for nple in two. The second led container in neutral Histology laboratory. |
| Turnananal | | | | |
| Turnaround: | 4-6 weeks (from da | - | rial is sent to rere | rral institution) |
| Ref. Range: | 20-50 µg/g Dry We | | | |
| | Iney Microsome An | | | |
| Laboratory: | Autoimmune Serolo | • · | | |
| Specimen: | Blood, 4 mL red top | • | | or clotted blood) |
| Comment: | Reported if seen on | Autoantibody S | Screen. | |
| Turnaround: | 24 Hours | | | |
| Ref. Range: | Not applicable | | | |
| Low Density lip | ooprotein (LDL) | | | |
| Laboratory: | Clinical Biochemistr | У | | |
| Specimen: | 4.0 mL blood in pla | in tube (clotted | sample) | |
| Comment: | Calculation. Result | s not reported i | f Triglyceride > 4. | 5 mmol/L |
| Turnaround: | MGH: - 3 hours 151 | Legain151. GP | or OPD- Results p | CUMH, SI, SF, SMOH, osted within 4 days. |
| Ref. Range: | Up-to-date reference appropriate. | e intervals will | be applied to all B | liochemistry reports as |
| Low Vaginal Sv | wab | | | |
| Laboratory: | Microbiology (Main | laboratory) | | |
| Specimen: | suitable transport n | nedium will be p nay be dry and Is ASAP. If proc | processed – swabs may not yield the essing is delayed, | . Only swabs sent in s that are sent without targeted organisms. refrigeration is |
| | | וכ מנ מדווחכות יב | emperature. | |
| Comment | | | | by urgent request |
| Comment: Turnaround: | | tinely Monday t | o Friday 9-5pm or | by urgent request. |

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Lupus Anticoagulant Screen (ACAB IgG /IgM/B2GP1) Antiphospholipid screen

| | Julant Screen (ACAB 19G / 19M/B2GP1) Antiphospholipid screen |
|-------------------------------|--|
| Laboratory: | Haematology |
| Specimen: | Blood $3mL \times 2$, blue Vacuette® (sodium citrate 3.2%) and $1 \times 4mL$ red top Vacuette (clotted). |
| | (Specimens which are haemolysed, underfilled or overfilled cannot be |
| | analysed, check coagulation sample bottles are not expired to ensure correct filling). |
| | Samples must be received within 4 hours. |
| | Note: BCSH guidelines on thrombophilia testing must be adhered to. |
| Comment: | Test available Monday to Friday, during routine working hours. Lupus anticoagulants are immunoglobulins that interfere with phospholipid- dependent coagulation tests. The screen comprises the following tests: PT, APTT, Fibrinogen assay, AFSL, and DVVT. Anti-Cardiolipin antibodies and B2 glycoprotein 1 are also included as part of the screen if a clotted sample is received. |
| | Samples without Request Form WILL NOT be processed. |
| | Thrombophilia request form FOR-CUH-PAT-1575 includes documentation of |
| | patient consent must be received with all requests and is available on the CUH |
| | website. |
| Turnaround: | 3 – 4 weeks (Refer to the main Haematology Section on Coagulation). |
| Report: | Reported Positive or Negative |
| Lyme Serology | / Borrelia burgdorferi Antibodies |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood, CSF (1mL) |
| Comment: | CSF only tested where antibody confirmed in blood. |
| | If clinically suspicious, the test should be repeated after a month as |
| | antibodies take some time to develop. |
| | Serum samples testing positive in house and CSF specimens are sent to a reference laboratory (Rare and Imported Pathogens Laboratory (RIPL), |
| | Porton Down). |
| Turnaround: | Negative serum samples: 36 hours |
| | Serum samples positive in house and CSF: 28 working days |
| Report: | Qualitative result |
| | oma venereum LGV |
| Laboratory: | Microbiology |
| Specimen: | Male Rectal swab. Appropriate PCR STD Specimen Collection and Transport Kits must be used. Please read the kit insert for information on specimen collection and associated limitations. |
| Comment: | Performed by a reference laboratory (Molecular Microbiology, Central Pathology Laboratory, St James Hospital. Dublin 8). |
| | This test is only performed on male rectal specimens that have tested positive for Chlamydia tracomatis and where the patient has the following clinical details: • HIV positive |
| | A contact of a known LGV confirmed case |
| Turnaraundu | Symptomatic of LGV |
| Turnaround: | 14 days Detected or not detected |
| Ref. Range: | |
| Lysosomal Enzy Laboratory: | |
| Laboratory. | Referred from Biochemistry to Willink Biochemical Genetics Unit, St Mary's |
| Specimen: | Hospital, Manchester |
| Specimen. | 5 ml EDTA whole blood |

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| | | Addion | | |
| Comment: | Express post (M,T,V | V) Delivery <72 | hrs | |
| Turnaround: | 8 weeks | | | |
| Ref. Range: | See report form | | | |
| 12 (Pyruvate I | Dehydrogenase Elis | a Test) | | |
| Laboratory: | Autoimmune Serolo | gy | | |
| Specimen: | Blood, 4 mL red top | Vacuette (or s | imilar container fo | r clotted blood) |
| Comment: | Quantitative Elisa. U Mitochondrial Immu | | | era showing specific An Screen. |
| Turnaround: | 96 Hours | | | |
| Ref. Range: | 0 - 5 IU/ML | | | |
| lagnesium (B | lood) | | | |
| Laboratory: | Clinical Biochemistr | • | | |
| Specimen: | 4.0 mL blood in plai | | sample) | |
| Comment: | Haemolysis invalida | | | |
| Turnaround: | MGH: - 3 hours 153 | egain153. GP | or OPD- Results p | CUMH, SI, SF, SMOH, osted within 4 days. |
| Ref. Range: | Up-to-date referenc appropriate. | e intervals will | be applied to all B | iochemistry reports as |
| lagnesium (U | | | | |
| Laboratory: | Clinical Biochemistr | У | | |
| Specimen: | 24 Hr collection | | | |
| Turnaround: | 1 Day | | | |
| Ref. Range: | 3.0 – 5.0mmol/24 H | | | |
| Comment | Up-to-date referenc appropriate. | e intervals will | be applied to all B | iochemistry reports as |
| lalaria PCR, A | ntigen and Blood Fi | Im Screen | | |
| Laboratory: | Haematology | | | |
| Specimen: | Blood 3mL purple V | • | • | |
| Comment: | request. An immun Plasmodium falcipal species of malaria, ovale, and plasmod confirm presence of ovale, P. falciparum percentage of infest | at all other tim odiagnostic tes rum antigens an Plasmodium fal- ium malariae in same, to ident same, to ident c. P. vivax and P cation of Plasmo y may produce ning test is not | t is used for the dent is used for the dent an antigen that ciparum, Plasmodi whole blood. Block ify other forms of <i>c. knowlesi</i> , also to be dium falciparum of a negative result of | aboratory when sending etection of circulating is common to four <i>ium vivax, Plasmodium</i> od films are examined t Malaria. <i>P. malariae, P.</i> o estimate the or <i>P. knowlesi</i> if presen on the antigen screenin |
| | of Malaria present a Note: Where a mal- a positive screen re and %parasitaemia Positive samples are Laboratory, Faculty | nd also to estir aria sample is > quires a fresh s .(as per BCSH (e referred from of Infectious & Medicine, Kepp | nate the percentage 4 hrs old when re ample <4hrs old t Guidelines). Haematology to the Tropical Diseases, el Street, LONDON | ceived in the laborator to confirm the species he Malaria Reference , London School of I, WC1E 7HT. Please |

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| | | | | |
| Turnaround: | A verbal report is a | lways given on | day of sample red | ceipt. |
| | Emergency specime | | | |
| | Routine specimens | | | |
| | Positive samples re | | ed above: 28 day | s (phoned report |
| | available within 3 v | | | |
| Result: | - | • | • | arum or P. knowlesi). |
| | Referral report: Se | nt to referring c | linician and copy | filed in laboratory |
| Manganese | | | | |
| Laboratory: | Referred from Clinical | Biochemistry to | o SAS Laboratory | for Trace Elements, |
| C | Guildford | a tuba (navu tan) | | |
| Specimen: | Sod Hep trace metal fre | | | n a concernent de collecter |
| Comment: | | | | s necessary to collect a wn for other analyses a |
| | the same time, other | - | | |
| | Alternatively, a plastic | | | |
| | | | | |
| Turnaround: | 10 days from receipt | in Referral labor | atory | |
| Ref. Range: | Up-to-date reference | | | chemistry reports as |
| | appropriate | | | <i>,</i> , |
| Maturity Onse | et Diabetes of the Yo | oung (MODY) | | |
| Laboratory: | Referred from Mole | cular Genetics I | ab in Biochemisti | ry to Royal Devon & |
| | Exeter NHS(via NC | MG) | | |
| Specimen: | 3-5ml EDTA blood | | | |
| Comment: | Special request for | m available fron | n | |
| | http://www.diabetesgen | es.org/sites/default/ | files/mody request fo | orm april 2013 0.doc |
| | Please note: invoic | | • • | · · · · · · · · · · · · · · · · · · · |
| Turnaround: | 8 weeks | | | Jerney enneren |
| Report: | Sent to referring cl | inician and copy | filed in pathology | v |
| | r Liposarcomas | | | 1 |
| Laboratory: | Referred by Patholo | oav to HSL Adva | anced Diagnostics | . London |
| Specimen: | - | | - | IO slides by Pathology |
| Turnaround: | 14 working days (f | · · | • | , |
| Measles IgG A | | | | |
| Laboratory: | Microbiology (Infec | tious Diseases S | Serology) | |
| Specimen: | 4mL clotted blood | | 577 | |
| Turnaround: | 36 hours | | | |
| Report: | Qualitative result | | | |
| Measles IgM | | | | |
| Laboratory: | Microbiology (Infec | tious Diseases S | Serology) | |
| Specimen: | 4mL clotted blood, | | 2,,, | |
| Comment: | | | ry (National Virus | Reference Laboratory |
| | (NVRL), Dublin) | | | , |
| Turnaround: | 14 working days | | | |
| Report: | Qualitative result | | | |
| Measles Mole | cular | | | |
| Laboratory: | Microbiology (Infec | tious Diseases S | Serology) | |
| , Specimen: | 4mL clotted blood, | | 2., | |
| Comment: | | | ry (National Virus | Reference Laboratory |
| | (NVRL), Dublin) | | - | , |
| Turnaround: | 14 working days | | | |
| Report: | Detected or not det | tected | | |
| • | | | | |

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| Meningitis C Va | accine Antibodies |
|--------------------------------------|---|
| Laboratory: | Clinical Biochemistry |
| Specimen: | Blood 4mL red top Vacuette [®] (or similar container for clotted blood) |
| Comment: | Performed by a reference laboratory (Irish Meningococcal and Meningitis |
| | Reference Laboratory, The Children's Hospital, Temple Street, Dublin). |
| Turnaround: | 8-10 weeks |
| Report: | Positive or negative |
| Meningococcal | PCR |
| See Neisseria | meningitidis PCR |
| | een / Blood (Amino Acid Chromatography) |
| Laboratory: | Sample referred from Clinical Biochemistry to The Children's Hospital, Temple Street, Dublin |
| Specimen: | Lithium Heparin sample which must be separated within 30 minutes of collection |
| Turnaround: | 4 weeks |
| Ref. Range: | See report or contact Biochemistry Laboratory Temple Street Hospital. |
| Metabolic Scre | |
| Laboratory: | Sample referred from Clinical Biochemistry to The Children's Hospital, Temple Street, Dublin |
| Specimen: | Spot urine, transport to Bio lab immediately for the addition of 5% Merthiolate |
| Comment: | Sample assayed for Creatinine, Protein, Ph, reducing substances, blood, glucose, ketones, mucopolysaccharides, sulphur amino acids, amino acid chromatography, ketoacids (DNPH) |
| Turnaround: | 1 week |
| Ref. Range: | See report or contact Biochemistry Laboratory, Temple Street Hospital. |
| Metanephrines | |
| Laboratory: | Sample referred from Clinical Biochemistry to Biochemistry Department, Freeman Hospital, Newcastle |
| Specimen: | 2 EDTA blood samples (5-7 mLs) taken 10 minutes apart. Send to laboratory on ice. |
| Comment: | Consultant request only |
| Turnaround: | 5 weeks |
| Metanephrines | (Urinary) |
| Laboratory: | Sample referred from Clinical Biochemistry to Beaumont Hospital |
| Specimen: | 24-hour urine sample collected into a container that has acid added. 24 hr urine containers are available from stores; acid is added in the Biochemistry lab. |
| Turnaround: | 5 weeks |
| Ref. Range: | See report form |
| Methadone | |
| Laboratory: Specimen: Comment: | Sample referred from Clinical Biochemistry to Toxicology Laboratory BEAUMONT Hospital Dublin, posted Monday, Tuesday, Wednesday and Thursday. Spot urine |
| | See Toxicology / Drug Screen |
| Turnaround: Ref. Range: | 1 week See report form or contact Toxicology Laboratory BEAUMONT Hospital 01- 8092673 / (01) 8092675, Emergency after hours (087) 2590749, Fax (01) 8093986 |

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| Specimen:LithiTurnaround:1 hoRef. Range:< 1.Methicillin-ResistantLaboratory:MicroSpecimen:SwasteriIf prtempComment:TestLabeprevcultuaxillanareThea moan ocomguidFor efor nanyTurnaround:Report:MRSMethotrexate (HighLaboratory:CliniSpecimen:4.0 mfor nComment:MeasBiocdose | t Staph aureus (MRSA) obiology (Main laboratory) bs should be placed in charcoal containing transport media. Use a clean, le, leakproof container for CSU and sputum. Transport specimens ASAP. occessing is delayed, refrigeration is preferable to storage at ambient berature. performed Monday to Friday (cut-off is 1pm). el all Microbiology forms with MRSA SCREEN. Indicate if the patient was iously MRSA positive. In screening investigations, patient surveillance ures usually include one swab from both nares, one swab from both ae and one swab from both sides of groin (3 swabs in all). Swabs from s, axillae and umbilicus are sufficient for infants and neonates. anterior nares are the usual site cultured from hospital staff. Occasionally ore extensive screening of staff who are carriers is required e.g. during utbreak. When MRSA is detected in any microbiological specimen, on pletion of treatment rescreen as recommended by national and local elines. electronic orders through the iCM system, one request should be entered aares, one for axilla and groin (one number, print two labels), and one for other site that is to be tested. m: 24 hours; Final: 24-48 hours A not isolated or MRSA isolated. Appropriate sensitivities on new isolates. |
|---|--|
| Turnaround:1 hoRef. Range:< 1. | ur 15 mins 5% Staph aureus (MRSA) obiology (Main laboratory) bs should be placed in charcoal containing transport media. Use a clean, le, leakproof container for CSU and sputum. Transport specimens ASAP. occessing is delayed, refrigeration is preferable to storage at ambient berature. performed Monday to Friday (cut-off is 1pm). el all Microbiology forms with MRSA SCREEN. Indicate if the patient was iously MRSA positive. In screening investigations, patient surveillance ures usually include one swab from both nares, one swab from both ae and one swab from both sides of groin (3 swabs in all). Swabs from s, axillae and umbilicus are sufficient for infants and neonates. anterior nares are the usual site cultured from hospital staff. Occasionally pre extensive screening of staff who are carriers is required e.g. during utbreak. When MRSA is detected in any microbiological specimen, on pletion of treatment rescreen as recommended by national and local elines. electronic orders through the iCM system, one request should be entered hares, one for axilla and groin (one number, print two labels), and one for other site that is to be tested. m: 24 hours; Final: 24-48 hours A not isolated or MRSA isolated. Appropriate sensitivities on new isolates. |
| Ref. Range:< 1.Methicillin-ResistantLaboratory:MicroSpecimen:SwasteriIf prtempComment:TestLabeprevComment:TestLabeprevcultuaxillanareThea moan ocomguidFor efor nanyTurnaround:PreliReport:Methotrexate (HighLaboratory:CliniSpecimen:4.0 nfor mfor nSpecimen:4.0 nfor mfor mSpecimen:4.0 nfor mfor mComment:MeasBiocdose | 5% t Staph aureus (MRSA) obiology (Main laboratory) bs should be placed in charcoal containing transport media. Use a clean, le, leakproof container for CSU and sputum. Transport specimens ASAP. ocessing is delayed, refrigeration is preferable to storage at ambient berature. performed Monday to Friday (cut-off is 1pm). el all Microbiology forms with MRSA SCREEN. Indicate if the patient was iously MRSA positive. In screening investigations, patient surveillance ures usually include one swab from both nares, one swab from both ae and one swab from both sides of groin (3 swabs in all). Swabs from s, axillae and umbilicus are sufficient for infants and neonates. anterior nares are the usual site cultured from hospital staff. Occasionally pre extensive screening of staff who are carriers is required e.g. during utbreak. When MRSA is detected in any microbiological specimen, on pletion of treatment rescreen as recommended by national and local elines. electronic orders through the iCM system, one request should be entered ares, one for axilla and groin (one number, print two labels), and one for other site that is to be tested. m: 24 hours; Final: 24-48 hours A not isolated or MRSA isolated. Appropriate sensitivities on new isolates. |
| Methicillin-Resistant Laboratory: Micro Specimen: Swa steri If pr temp Comment: Test Labe prev cultu axilla nare The a mo an o com guid For e for n any Turnaround: Preli Report: MRS Methotrexate (High Laboratory: Clini Specimen: 4.0 n for n Comment: Meas Bioc dose | t Staph aureus (MRSA) obiology (Main laboratory) bs should be placed in charcoal containing transport media. Use a clean, le, leakproof container for CSU and sputum. Transport specimens ASAP. occessing is delayed, refrigeration is preferable to storage at ambient berature. performed Monday to Friday (cut-off is 1pm). el all Microbiology forms with MRSA SCREEN. Indicate if the patient was iously MRSA positive. In screening investigations, patient surveillance ures usually include one swab from both nares, one swab from both ae and one swab from both sides of groin (3 swabs in all). Swabs from s, axillae and umbilicus are sufficient for infants and neonates. anterior nares are the usual site cultured from hospital staff. Occasionally pre extensive screening of staff who are carriers is required e.g. during utbreak. When MRSA is detected in any microbiological specimen, on pletion of treatment rescreen as recommended by national and local elines. electronic orders through the iCM system, one request should be entered nares, one for axilla and groin (one number, print two labels), and one for other site that is to be tested. m: 24 hours; Final: 24-48 hours A not isolated or MRSA isolated. Appropriate sensitivities on new isolates. |
| Laboratory: Micro Specimen: Swa steri If pr temp Comment: Test Labe prev cultu axilla nare The a mo an o com guid For e for n any Turnaround: Preli Report: MRS Methotrexate (High Laboratory: Clini Specimen: 4.0 n for n Comment: Meas | biology (Main laboratory) bs should be placed in charcoal containing transport media. Use a clean, le, leakproof container for CSU and sputum. Transport specimens ASAP. ocessing is delayed, refrigeration is preferable to storage at ambient berature. performed Monday to Friday (cut-off is 1pm). el all Microbiology forms with MRSA SCREEN. Indicate if the patient was iously MRSA positive. In screening investigations, patient surveillance ures usually include one swab from both nares, one swab from both ae and one swab from both sides of groin (3 swabs in all). Swabs from s, axillae and umbilicus are sufficient for infants and neonates. anterior nares are the usual site cultured from hospital staff. Occasionally pre extensive screening of staff who are carriers is required e.g. during utbreak. When MRSA is detected in any microbiological specimen, on pletion of treatment rescreen as recommended by national and local elines. electronic orders through the iCM system, one request should be entered mares, one for axilla and groin (one number, print two labels), and one for other site that is to be tested. m: 24 hours; Final: 24-48 hours A not isolated or MRSA isolated. Appropriate sensitivities on new isolates. |
| Specimen: Swa steri If pr temp Comment: Test Labe prev cultu axilla nare The a mo a no com guid For e for n any Turnaround: Preli Report: MRS Methotrexate (High Laboratory: Clini Specimen: 4.0 n for n Comment: Meas Bioc dose | bs should be placed in charcoal containing transport media. Use a clean, le, leakproof container for CSU and sputum. Transport specimens ASAP. ocessing is delayed, refrigeration is preferable to storage at ambient berature. performed Monday to Friday (cut-off is 1pm). el all Microbiology forms with MRSA SCREEN. Indicate if the patient was iously MRSA positive. In screening investigations, patient surveillance ures usually include one swab from both nares, one swab from both ae and one swab from both sides of groin (3 swabs in all). Swabs from s, axillae and umbilicus are sufficient for infants and neonates. anterior nares are the usual site cultured from hospital staff. Occasionally ore extensive screening of staff who are carriers is required e.g. during utbreak. When MRSA is detected in any microbiological specimen, on pletion of treatment rescreen as recommended by national and local elines. electronic orders through the iCM system, one request should be entered hares, one for axilla and groin (one number, print two labels), and one for other site that is to be tested. m: 24 hours; Final: 24-48 hours A not isolated or MRSA isolated. Appropriate sensitivities on new isolates. |
| Comment: Steri If pr temp Comment: Test Labe prev cultu axilla nare The a mo an o com guid For e for n any Turnaround: Preli Report: MRS Methotrexate (High Laboratory: Clini Specimen: 4.0 n for n Comment: Meas Bioc dose | le, leakproof container for CSU and sputum. Transport specimens ASAP. occessing is delayed, refrigeration is preferable to storage at ambient performed Monday to Friday (cut-off is 1pm). el all Microbiology forms with MRSA SCREEN. Indicate if the patient was iously MRSA positive. In screening investigations, patient surveillance ures usually include one swab from both nares, one swab from both ae and one swab from both sides of groin (3 swabs in all). Swabs from s, axillae and umbilicus are sufficient for infants and neonates. anterior nares are the usual site cultured from hospital staff. Occasionally ore extensive screening of staff who are carriers is required e.g. during utbreak. When MRSA is detected in any microbiological specimen, on pletion of treatment rescreen as recommended by national and local elines. electronic orders through the iCM system, one request should be entered hares, one for axilla and groin (one number, print two labels), and one for other site that is to be tested. m: 24 hours; Final: 24-48 hours A not isolated or MRSA isolated. Appropriate sensitivities on new isolates. |
| Laber prev cultu axilla nare The a mo a no com guid For e for n any Turnaround: Preli Report: MRS Methotrexate (High Laboratory: Clini Specimen: 4.0 n for n Comment: Meas Bioc dose | el all Microbiology forms with MRSA SCREEN. Indicate if the patient was iously MRSA positive. In screening investigations, patient surveillance ures usually include one swab from both nares, one swab from both ae and one swab from both sides of groin (3 swabs in all). Swabs from s, axillae and umbilicus are sufficient for infants and neonates. anterior nares are the usual site cultured from hospital staff. Occasionally ore extensive screening of staff who are carriers is required e.g. during utbreak. When MRSA is detected in any microbiological specimen, on pletion of treatment rescreen as recommended by national and local elines. electronic orders through the iCM system, one request should be entered ares, one for axilla and groin (one number, print two labels), and one for other site that is to be tested. m: 24 hours; Final: 24-48 hours A not isolated or MRSA isolated. Appropriate sensitivities on new isolates. |
| Labe prev cultu axilla nare The a mo a no com guid For e for n any Turnaround: Preli Report: MRS <u>Methotrexate (High</u> Laboratory: Clini Specimen: 4.0 n for n Comment: Meas Bioc dose | el all Microbiology forms with MRSA SCREEN. Indicate if the patient was iously MRSA positive. In screening investigations, patient surveillance ures usually include one swab from both nares, one swab from both ae and one swab from both sides of groin (3 swabs in all). Swabs from s, axillae and umbilicus are sufficient for infants and neonates. anterior nares are the usual site cultured from hospital staff. Occasionally ore extensive screening of staff who are carriers is required e.g. during utbreak. When MRSA is detected in any microbiological specimen, on pletion of treatment rescreen as recommended by national and local elines. electronic orders through the iCM system, one request should be entered ares, one for axilla and groin (one number, print two labels), and one for other site that is to be tested. m: 24 hours; Final: 24-48 hours A not isolated or MRSA isolated. Appropriate sensitivities on new isolates. |
| Turnaround: Preli Report: MRS Methotrexate (High Laboratory: Clini Specimen: 4.0 r for n Comment: Meas Bioc dose | m: 24 hours; Final: 24-48 hours A not isolated or MRSA isolated. Appropriate sensitivities on new isolates. |
| Report:MRSMethotrexate (HighLaboratory:CliniSpecimen:4.0 mfor mComment:MeasBiocdose | A not isolated or MRSA isolated. Appropriate sensitivities on new isolates. |
| Methotrexate (High Laboratory: Clini Specimen: 4.0 r for n Comment: Meas Bioc dose | |
| Laboratory: Clini Specimen: 4.0 r for n Comment: Meas Bioc dose | Dose) |
| Specimen: 4.0 r for n Comment: Meas Bioc dose | |
| for n Comment: Meas Bioc dose | cal Biochemistry |
| Bioc dose | mL blood in plain tube (Gel free clotted sample) Serum samples tested nethotrexate should be protected from light |
| Turnaround: Sam | sured in CUH only on patients with high-dose Methotrexate. Contact hemistry laboratory in advance – it is desirable to check the 48hr post level on Wednesdays. |
| | e day |
| 24hr | high dose Methotrexate levels are measured at 48hr, 72hr and every s until level is $<0.05 \ \mu$ mol/L to guide Calcium Folinate (Leucovorin) ue therapy. |
| Microarray (Array C | GH) Analysis |
| Laboratory: Refe | rred from Biochemistry to NCMG |
| | cimen: Adults: 5ml EDTA blood nts: 2ml min EDTA blood |
| Comment: NCM Pleas | G request form available on <u>www.genetics.ie</u> se note: invoices will be issued to the referring clinician for tests not |
| - | prmed in NCMG. |
| Turnaround: 6-10 Report: Sent | |

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| Microsatellite | | | | | |
|----------------|---|--|--|--|--|
| Laboratory: | Specimen referred from Histopathology to Department of Histopathology, Beaumont, D9 | | | | |
| Specimen: | Tissue block | | | | |
| Turnaround: | Turnaround: 20 days | | | | |
| Mineral Analy | Mineral Analysis (copper/iron) | | | | |
| Laboratory: | Histopathology | | | | |
| Specimen: | Liver biopsy unfixed | | | | |
| Comment: | Place specimen on filter paper in dry universal container | | | | |
| Turnaround: | 4-6 weeks (specimen is referred to external laboratory) | | | | |
| Mitochondrial | Antibodies (Immunofluorescence Test) | | | | |
| Laboratory: | Autoimmune Serology | | | | |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) | | | | |
| Comment: | Immunofluorescence assay. Part of Autoantibody Screen. Quantitative Anti- M2 assay automatically undertaken on all immunofluorescence positive sera. | | | | |
| Turnaround: | 24 Hours | | | | |
| Ref. Range: | Contact Laboratory | | | | |
| Mitochondrial | | | | | |
| Laboratory: | Referred from Molecular Genetics lab in Biochemistry to Newcastle Mitochondrial NCG via NCMG | | | | |
| Specimen: | 3-5ml EDTA blood | | | | |
| Comment: | Special request form available at | | | | |
| | http://www.mitochondrialncg.nhs.uk/documents/NCG_Referral_Form.pdf | | | | |
| | Please note: invoices will be issued directly to the referring clinician. | | | | |
| Turnaround: | 8-10 weeks | | | | |
| Report: | Sent to referring clinician and copy filed in pathology | | | | |
| Mitotane | | | | | |
| Laboratory: | Referred from Biochemistry to Cardiff Toxicology Laboratory, Cardiff and Vale University Health board | | | | |
| Specimen: | EDTA sample | | | | |
| Comment: | Trough sample >12hr post dose | | | | |
| Turnaround: | 4 weeks | | | | |
| Report: | | | | | |
| Molecular gen | etics for the diagnosis of AML, CML and ALL | | | | |
| Laboratory: | Referred from Haematology to Munich Leukaemia Laboratory (MLL MVZ GmbH), Germany | | | | |
| Specimen: | 10-15 ml bone marrow or 10-15 ml bone marrow aspirate/peripheral | | | | |
| | Blood (EDTA or heparin) | | | | |
| Comment: | Must arrange with Haematology, transport within 24 hours, complete form from referral laboratory | | | | |
| Turnaround: | 2-10 working days | | | | |
| Report: | Sent to referring clinician and copy filed in laboratory | | | | |
| Monkey pox R | NA | | | | |
| Laboratory: | Microbiology (Main laboratory) | | | | |
| Specimen: | Viral swab from vesicle/lesions | | | | |
| Comment: | Microbiology Medical/Infectious Disease Medical/Public Health team must be contacted prior to taking samples | | | | |
| _ | Site must be specified as Genital Herpes is a notifiable disease, also report includes HSV1, HSV2, VZV (part of differential diagnosis) | | | | |
| Turnaround: | 5 working days | | | | |
| Report: | Detected or Not detected | | | | |

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| Mouth Swab | |
|----------------------|--|
| Laboratory: | Microbiology (Main laboratory) |
| Specimen: | Specimen pus if present otherwise swab any lesions or inflamed areas. A tongue depressor or spatula may be helpful to aid vision and avoid contamination from other parts of the mouth. Swabs should be transported as soon as possible in charcoal containing transport media. If processing is |
| Comment: | delayed, refrigeration is preferable to storage at ambient temperature. Test performed routinely Monday to Friday 9-5pm or by urgent request. For possible herpes infection consider a Viral Culture. A separate swab in appropriate viral transport media is necessary. |
| Turnaround: | Microscopy for Vincent's angina: 24 hours Culture Final: 24-48 hours |
| Report: | Presence or absence of Vincent's organisms. Culture: Any clinically significant isolate with the appropriate sensitivities. |
| MSU – Midstre | am Urine |
| See Urine Mi | croscopy and Culture or Cytology |
| MTHFR (Methy | lenetetrahydrofolate Reductase) C667T Mutation |
| Laboratory: | Sample referred from Haematology to Eurofins-Biomnis |
| Specimen: | 3.0 mL blood EDTA |
| Comment: | When the body is deficient in methylenetertahydrofolate reductase its ability to absorb folate is inhibited. Folic acid is essential for red cell production and for the development and health of the foetus and deficiency may lead to hyperhomocystenemia and preeclampsia. |
| | A combined request/consent form as part of the new EU GDPR rules is required to be completed and is available on the Eurofins website |
| Turnaround: | 32 days |
| Result: | Sent to referring clinician and copy filed in laboratory |
| Mumps IgG An | tibody |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Turnaround: | 36 hours |
| Report: | Qualitative result |
| Mumps IgM An | Itibody |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood, oral fluid |
| Comment: | Performed by a reference laboratory (National Virus Reference Laboratory (NVRL), Dublin) |
| Turnaround: | 14 working days |
| Report: | Qualitative result |
| Mumps Molecu | llar |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | Oral fluid, throat swab, CSF, urine |
| Comment: | Performed by a reference laboratory (National Virus Reference Laboratory (NVRL), Dublin) |
| Turnaround: | 14 working days |
| Report: | Detected or not detected |
| Muscle Biopsy | |
| Laboratory: | Neuropathology |
| Specimen: | Fresh Muscle (universal precautions) |
| | |

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| Comment: | The muscle biopsy must be at least 1.5cm x 1.5cm x 1.5cm in size. For |
|----------|---|
| | certain suspected metabolic or mitochondrial disorders, a larger sample may |
| | be required for molecular or biochemical analysis. Please contact the |
| | Neuropathologist to discuss the case in advance. |

The biopsy should be sent immediately FRESH to the Neuropathology Department. Universal safety precautions for fresh tissue should apply. For specimens which have to be sent over a distance (e.g. Mercy, Bantry, Mallow, Limerick etc.) the biopsy can be wrapped in clingfilm to avoid drying out during transport. Telephone 021 4922519 to let us know that the biopsy is en route. The biopsy should be delivered directly to a staff member in the Neuropathology Dept. Please pack sample according to Packing Instruction 650. Taxi driver/courier should be instructed not to leave specimen at laboratory reception and also instructed in how to deal with spillages. The muscle biopsy should reach the department by 4.00pm. On receipt of the specimen a staff member will telephone the referring hospital laboratory to confirm that the tissue has arrived safely.

Muscle histochemistry is performed in batches once weekly, on Wednesdays. The biopsy can be taken on any day and sent to arrive in the Neuropathology Department no later than 4.00pm.

Additional information is available in the protocol for muscle biopsy (available from the Neuropathology Dept.).

| Muscle Mitochondrial Enzyme and Genetic Analysis | | | | | |
|---|---|--|--|--|--|
| Laboratory: | Neuropathology | | | | |
| Specimen: | Frozen Muscle | | | | |
| Comment: | Please refer to muscle biopsy protocol above. Specimens sent to Newcastle Mitochondrial NCG Diagnostic Service, Newcastle Upon Tyne, UK. | | | | |
| Turnaround: | 4-12 weeks but may be up to 6 months depending on case complexity. | | | | |
| Mutation analysis for inherited bleeding disorders, Haemophilia carrier testing for | | | | | |
| | direct mutational detection, mutation analysis for inherited Factor VIII or Factor IX | | | | |
| deficiency | | | | | |
| Laboratory: | Referred from Haematology Dept. to Haemostasis Molecular Diagnostics (HMD), National Coagulation Laboratory, Centre for Clinical and Laboratory Medicine, CPLM, St James Hospital, Dublin 8 | | | | |
| Specimen: | Min x 2 EDTA, 6-20 ml | | | | |
| Comment: | Contact Coagulation Medical Team at 01 4162141 | | | | |
| | Counselling and consent required before testing | | | | |
| | Samples must be received in the laboratory within 7 days of phlebotomy | | | | |
| Turnaround: | 120 working days but can vary depending on gene | | | | |
| Report: | Sent to referring clinician and copy filed in laboratory | | | | |

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Mycobacteria Testing

Laboratory: Microbiology (TB Laboratory)

Specimen Types

- Sputum Collect early in the morning on at least 3 consecutive days. Sputum should be expectorated from the lower respiratory tract by deep coughing. Preferably, collect a minimum volume of 5mL per specimen. Saliva and postnasal secretions are not suitable. Specimens collected on 3 consecutive days should not be pooled. This may be important if Mycobacteria other than *Mycobacterium tuberculosis* are isolated as interpretation is based on repeated isolation.
- Bronchial washings Minimum specimen size is preferably 5mL.
- Urine Only processed after prior consultation with Microbiology Medical Team. Collect early morning urine on 3 consecutive days. A minimum volume of 20mL is desirable.
- Gastric lavage fluid Only processed after prior consultation with Microbiology Medical Team. Collect samples only on Monday to Friday. Collect early in the morning (before breakfast) on 3 consecutive days. Preferably, collect a minimum volume of 5mL per specimen. If the samples are not delivered promptly to Microbiology, gastric acid present in sample will render them useless for processing. Deliver samples straight to the Microbiology laboratory by 9.00am.

Gastric lavage samples must be accompanied by a Handwritten Green Microbiology request form. Gastric lavage samples should not be ordered through iCM.

- Blood Culture for Mycobacterial investigation Only processed after prior consultation with Microbiology Medical Team. Please contact the TB laboratory first as specific bottles for TB culture are available from the laboratory on request (ext. 22823), (Mallow General Hospital, Bantry General Hospital and Mercy University Hospital laboratories must contact the Microbiology medical team on ext 22500/20120 to request bottles for sampling). Blood is added directly to the culture bottles (1-5mL of blood or marrow). The culture bottles should be transported immediately to the laboratory; Samples processed Monday to Friday 9-5.
- Bone marrow is added directly to the culture bottles; see procedure for blood above.
- CSF, body fluids, aspirates, pus Collect aseptically as much as possible into a sterile container. Preferably, a volume of 5-10mL of CSF is required.
- Skin / tissue biopsy / post-mortem specimens Collect aseptically into a sterile container without preservative. Select a caseous portion if possible. The majority of organisms will be found in the periphery of a caseous lesion. As large a specimen as possible should be sent. Microscopy is generally not performed on swabs.

| | generally not performed on on abor | | |
|-------------|--|--|--|
| Comment: | Microscopy and culture performed routinely Monday to Friday 9-5pm. If smear results are desired on the same day that the specimen is submitted, the specimen should reach the laboratory before 3pm and the TB laboratory notified. | | |
| | For the initial diagnosis of mycobacterial infection all specimens should be fresh and taken when possible before anti-tuberculosis treatment is started Specimens should be transported as soon as possible. | | |
| | Specimens other than blood should be refrigerated if transport to the laboratory or specimen processing is delayed for more than 1 hour. | | |
| | For body fluids use a sterile, leakproof, disposable plastic container. | | |
| | Swabs should be transported in Amies transport medium with charcoal. Laryngeal swabs are not recommended and only be used when pus or sputum is unobtainable. | | |
| | Isoniazid, rifampicin, ethambutol Pyrazinamide and streptomycin susceptibility testing performed in IMRL, St James' Hospital. | | |
| Turnaround: | Microscopy: 24-72 hours | | |
| | Culture: 6-8 weeks | | |
| | Positive smear and culture results are telephoned to requesting clinician. | | |
| Report: | Microscopy: Acid-Alcohol fast bacilli not seen or seen with enumerator | | |
| | | | |

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| | | culture for mycobacterium negative or mycobacterium species solated with sensitivities where appropriate | |
|--------------------------|---|---|--|
| Mycology – Fu | ngal Microscopy | and Culture (Dermatophytosis – skin, hair, nails) | |
| Laboratory: Specimen: | Microbiology (Mycology section) Scalp specimens are best obtained by scraping with a blunt scalpel. The contents should include hair stubs, the contents of plugged follicles and skin scales. Hair may also be plucked from the scalp with forceps (infected hairs are usually easy to remove in this way). Cut hairs are unsatisfactory as the focus of infection is usually below or near the surface of the scalp. Nail clippings should be taken from any discoloured, dystrophic or brittle parts of the nail. These should be cut as far back as possible from the free edge of the nail and include its full thickness, scrapings can also be taken from beneath the nail to supplement the clipping specimen. Skin specimens should be collected by scraping outwards from the edges of the lesions, with either a blunt scalpel blade or with the edge of a glass microscope slide. The edge of the lesion is where there is likely to be the most fungus. | | |
| Comment: | It is often helpfe with surgical sp improves the ch reduces the like is essential if gr Transport at roc Do not use fixat All specimens sl sealed sterile cc or glass slides in used. The use o Important not causes dermato <i>nigra</i> , which is a on the skin of th | Some general points on specimen collection are given below: t is often helpful to clean the lesions of the skin or scalp (and sometime nail) with surgical spirit or 70% alcohol prior to collection of specimens as this mproves the chances of detecting the fungus by microscopy and also educes the likelihood of contamination of subsequent cultures. Prior cleaning a essential if greasy ointments or powders have been applied to the region. Transport at room temperature. No not use fixatives. All specimens should be collected and transported in a properly labelled, ealed sterile container i.e. universal containers, Mycological Transport Pack or glass slides in the appropriate slide holder. Loose slides should not be used. The use of clear sticky tape (sellotape) is not recommended. mportant note: If you clinically suspect Hendersonula toruloidea which auses dermatophyte-like lesions of the palms, soles and toe-webs or <i>Tinea</i> <i>sigra</i> , which is a rare condition which causes dark pigmented areas, usually on the skin of the palm, and is clinically distinctive from dermatophyte | |
| Test method: | lesions, please inform the laboratory when sending skin samples for ar Keratinised tissues are treated with potassium hydroxide in the laborate detect hyphae of dermatophytes. Many pathogenic fungi will grow slow conventional media but may be recovered more reliably on special fundation media, which require incubation for up to 4 weeks. Some isolates may require referral to the Mycology Referral Laboratory in Bristol for identiand/or susceptibility testing which can take up to an additional 4 week | | |
| Turnaround: | Direct smear: Culture: | 1 week. 1-3 weeks | |
| Report: | Direct smear: | Fungal elements seen or not seen. Typical microscopic appearance indicates fungal infection but does not identify the particular fungal species. Culture of yeast or fungus provides species identification. Positive microscopy is diagnostic for a fungal infection, however a negative microscopy result does not exclude a diagnosis of fungal infection. | |
| | Culture: | Fungus not isolated or organism name isolated | |

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| Mycoplasma ge | enitalium RNA |
|-----------------------|--|
| Laboratory: | Microbiology (Main Lab) |
| Specimen: | Genital swab /Urine collected using Aptima collection device available from the NVRL, |
| Comment: | Performed by a reference laboratory (National Virus Reference Laboratory (NVRL), Dublin). Test |
| Turnaround: | 14 days |
| Report: | Detected or Not Detected |
| Mycoplasma pi | neumoniae IgM |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Comment: | Performed by a reference laboratory (National Virus Reference Laboratory (NVRL), Dublin). Test is validated only for patients less than or equal to 20 years of age. |
| Turnaround: | 14 days |
| Report: | Qualitative result |
| | Acid (Mycophenolate) |
| Laboratory: | Sample referred from Clinical Biochemistry to Harefield Hospital |
| Specimen: | 0.5ml Plasma EDTA, pasma needs to be separated within 6 hours. |
| Comment: | 12 hour trough level |
| Turnaround: | 6 weeks |
| Therapeutic Range: | Interpretation of Mycophenolic Acid is dependent on time interval between sample and last dose, clinical indication for use of the drug, duration of therapy, other drug therapy and method of measurement |
| Myeloperoxida | ase Antibodies |
| Laboratory: | Autoimmune Serology |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) |
| Comment: | Quantitative Elisa |
| Turnaround: | 72 Hours |
| Ref. Range: | 0 - 20 AU/mL |
| Myoglobin | |
| Laboratory: | Sample referred from Clinical Biochemistry to Sheffield Northern General's Protein Reference Unit Diagnostic Service |
| Specimen: | 2 ml serum or 2 ml urine |
| Turnaround: | 3 weeks |
| Ref. Range: | See report form |

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| eisseria gono Laboratory: | Microbiology |
|-------------------------------------|--|
| Specimen: | Nucleic acid amplification method. Appropriate PCR STD Specimen Collection and Transport Kits must be used. Please read the kit insert for information o specimen collection and associated limitations. |
| Comment: | Test available Monday to Friday 9-5pm. Specimens received for Neisseria gonorrhoea PCR will also be tested for <i>Chlamydia trachomatis</i> DNA. |
| | The assay is verified for use with female Endocervical swab specimens, High Vaginal Swab specimens and male/female Urine specimens. The preferred specimen type for <i>N. gonorrhoea</i> testing in female patients is urine due to increased sensitivity and fewer problems during specimen processing. Underfilled or overfilled Urine specimen containers are unsuitable for testing Endocervical/HVS specimen tubes with no swab or with two swabs cannot be tested. |
| | Use only flocked swabs for Endocervical sampling (this is the thinner of the 2 swabs in the sample collection kit). Woven swabs from Endocervical sites are not processed. |
| | Use woven swabs provided for all other sites, other than Endocervical sites. |
| | Specimens that appear bloody or have a dark brown colour are unsuitable for testing (may give false negative results). |
| | The presence of mucous may inhibit PCR and cause false negative test results. Mucous free specimens are required for optimal test performance. Do not use collection devices beyond their expiry date. |
| Turnaround: | 96 – 120 hour |
| Report: | RT: PCR <i>Neisseria gonorrhoea</i> Target Not Detected or Target Detected. A Target Not Detected result does not automatically exclude infection from <i>Neisseria gonorrhoea</i> as the level of DNA present may be lower than the lim of detection of the assay. |
| | The assay is only verified for use with female Endocervical/HVS swab specimens and male/female Urine specimens. Results from other specimen types should be interpreted with caution. |
| | ingitidis PCR |
| | Microbiology (Infectious Diseases Serology) |
| Specimen: | 1mL EDTA blood, CSF (0.5mL) |
| Comment: | Performed by Irish Meningitis & Sepsis Reference Laboratory (IMSRL), Dubli |
| Turnaround: | 10 working days. Samples received by IMSRL before 11am, verbal result between 4pm and 5pm the same day (positive only). |
| Report: | Detected or not detected |

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| Nerve Biopsy | |
|--------------|--|
| Laboratory: | Neuropathology |
| Specimen: | Fresh nerve (universal precautions) |
| Comment: | Please refer to the nerve biopsy protocol (Neuropathology Information for Users). |
| | The biopsy site should be chosen by the primary care physician. In general, the sural nerve is the most frequently biopsied nerve. A fascicular or complete nerve biopsy can be done. In practice approximately two centimetres of the entire nerve including the perineurium is cut. The laboratory should be notified in advance that a nerve biopsy is en route. It should be sent immediately FRESH to the Neuropathology Dept. Universal safety precautions for fresh tissue should apply. |
| | For specimens which have to be sent over a distance (e.g. Bantry, Mallow etc.) the biopsy can be wrapped in gauze lightly moistened with NORMAL SALINE, to keep moist during transport. Telephone ext 021 4922519 to let us know the biopsy is en route. The biopsy should be delivered directly to a staff member in the Neuropathology Dept. Sample should be packed according to Packing Instruction 650. Taxi driver/courier should be instructed not to leave specimen at laboratory reception and also instructed in how to deal with spillages. The nerve biopsy should reach the department by 4.00pm. On receipt of the specimen a staff member will telephone the referring hospital laboratory to confirm that the tissue has arrived safely. Please indicate on the Neuropathology request form the clinician to whom the result should be sent and if a copy is needed for another clinician. The primary care team should fill out the clinical details on the request form before the patient goes to theatre. For any further queries please contact the Neuropathology laboratory (021 4922519) or Dr Bermingham (021 4920475). |
| Turnaround: | 3 weeks. Certain cases may take longer. |
| | a Screen (Catecholamines and Metanephrines) |
| Laboratory: | Sample referred to Beaumont Hospital, Dublin |
| Specimen: | Fresh spot urine (20 mL, if possible). MUST be acidified in lab within 10 minutes of collection. |
| Comment: | Please notify the Biochemistry laboratory in advance. |
| | State what drugs the patient (<16years) is on during collection. |
| Turnaround: | 3 weeks |
| Ref. Range: | Contact CUH Clinical Biochemistry Laboratory |
| Neuromuscula | r genetics (HNPP, CMT, DM, DMD, FA, SCA etc) |
| Laboratory: | Referred from Molecular Genetics lab in Biochemistry to NCMG |
| Specimen: | 3ml EDTA blood |
| Comment: | Contact 22531 for further information |
| | Please note: invoices will be issued to the referring clinician for tests not performed in NCMG. |
| Turnaround: | See website: <u>www.genetics.ie</u> |
| Report: | Sent to referring clinician and copy of report filed in pathology |
| | Biopsies (Routine) |
| Laboratory: | Neuropathology |
| Specimen: | Formalin-fixed tissue |
| Turnaround: | 5 days |
| | Biopsies (High-Risk) |
| Laboratory: | Neuropathology |

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| Specimen: | Formalin-fixed tissu | 10 | | |
| Comment: | | | , investigation of | faturical domentia and |
| Comment: | · · | | | f atypical dementia and |
| | | | | must be used. Contact the |
| | Neuropathologist or | | | a the CID even will a see |
| | | | | n the CJD surveillance |
| Turnaraundu | centre in Beaumont | • | 0///00 | |
| Turnaround: | N/A, case depender on Sequencing (cfT | | | |
| | | | | |
| Laboratory: | | | | TNA Plasma Molecular |
| | testing in the pathol | | | n patients with Lung |
| | consultant histopati cancer. | iologists on pla | sina samples noi | in patients with Lung |
| | The cut-off for recei | at of those cam | plac into the labo | vistory is 15,00 |
| Chasiman | | | • | |
| Specimen: | 2 K2 EDTA Blood tub | bes (must react | i iad within 4 hou | lts) |
| | <u>OR</u> | | | |
| Comment: | at least 1 Roche cfD | | to tolying the com | and at Eve 22512 (22702 |
| comment: | | | | ple at Ext.22513 /22792 |
| | hand directly to th | | | atory immediately and |
| Turnaround: | 5-10 working days | e medical Scie | | |
| Turnarounu. | J-10 WORKING Udys | | | |
| Norovirus – N | orwalk-like viruses | (NLV) /Small | Round Structu | red Viruses (SRSV) |
| Laboratory: | Microbiology (Cated | • • | | |
| , Specimen: | A fresh liquid faece | | • • | s sufficient. |
| Comment: | - | | | le in-house, otherwise |
| connenti | test will be referred | | • | |
| | Microbiology Medica | | • | |
| | Urgent Norovirus te | - | | al from Medical |
| | Microbiology team | 5 | | |
| | | | | |
| | A Target Not Detec | ted result does | not automatically | <pre>/ exclude infection from</pre> |
| | • | - | | sent may be lower than |
| | the limit of detection | n of the assay. | | |
| | | | | |
| Turnaround: | In-house: 5 workin | | | |
| Report: | Target Detected or | Target Not Det | ected for Norovir | us. |
| Nose Swab | | | | |
| Laboratory: | Microbiology (Main | | | |
| Specimen: | • | J , | 5 | n the surface. Transport |
| | • | | . . | edia. If processing is |
| | | | | bient temperature. |
| Comment: | | • | | clinical details (recurrent |
| | boils, infected ecze | | • • | |
| | Aerobic culture – To | | 2 | |
| | | | | ylococcal infection. Test |
| _ | performed routinely | | | urgent request. |
| Turnaround: | Prelim: 24 hours; Final: 48-72 hours Presence of <i>Staphylococcus aureus</i> usually reflects carrier state. | | | |
| Report: | Presence of Staphy | iococcus aureus | s usually reflects | carrier state. |

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| Laboratory: | Haematology |
|--|---|
| Specimen: | Blood 3mL purple Vacuette [®] (EDTA) |
| | Paediatric (1mL purple (EDTA) or 1.3 mL red) |
| | Note: 6ml purple EDTA Vacuette or any other sample type is unsuitable for |
| | NRBCs. |
| | Blood Films are made in the laboratory as required. |
| Comment: | Please refer to section: Full Blood Count including automated WBC |
| | DifferentialBlood Films for Manual White Cell Differentials, Slide Platelets and |
| | Red Cell Morphology (peripheral blood smear) |
| Oestradiol | |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Turnaround: | 4 Days |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as |
| | appropriate. |
| Oncotype DX Te | |
| Laboratory: | Referred from Pathology to Genomic Health Inc., California |
| Specimen: | FFPE tissue block |
| Turnaround: | 20 working days (from date testing material is sent to referral institution) |
| Ophthalmic Bio | psies |
| Laboratory: | Neuropathology |
| Specimen: | Formalin fixed tissue |
| Turnaround: | 5 days |
| Ophthalmic Bio | psies – corneal smears (acanthamoeba) |
| Laboratory: | Neuropathology |
| Specimen: | Corneal scrape – special fixative required, (CytoLyt), available from Neuropathology. |
| Comment: | Please contact Neuropathology Department in advance on 4922520 |
| Turnaround: | 1-2 days |
| Opiates | |
| Laboratory: | Sample referred from Clinical Biochemistry to Toxicology Laboratory BEAUMONT Hospital Dublin, posted Monday, Tuesday, Wednesday and Thursday. |
| Specimen: | Spot urine |
| Comment: | See Toxicology / Drug Screen |
| Turnaround: | 1 week |
| Ref. Range: | See report form or contact Toxicology Laboratory BEAUMONT Hospital 01- 8092673 8092673 / (01)8092675, Emergency after hours (087) 2590749, Fax (01) 8093986 |
| Organic Acids | |
| Laboratory: | Sample referred from Clinical Biochemistry to The Children's Hospital, Temple Street, Dublin |
| Specimen: | Spot Urine |
| • | Sample must be frozen immediately |
| Comment: | |
| Comment: Turnaround: | 8 Weeks |
| Comment: Turnaround: Ref. Range: | 8 weeks See report or contact Biochemistry Laboratory Temple Street Hospital |

Osmolality (Serum)

Laboratory: Clinical Biochemistry

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|---|-----------------------|-----------------------|------------------|-------------------------------|---------------------------|
| Approved By: Dr Vitaliy Mykytiv, Ms Sinead Creagh Mr Paul Cantwell Specimen: 4.0 mL blood in plain tube (clotted sample) Turnaround: 24 Hours Ref. Range: Up-to-date reference intervals will be applied to all Biochemistry reports a appropriate. Osmolality (Urine) Laboratory: Laboratory: Clinical Biochemistry Specimen: Spot urine sample Turnaround: 24 Hours Ref. Range: Dependant on the patient's state of hydration Ovarian Antibodies Dependant on the patient's state of hydration Covarian Antibodies Dependant on the patient's state of hydration Ovarian Antibodies Eaboratory: Sample referred from Autoimmune Serology to Eurofins-Biomnis Laborator Specimen: Blood, 4 mL red top Vacuette (or similar container for clotted blood) Turnaround: Approve Alaboratory: Specimen referred directly from ward (through Stores department) to Haematology, Our Lady's Hospital Crumlin Specimen: Blood 3mL, purple, Vacuette® (EDTA) Specimen must reach referral laboratory within 3 ½, hours of phlebotomy and Bibordo 3mL, purple, Vacuette® (EDTA) Specimen must reach referral laboratory of the esent by taxi at 8.0 am. | Laboratory M | | | 03/11/2023 | Page: 167 of 206 |
| Specimen: 4.0 mL blood in plain tube (clotted sample) Turnaround: 24 Hours Ref. Range: Up-to-date reference intervals will be applied to all Biochemistry reports a appropriate. Osmolality (Urine) Laboratory: Clinical Biochemistry Specimen: Spot urine sample Turnaround: 24 Hours Ref. Range: Dependant on the patient's state of hydration Ovarian Antibodies Laboratory: Sample referred from Autoimmune Serology to Eurofins-Biomnis Laborato Specimen: Blood, 4 mL red top Vacuette (or similar container for clotted blood) Turnaround: Approx. 3 Weeks Ref. Range: See report form, or visit internet site https://www.eurofins.le/biomnis/ for up t date referral test information. Oxidative Burst analysis Laboratory: Specimen referred directly from ward (through Stores department) to Haematology, Our Lady's Hospital Crumlin Specimen: Blood 3mL, purple, Vacuette [®] (EDTA) Specimen must reach referrel laboratory within 3 ½ .hours of phlebotomy and delivery is organised with Stores Department to be sent by taxi at 8.0 am. Sample referred from Haematology to Eurofins-Biomnis Specimen: Blood 3mL, purple, Vacuette [®] (solium citrate 3.2%) x 3 fill to mark on tubes Comment: Requested by | | | | Dr Vitaliy Mykytiv, M | |
| Turnaround: 24 Hours Ref. Range: Up-to-date reference intervals will be applied to all Biochemistry reports a appropriate. Osmolality (Urine) Laboratory: Clinical Biochemistry Specimen: Spot urine sample Turnaround: 24 Hours Ref. Range: Dependant on the patient's state of hydration Ovarian Antibodies Laboratory: Sample referred from Autoimmune Serology to Eurofins-Biomnis Laborato Specimen: Blood, 4 mL red top Vacuette (or similar container for clotted blood) Turnaround: Approx. 3 Weeks Ref. Range: See report form, or visit internet site https://www.eurofins.ie/biomnis/ for up t Laboratory: Specimen referred directly from ward (through Stores department) to Haematology, Our Lady's Hospital Crumlin Specimen: Blood 3mL, purple, Vacuette® (EDTA) Specimen must reach referral laboratory within 3 ½ .hours of phlebotomy and delivery is organised with Stores Department to be sent by taxi at 8.0 am. Sample msut be taken between 07:30 and 08:00 Comment: Requested by Consultant Haematologist Turnaround: 3 weeks Specimen: Songle referred from Haematology to Eurofins-Biomnis Specimen: Songle and component | | | Author: | Mr Paul Cantwell | |
| Turnaround: 24 Hours Ref., Range: Up-to-date reference intervals will be applied to all Biochemistry reports a appropriate. Osmolality (Urine) Laboratory: Laboratory: Clinical Biochemistry Specimen: Spot urine sample Turnaround: 24 Hours Ref., Range: Dependant on the patient's state of hydration Ovarian Antibodies Laboratory: Sample referred from Autoimmune Serology to Eurofins-Biomnis Laborato Specimen: Blood, 4 mL red top Vacuette (or similar container for clotted blood) Turnaround: Approx. 3 Weeks Ref. Range: See report form, or visit internet site https://www.eurofins.ie/biomnis/ for up t date referral test information. Oxidative Burst analysis Laboratory: Laboratory: Specimen referred directly from ward (through Stores department) to Haematology, Our Lady's Hospital Crumlin Specimen Blood 3mL, purple, Vacuette® (EDTA) Specimen must reach referral laboratory within 3 ½ .hours of phlebotomy and delivery is organised with Stores Department to be sent by taxi at 8.0 am. Sample must be taken between 07:30 and 08:00 Comment: Requested by Consultant Haematology to Eurofins-Biomnis Specimen: Blood 3mL; blue Vacuette® (sodium citrate 3. | Specimen: | 4.0 mL blood in plai | n tube (clotted | sample) | |
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| PAI-1 (Plasminogen Activator Inhibitor)Laboratory:Sample referred from Haematology to Eurofins-BiomnisSpecimen:Blood 3mL; blue Vacuette® (sodium citrate 3.2%) x 3 fill to mark on tubesComment:Request must be booked in advance with the Haematology Laboratory CU(PAI-1) is an important component of the coagulation system that down- regulates fibrinolysis in the circulation. Reduced PAI-1 levels may result in increased fibrinolysis and an associated bleeding diathesis. A combined request/consent form as part of the new EU GDPR rules is required to be completed and is available on the Eurofins websiteTurnaround:40 working daysReport:Sent to referring clinician and copy filed in laboratoryParacetamol4.0 mL blood in or plain tube (clotted sample)Comment:Sample 4 – 12 Hours post ingestionTurnaround:1 Hour 15 minsRef. Range:Interpretation of Paracetamol toxicity is highly dependent on time of putation | | | nician and copy | filed in laboratory | |
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| Specimen: Comment:Blood 3mL; blue Vacuette® (sodium citrate 3.2%) x 3 fill to mark on tubes Request must be booked in advance with the Haematology Laboratory CU (PAI-1) is an important component of the coagulation system that down- regulates fibrinolysis in the circulation. Reduced PAI-1 levels may result in increased fibrinolysis and an associated bleeding diathesis. A combined request/consent form as part of the new EU GDPR rules is required to be completed and is available on the Eurofins websiteTurnaround: 40 working days Report:Sent to referring clinician and copy filed in laboratoryParacetamolClinical Biochemistry 4.0 mL blood in or plain tube (clotted sample) Comment:Sample 4 – 12 Hours post ingestion Turnaround: 1 Hour 15 mins Ref. Range:Interpretation of Paracetamol toxicity is highly dependent on time of putat | Laboratory: | Sample referred from | m Haematolog | / to Eurofins-Biom | nis |
| Comment:Request must be booked in advance with the Haematology Laboratory CUI (PAI-1) is an important component of the coagulation system that down- regulates fibrinolysis in the circulation. Reduced PAI-1 levels may result in increased fibrinolysis and an associated bleeding diathesis. A combined request/consent form as part of the new EU GDPR rules is required to be completed and is available on the Eurofins websiteTurnaround:40 working daysReport:Sent to referring clinician and copy filed in laboratoryParacetamolClinical BiochemistrySpecimen:4.0 mL blood in or plain tube (clotted sample) Comment:Comment:Sample 4 – 12 Hours post ingestion Turnaround:Turnaround:1 Hour 15 mins Ref. Range:Ref. Range:Interpretation of Paracetamol toxicity is highly dependent on time of putation | • | - | | | |
| (PAI-1) is an important component of the coagulation system that down-regulates fibrinolysis in the circulation. Reduced PAI-1 levels may result in increased fibrinolysis and an associated bleeding diathesis. A combined request/consent form as part of the new EU GDPR rules is required to be completed and is available on the Eurofins website Turnaround: 40 working days Report: Sent to referring clinician and copy filed in laboratory Paracetamol Laboratory: Clinical Biochemistry Specimen: 4.0 mL blood in or plain tube (clotted sample) Comment: Sample 4 – 12 Hours post ingestion Turnaround: 1 Hour 15 mins Ref. Range: Interpretation of Paracetamol toxicity is highly dependent on time of putation | • | - | • | • | |
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| required to be completed and is available on the Eurofins websiteTurnaround:40 working daysReport:Sent to referring clinician and copy filed in laboratoryParacetamolLaboratory:Clinical BiochemistrySpecimen:4.0 mL blood in or plain tube (clotted sample)Comment:Sample 4 – 12 Hours post ingestionTurnaround:1 Hour 15 minsRef. Range:Interpretation of Paracetamol toxicity is highly dependent on time of putation | | - | | - | |
| Turnaround:40 working days Report:Report:Sent to referring clinician and copy filed in laboratoryParacetamolLaboratory:Clinical Biochemistry Specimen:Specimen:4.0 mL blood in or plain tube (clotted sample) Comment:Comment:Sample 4 – 12 Hours post ingestion 1 Hour 15 mins Ref. Range:Interpretation of Paracetamol toxicity is highly dependent on time of putation | | | | | |
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| Laboratory:Clinical BiochemistrySpecimen:4.0 mL blood in or plain tube (clotted sample)Comment:Sample 4 – 12 Hours post ingestionTurnaround:1 Hour 15 minsRef. Range:Interpretation of Paracetamol toxicity is highly dependent on time of putat | | Sent to referring cli | nician and copy | filed in laboratory | |
| Specimen:4.0 mL blood in or plain tube (clotted sample)Comment:Sample 4 – 12 Hours post ingestionTurnaround:1 Hour 15 minsRef. Range:Interpretation of Paracetamol toxicity is highly dependent on time of putat | | | | | |
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| Ref. Range: Interpretation of Paracetamol toxicity is highly dependent on time of putat | | | | | |
| | | | racotamal taxia | vity in highly donas | dont on time of nut-ti |
| | ker. kange: | - | | ity is highly depen | dent on time of putativ |
| | | | loniografii | | |

Paraneoplastic screen (See anti-neuronal antibodies)

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Parasitology (enteric) – Ova, Cysts and Parasites (OCPs)

| Parasitology (e | enteric) – Ova, Cysts and Parasites (OCPs) |
|-----------------|--|
| Laboratory: | Microbiology (Category 3 Laboratory) |
| Specimen: | Fresh faeces specimen in a sterile leak-proof container. |
| | Do not refrigerate or incubate specimens. |
| | Three examinations spaced 2-3 days apart are recommended for best |
| | recovery of parasites. Unless the patient has severe diarrhoea or dysentery, |
| | no more than one specimen should be examined within a single 24-hour |
| | period, as shedding of cysts and ova tends to be intermittent. |
| | |
| | If <i>Entamoeba histolytica</i> or <i>Giardia lamblia</i> are suspected and the first 3 specimens are negative, ideally 3 additional specimens should be submitted |
| | at weekly intervals. |
| | at weekly intervals. |
| | <i>Note:</i> Fresh specimens are essential for the examination of trophozoites. |
| | Transport specimens ASAP. Protozoan trophozoites will not survive if the |
| | specimen dries out. Cysts will not form once the specimen has been passed. |
| Comment: | Full clinical details are essential. Faeces specimens from patients with chronic |
| | diarrhoea, patients with a history of foreign travel, immunocompromised |
| | patients or FMT (Faecal Microbiota Transplant) patients will be processed. If |
| | in doubt, please contact the medical staff. Please indicate if specific organisms are sought. Specifically indicate on the |
| | request form if Cyclospora or Microsporidia are sought. |
| | Oocysts of Cryptosporidium spp. Can be identified with special staining |
| | techniques; (Cryptosporidium parvum/hominis detected via molecular |
| | techniques in faeces) their presence may indicate active infection or carriage. |
| Turnaround: | 7 working days |
| Report: | OCP not seen or a report on any parasites seen. |
| | The presence of white or red cells is significant and indicates mucosal |
| | inflammation. |
| | Diagnosis of amoebic colitis requires the presence of <i>Entamoeba histolytica</i> trophozoites containing ingested red cells. |
| | Cysts or trophozoites of <i>Giardia intestinalis</i> confirm a diagnosis of giardiasis. |
| | The presence of characteristic ova can identify infection with hookworms and |
| | other roundworms (nematodes) e.g. <i>Enterobius vermicularis</i> in sticky tape |
| | preparations, Ascaris lumbricoides; flat flukes (trematodes) e.g. Fasciola |
| | hepatica, tape worms e.g. Taenia saginata, Taenia solium. Occasionally |
| | complete worms are passed, enabling specific identification of the adult |
| | worm. |
| Parechovirus M | |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | Respiratory secretions, stool, CSF |
| Comment: | Performed by a reference laboratory (National Virus Reference Laboratory |
| | (NVRL), Dublin) |
| Turnaround: | 14 working days |
| Report: | Detected or not detected |
| Parvovirus B19 | |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Turnaround: | 36 hours |
| Report: | Qualitative result |
| • | - |

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| PCP (Pneumoc | ystis jirovecii) | | | | |
|-----------------|--|--|--|--|--|
| Laboratory: | Histopathology (Cytology Department) | | | | |
| Specimen: | Bronchial lavage (neat or in cytolyt) | | | | |
| Comment: | Tests are performed routinely Monday to Friday during routine working hours | | | | |
| Turnaround: | Samples can be processed as urgent with prior communication with | | | | |
| | laboratory. | | | | |
| Ref. Range: | Not applicable | | | | |
| PCP (Pneumoc | ystis jirovecii) | | | | |
| Laboratory: | Microbiology | | | | |
| Specimen: | Sputum or Brochial lavage (BAL) | | | | |
| Comment: | Test performed by National Virus Reference Laboratory (NVRL), Dublin | | | | |
| Turnaround: | 28 working days | | | | |
| Penile swab | | | | | |
| Refer to Genit | al swab | | | | |
| Pericardial Flu | id / Peritoneal Fluid / Pleural Fluid | | | | |
| See Sterile Bo | dy Fluid – Microscopy and Culture | | | | |
| Perinatal: Plac | enta, Products of Conception, Ectopic Pregnancies | | | | |
| Laboratory: | See formalin fixed histopathology speciments. | | | | |
| Peritoneal Dial | ysis Fluid | | | | |
| See Continuo | us Ambulatory Peritoneal Dialysis Fluid | | | | |
| Pernasal Swab | /Pertussis | | | | |
| See Bordetella | a species – Culture | | | | |
| PFA 100 (Plate | let Aggregation Screen) | | | | |
| Laboratory: | Haematology | | | | |
| Specimen: | Blood 3mL; blue Vacuette [®] (sodium citrate 3.2%) x2. Specimens must be | | | | |
| | sent to the Haematology Lab. Within 2 hours of collection. | | | | |
| | Samples must not be sent in the pneumatic tube system. | | | | |
| | Patients on aspirin are unsuitable for this test. | | | | |
| | Specimens that are haemolysed, underfilled or overfilled cannot be analysed, | | | | |
| | check coagulation sample bottles are not expired to ensure correct filling | | | | |
| | Specimens with platelet counts $<150 \times 10^9$ /l are unsuitable for testing. | | | | |
| Comment: | Test available Mon-Fri before 4pm hours by arrangement with the | | | | |
| | Haematology dept. The process of platelet adhesion and aggregation | | | | |
| | following a vascular injury is simulated in vitro, based on change in vacuum /pressure brought about by platelet plug formation. The most common | | | | |
| | causes of platelet dysfunction are related to uremia, von Willebrand disease | | | | |
| | and exposure to agents such as acetyl salicylic acid. | | | | |
| Turnaround: | 8-24 hours | | | | |
| Ref. Range: | Collagen/Epinephrine 82 – 150 secs Collagen/ ADP 62 – 100 secs | | | | |
| | ytoma & Paraganglioma Genetic Screen | | | | |
| Laboratory: | Referred from molecular genetics lab in Biochemistry to LEEDS NHS via NCMG | | | | |
| Specimen: | 3-5ml EDTA blood | | | | |
| Comment: | NCMG request form available at <u>www.genetics.ie</u> | | | | |
| | Please note: invoices will be issued directly to the referring clinician. | | | | |
| | | | | | |
| Turnaround: | 40 days for 8 gene screen | | | | |

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| Dhamanali dina | |
|--------------------------|--|
| Phencyclidine | |
| Laboratory: | Sample referred from Clinical Biochemistry to Toxicology Laboratory BEAUMONT Hospital Dublin, posted Monday, Tuesday, Wednesday and Thursday. |
| Specimen: | Spot urine |
| Comment: | See Toxicology / Drug Screen |
| Turnaround: | 1 week |
| Ref. Range: | See report form or contact Toxicology Laboratory BEAUMONT Hospital 01- 8092673 / (01)8092675, Emergency after hours (087) 2590749, Fax (01) 8093986 |
| Phenobarbiton | e / Phenobarbital |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Comment: | Take trough sample immediately before next dose. When making |
| Turnaround: | comparative measurements, it is advisable that sampling times be consistent 4 Days. Urgents on request |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as |
| Ker. Kange. | |
| Dhanaturina D | appropriate. |
| | ed Cell Antigens |
| Laboratory: | Blood Transfusion Laboratory |
| Specimen: | 1 X 6 mL EDTA Pink Capped Tube |
| Comment: | Phenotypic analysis of patient red cell antigens (e.g. male partners of antenatal patients found to have developed red cell antibodies during pregnancy in the prediction of HDNB) |
| | Complete the Blood Transfusion or Antenatal Serology request forms LF-C-BTR-XMATCH or LF-C-BTR-ANTENAT. This is an INAB accredited test. |
| Turnaround: | 3 Hours |
| | |
| Ref. Range: | Not Applicable |
| Phenytoin | Clinical Piechomistry |
| Laboratory: Specimen: | Clinical Biochemistry 4.0 mL blood in plain tube (clotted sample) |
| Comment: | Take trough sample immediately before next dose. When making |
| | comparative measurements, it is advisable that sampling times be consistent |
| Turnaround: | 1 Day |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate. |
| Phosphate (Blo | ood) |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Comment: | Haemolysis invalidates result |
| Turnaround: | A/E or urgent sample: - 1 hour 30 mins CUH wards, CUMH, SI, SF, SMOH, MGH: - 3 hours 170egain170. GP or OPD- Results posted within 4 days. |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate. |
| Phosphate (Ur | |
| Laboratory: | Clinical Biochemistry |
| , Specimen: | 24 Hour urine collection, to be acidified as soon as possible in laboratory. |
| Turnaround: | 1 Day |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate. |

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| s vermicularis gation Tests |
|--|
| |
| |
| Haematology |
| Six (minimum) Blood 3mL; blue Vacuette® (sodium citrate 3.2%). |
| Samples must not be sent in the pneumatic tube system. |
| Specimens must be sent to the Haematology Lab. within 2 hours of |
| collection. |
| Limitations: Patients on aspirin are unsuitable for this test. |
| Specimens that are haemolysed, underfilled or overfilled cannot be analysed |
| check coagulation sample bottles are not expired to ensure correct filling. |
| Specimens with platelet counts <150x109/l are unsuitable for testing. |
| Test available Mondays only, by prior arrangement with the Haematology |
| dept. The process of platelet adhesion and aggregation following a vascular |
| injury is simulated in vitro, and the platelets aggregates, which form as a result of being exposed to collagen, ristocetin, ADP and adrenaline, are |
| detected by changes in light transmittance. The most common causes of |
| platelet dysfunction are related to uremia, von Willebrand disease and |
| exposure to agents such as acetyl salicylic acid. |
| 8-24 hours, |
| Reported as Normal / Reduced / No Response / Inconclusive |
| dy Investigation |
| Blood Transfusion Laboratory |
| 1 x 4 ml Clotted sample (red cap with yellow ring). |
| Referred to: I.B.T.S., National Blood Centre, James's St., Dublin 8. |
| Complete the Blood Transfusion request forms LF-C-BTR-ANTENAT or LF-C-BTR-XMATCH |
| This is not an INAB accredited test. |
| 3 weeks |
| Not applicable. |
| Antibodies (IgG) |
| Clinical Biochemistry |
| Blood 4mL red top Vacuette [®] (or similar container for clotted blood) |
| Test performed by reference laboratory (HPA Laboratory, Manchester). |
| 2-3 weeks |
| Refer to specific laboratory report |
| PCR |
| Microbiology (Infectious Diseases Serology) |
| 1mL EDTA blood, CSF (0.5mL) |
| Performed by Irish Meningitis & Sepsis Reference Laboratory (IMSRL), Dubli |
| 10 working days. Samples received by IMSRL before 11am, verbal result |
| between 4pm and 5pm the same day (positive only). |
| Detected or not detected |
| nal nocturnal haemoglobinuria |
| Referred by Haematology to Haematology, St James Hospital, Dublin 8 |
| Blood 3mL x 2, purple Vacuette [®] (EDTA). |
| Test available Monday to Wednesday, before 12.00 noon. PNH is |
| characterised by intermittent intravascular haemolysis due to hypersensitivi of RBC'S to the haemolytic action of complement caused by the lack of proteins DAF and MIRL. Diagnosis is possible by using monoclonal antibodie where the abnormal RBC population is identified by agglutination technique. |
| |

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| | | • | | |
| Turnaround: | Positive results pho 60 working days | ned within 24 ł | nours of receipt o | f result, printed reports i |
| Report: | Sent to referring clinician and copy filed in laboratory | | | |
| - | No evidence of PNH | | | , |
| Polio Antibodie | | , | | |
| Laboratory: | Clinical Biochemistr | v | | |
| Specimen: | Blood 4mL red top | | similar container | for clotted blood) |
| Comment: | • | • | | y Infections Laboratory, |
| | Colindale, London). | | | ,, |
| Turnaround: | 4 weeks | | | |
| Report: | Quantitative report | with an interpr | etative comment | |
| Porphyrin Scre | | • | | |
| Laboratory: | | m Clinical Bioc | hemistry to St. Ja | mes Hospital Dublin |
| , Specimen: | Spot urine sample | | EDTA whole bloc | |
| • | Faeces sample | | Lithium Heparin | |
| Comment: | All samples must be | e protected from | | |
| Turnaround: | 3weeks | | 5 | |
| Ref. Range: | See report or conta | ct Biochemistry | Dept. St James' | Hospital |
| Post-Mortems | ł | | • | • |
| | ost-Mortems Section | 3.5 Dept. of Pa | tholoav | |
| Potassium (Blo | | | | |
| Laboratory: | Clinical Biochemistr | v | | |
| Specimen: | 4.0 mL blood in plai | • | sample) | |
| Comment: | Haemolysis invalidates result | | | |
| Turnaround: | • | le: - 1 hour 30 | | s, CUMH, SI, SF, SMOH, 4 days. |
| Ref. Range: | | | • | Biochemistry reports as |
| Potassium (Uri | | | | |
| Laboratory: | Clinical Biochemistr | V | | |
| Specimen: | Spot or 24 Hr samp | | | |
| Turnaround: | 1 Day | | | |
| Ref. Range: | • | e intervals will | be applied to all | Biochemistry reports as |
| _ | appropriate. | | | · · |
| Pouch of Doug | las Fluid | | | |
| See Sterile Bo | dy Fluid – Microscopy | / and Culture | | |
| Prader Willi Sy | ndrome (PWS) | | | |
| Laboratory: | Referred from Bioch | nemistry to Nat | ional Centre for M | 1edical Genetics (NCMG) |
| Specimen: | Infants: 1ml EDTA I | • | | . , |
| | Adults 3-5ml EDTA | blood | | |
| Comment: | Copy of NCMG requ | est form availa | ble on website w | <u>ww.genetics.ie</u> |
| Turnaround: | 6 weeks | | | - |
| Report: | Sent to referring cli | nician and copy | <u>v of report filed in</u> | pathology |
| | | | | |
| Pregnancy Tes | 15 | | | |
| | Haematology | | | |
| Pregnancy Tes | Haematology | en (must be <4 | 18 hrs old, prefer | ably refrigerated), early |

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| Comment: | of human chorionic | gonadotropin (l nts about 10 day | HCG) which the pl s after fertilisation | etecting elevated levels acenta begins to produce on. Test available Monday |
| | other times. | time working ne | | |
| Turnaround: | Emergency specime Routine specimens: | 8 – 24 hours | | |
| Report: | Positive, Negative o | or Inconclusive | | |
| Procalcitonin | | | | |
| Specimen: S | linical Biochemistry erum. Appropriate cli eeded. When monitor proughout the evaluat | ring patients use | | l patient preparation en collection tube type |
| Turnaround: S | ame day | | | |
| | efer to PCT Interpret | ation Guidelines | 5. | |
| Progesterone | | | | |
| Laboratory: Specimen: | | in tube (clotted to expected day | | ence of ovulation draw Confirm correctness of |
| Turnaround: | 4 Days | | | |
| Ref. Range: | Up-to-date reference appropriate. | ce intervals will | be applied to all E | Biochemistry reports as |
| Prolactin | | | | |
| Laboratory: | Clinical Biochemistr | • | | |
| Specimen: | 4.0 mL blood in plai | in tube (clotted | sample) | |
| Turnaround: Ref. Range: | 4 Days Up-to-date referenc appropriate. | ce intervals will | be applied to all E | liochemistry reports as |
| Propoxyphene | | | | |
| Laboratory: | Sample referred fro BEAUMONT Hospita Thursday. | | | 3, , |
| Specimen: | Spot urine | | | |
| Comment: | See Toxicology / D | rug Screen | | |
| Turnaround: Ref. Range: | 8092673 / (01)809 | | | AUMONT Hospital 01- 37) 2590749, Fax (01) |
| Protein (Total) | 8093986 | | | |
| Laboratory: | Clinical Biochemistr | v | | |
| Specimen: | 4.0 mL blood in plai | | sample) | |
| Turnaround: | A/E or urgent samp MGH: - 3 hours 173 | le: - 1 hour 30 Begain173. GP | min. CUH wards, or OPD- Results p | CUMH, SI, SF, SMOH, osted within 4 days. |
| Ref. Range: | Reference ranges. | | | c and Pregnancy-related Siochemistry reports as |
| | appropriate. | | | |
| Protein (Urinar | | | | |
| Laboratory: | Clinical Biochemistr | • | | |
| Specimen: | Spot or 24 Hr samp | le | | |
| Turnaround: | 1 Day | | | |

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| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as |
|-------------|---|
| | appropriate. |

| | appropriate | e. | | | | |
|---------------|---|--|-----------------------|---|--|--|
| Protein C | | | | | | |
| Laboratory: | Haematolo | av | | | | |
| Specimen: | Blood 3mL; blue Vacuette® (sodium citrate 3.2%). | | | | | |
| opeemen | | | - | ed or overfilled cannot be analysed | | |
| | • | | | | | |
| . . | - | - | | expired to ensure correct filling. | | |
| Comment: | | , | , 2 | itine working hours, and for | | |
| | | | | this assay the Protein C present in | | |
| | | | | , this in turn hydrolyses a | | |
| | | | | ured. Decreased levels are | | |
| | | | | in patients with hepatic disorders, | | |
| | | | | cases of DIC. Congenital | | |
| | | | | ent venous thrombosis. This assay | | |
| | • | | • | e Main Haematology Section on | | |
| | | _ | tion of Thrombophi | | | |
| | - | | eived within 4 ho | | | |
| | Thromboph | nilia request | form FOR-CUH-PAT | I-1575, including documentation of the second se | | |
| | patient con | isent, must b | e received with all r | requests and is available on the CU | | |
| | website. | | | | | |
| Turnaround: | | ecimens: 3 - | · 4 weeks | | | |
| | • | | matology Section o | n Coagulation). | | |
| Ref. Range: | Age | Mean (%) | Range (%) | | | |
| iten italiger | Day 1 | 35 | 17 - 53 | | | |
| | Day 5 | 42 | 20 - 64 | | | |
| | Day 30 | 43 | 21 - 65 | | | |
| | Day 90 | 54 | 28 - 80 | | | |
| | Day 180 | 59 | 37 - 81 | | | |
| | Adult | 95 | 70 - 120 | | | |
| Protein S | Addit | 55 | 70 120 | | | |
| Laboratory: | Haematolo | <u>av</u> | | | | |
| | | | tto® (andium situat | - 2 20/) | | |
| Specimen: | | • | tte® (sodium citrat | | | |
| | | Specimens that are haemolysed, underfilled or overfilled cannot be analysed, check coagulation sample bottles are not expired to ensure correct filling. | | | | |
| . . | - | | | | | |
| Comment: | Test available Monday to Friday, during routine working hours. Protein S is a | | | | | |
| | vitamin K dependent protein, which serves as a co – factor for the | | | | | |
| | anticoagulant activity of activated protein C in the degradation of factors V | | | | | |
| | and VIII. This assay forms part of the Thrombophilia screen, see Main | | | | | |
| | Haematology Section on Guidelines for Investigation of Thrombophilia. | | | | | |
| | Samples must be received within 4 hours | | | | | |
| | Thrombophilia request form FOR-CUH-PAT-1575, including documentation of | | | | | |
| | patient consent, must be received with all requests and is available on the | | | | | |
| | CUH websi | | | | | |
| Turnaround: | 3 – 4 week | S | | | | |
| Ref. Range: | | | | | | |
| | Ag | е | Range | | | |
| | Da | y 1 | 12-60% | | | |
| | Da | у 5 | 22-78% | | | |
| | Da | y 30 | 33-93% | | | |
| | | y 90 | 54-118% | | | |
| | | y 180 | 55-119% | | | |
| | | ult male | 68% - 139% | | | |
| | A 4 | ult famala | 60 114 0/ | | | |

Adult female

60 - 114 %

| Title: Laboratory Medicine User Handbook | Reference: | PPG-CUH-PAT-31 | Revision: 22 |
|--|--------------|------------------------|------------------|
| | Active Date: | 03/11/2023 | Page: 175 of 206 |
| | Approved By: | Dr Vitaliy Mykytiv, Ms | Sinead Creagh |
| | Author: | Mr Paul Cantwell | |

| Laboratory: Clinical Biochemistry Specimen: Spot urine Turnaround: 1day Ref. Range: Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate. Prothrombin DNA Mutation Studies (G20210A) Laboratory: Haematology Molecular Genetics Specimen: Blood 3mL purple Vacuette® (EDTA) Comment: Forms part of a Thrombophilia screen. Thrombophilia request form FOR-CUH-PAT-1575, including documentation of patient consent, must be received with all requests and is available on the CUH website. Turnaround: 6 - 8 weeks Report: (Negative/Positive-Heterozygous /Homozygous), see final report Prothrombin Time (PT) Laboratory: Laboratory: Haematology Specimens which are haemolysed, underfilled or overfilled cannot be analysed, check coagulation sample bottles are not expired to ensure correct filling). Comment: Test available Monday to Friday, during routine working hours and for emergency reasons at all other times. The test is used as a screen to detect (a) single or combined deficiencies of the extrinsic coagulation system, (b) liver disease (c) vitamin K deficiency (d) monitoring oral anticoagulants, I assaying the specific coagulation Factor II. It also forms part of the Thrombophilia and/or Lupus screen. Specimens must be received within 48hrs | Protein/Creati | nine Ratio (Urinary) | | | | | |
|---|----------------|-------------------------------|--|--|--|--|--|
| Speciment Spot urine Turnaround: 1day Ref, Range: Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate. Prothrombin DNA Mutation Studies (G20210A) Laboratory: Haematology Molecular Genetics Specimen: Blood 3mL purple Vacuette® (EDTA) Comment: Forms part of a Thrombophilia screen. Turnaround: 6 - 8 weeks Report: (Negative/Positive-Heterozygous /Homozygous), see final report Prothrombin Time (PT) Laboratory: Laboratory: Haematology Specimens which are haemolysed, underfilled or overfilled cannot be analysed, check coagulation sample bottles are not expired to ensure correct filling). Comment: Test available Monday to Friday, during routine working hours and for emergency reasons at all other times. The test is used as a screen to detect (a) single or combined deficiencies of the extrinsic coagulation system, (b) liver disease (c) vitanin K deficiency (d) monitoring oral anticoagulants, I assaying the specific coagulation Factor II. It also forms part of the Thrombophilia and/or Lupus screen. Specimens must be received within 48hrs Many commonly administered drugs may affect the results. This should be kept in mind especially when unusual or unexpected results have been obtained. The prothrombin time (measured in seconds) is a very sensitive test to ad | | | | | | | |
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| The test is used as a screen to detect (a) single or combined deficiencies of the extrinsic coagulation system, (b) liver disease (c) vitamin K deficiency (d) monitoring oral anticoagulants, I assaying the specific coagulation Factor II. It also forms part of the Thrombophilia and/or Lupus screen. Specimens must be received within 48hrs Many commonly administered drugs may affect the results. This should be kept in mind especially when unusual or unexpected results have been obtained. 'The prothrombin time (measured in seconds) is a very sensitive test to advancing liver disease in patients with liver disorders. The PT ratio – the patients PT over the midpoint of the normal range is useful. The laboratory recognises that some protocols dealing with liver disease and paracetamol overdose use the INR. This is a less sensitive measure of liver disease as it is adapted for patients on warfarin.Turnaround: Ref. Range:Age Mean Range (seconds) Day 1 13.0 10.1 – 15.9 Day 5 12.4 9.5 – 15.3 Day 30 11.8 9.3 – 14.3 Day 90 11.9 9.6 – 14.2 Day 180 12.3 10.7 – 13.8 Adult See final reportPSA TotalLaboratory: Clinical Biochemistry Specimen:4.0 mL blood in plain tube (clotted sample) Turnaround: 4 Days Ref. Range: | Comment: | | | | | | |
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| | кег. кange: | appropriate. | | | | | |

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| PTH | | | | |
|-----------------------|---|--|--|--|
| Laboratory: | Clinical Biochemistry | | | |
| Specimen: | 4.0 mL EDTA plasma | | | |
| Turnaround: | 1 week | | | |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as | | | |
| _ | appropriate. | | | |
| Purines & Pyri | midines | | | |
| Laboratory: | Referred from Biochemistry to the Purine Research Lab, St. Thomas's Hospital, London | | | |
| Specimen: | Spot Urine (5-10mls) on ice – must be frozen immediately. EDTA blood (2-5mls) | | | |
| Comment: | Consultant request only | | | |
| Turnaround: | 5 Weeks | | | |
| Pyruvate Kinas | | | | |
| | | | | |
| Laboratory: | Sample referred from Haematology to The Red Cell Centre, King's College Hospital, London, SE5 9RS Westminister Bridge Rd., London0044 2032 999000 | | | |
| Specimen: | Blood 3mL, purple Vacuette [®] (EDTA), minimum 1 mL. | | | |
| Comment: | Request must be booked in advance with the Haematology Laboratory CUH, performed as part of the investigations into haemolytic anaemias. | | | |
| Turnaround: | 60 days | | | |
| Report: | Sent to referring clinician and copy filed in laboratory | | | |
| Q Fever | | | | |
| See <i>Coxiella b</i> | ournetii IgG and IgM | | | |
| QuantiFERON ® | -TB Gold Plus test (QFT) | | | |
| Laboratory: | Microbiology (TB Laboratory) | | | |
| Specimen: | Special kit available from the Microbiology Laboratory after prior agreement with medical team. Please follow the manufacturers instructions supplied with the kit. Note: | | | |
| | Fill to black mark on tube; under or overfilled bottles are not accepted. Immediately after filling tubes shake 10xtimes; just firmly enough to ensure the entire inner surface of the tube is coated with blood to dissolve antigens on tube walls. Hand-write patient details on tubes. Return the complete kit (in box) accompanied by a green Microbiology request form. | | | |
| Comment: | Errors in collecting or transporting blood specimens can decrease the accuracy of QFT. Do not refrigerate the kit at anytime. Blood specimens must be processed as soon as possible after collection while white blood cells are still viable. Before the QFT is conducted, confirm arrangements for testing with the laboratory. QuantiFERON®-TB Gold Plus test (QFT) - Specimens are only accepted by this laboratory Monday to Thursday before 4pm (excluding Bank Holidays). All samples received after this time will not be processed. Samples are also not accepted any day preceding a Bank Holiday (i.e. February bank holiday & St. Patricks day) Test performed by reference laboratory (Eurofins Biomnis, Sandyford | | | |
| Turnaround: | Industrial Estate). 2 Weeks | | | |

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| Report: | Positive (≥0.35), ne | egative (<0.35) | or indeterminate. | |
| | A positive result suggests that <i>M. tuberculosis</i> infection is likely; a n | | | |
| | | | | ninate result suggests |
| | QFT-G results canno | | | |
| | A positive result doe | es not distingui | sh between active | and latent infection. A |
| | repeat will be reque | sted where san | nples are close to (|).35 cut-off. |
| R90 gene pan | el for Inherited Coag | gulation bleed | ling, Thrombotic | and Platelet Disorder |
| Laboratory: | Referred from Haema | | rd Regional Genetic | s Laboratories |
| Specimen: | 2-5 ml peripheral blo | • | | |
| Comment: | - | ematology, trai | nsport within 24 ho | urs, complete form fror |
| | referral laboratory | | | |
| Turnaround: | Urgent 21 working da | • • • | | ing days |
| Report: | Sent to referring clini | cian and copy f | filed in laboratory | |
| Renal Biopsy | | | | |
| Laboratory: | Histopathology (Rer | • | lectron Microscopy | Department) |
| Specimen: | Renal Biopsy (unfixed | ed tissue)* | | |
| Comment: | Specimens are acce | pted Mon – Fri | 8am to 3:30pm. | |
| | It is essential to inform the laboratory in advance of the date and | | | |
| | approximate time of the procedure at Ext.21315. | | | |
| | On the day of the procedure, the specimen container for the biopsy is | | | |
| | collected from the EM/Renal laboratory. This consists of a universal container | | | |
| | with filter paper soaked in Phosphate Buffered Saline, into which the tissue is | | | |
| | placed directly after the procedure. The tissue is then brought to the Renal/EM department, where it is handed | | | |
| | | | | |
| | | | | d into portions for Light |
| | in the EM/Renal Lab | | Lence Microscopy a | nd Electron Microscopy |
| | *Note: All Renal Tra | | as are processed in | -house and |
| | slides/images are th | | - | |
| Turnaround: | | | | |
| ranaroanar | 80% of clinically urgent cases verbally reported in 2 days 80% of all cases fully authorised in 2 weeks | | | |
| | Approx. 6 weeks for renal transplant case referred to Beaumont | | | |
| Renal Stone | | | | Deddmone |
| Laboratory: | Sample referred fro | m Clinical Bioch | nemistry to the Mai | er Hospital Dublin |
| Specimen: | Sample referred from Clinical Biochemistry to the Mater Hospital Dublin. Renal Stone | | | |
| Comment: | Renal Stone assayed for NH4, Uric acid, Cystine, CO ₂ , Oxalate, Calcium, | | | |
| comment. | Phosphate, Magnesi | | | |
| Turnaround: | | | | |
| Ref. Range: | | ct Biochemistry | Dept. Mater Hospi | tal |
| | dosterone/Renin rat | | | |
| Retinol Bindir | • | | | |
| Laboratory: | Referred from Clinical Diagnostic Service | Biochemistry to | o Sheffield Norther | n General's PRU |
| Specimen: | 1 ml Serum | | | |
| • - | 2 ml Urine | | | |
| Turnaround: | 1 week from receipt in | Referral labora | atory | |
| Ref. Range: | See report or contact | | | |
| _ | ph: +44 (0) 114-271- | 5552 (Technica | I & Clinical Queries | 5) |
| | | | | |

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Respiratory Viral Screen (Molecular)

| | r al Screen (Molecular) Microbiology | | | |
|---|---|--|--|--|
| Laboratory: | 5, | | | |
| Specimen: Viral swab (nasopharyngeal, nose, throat), nasopharyngeal aspira broncho-alveolar lavage | | | | |
| | | | | |
| | Do not send through the pneumatic tube. | | | |
| | Note: If there are two swabs in the viral swab collection kit, please use the | | | |
| | thinner flocked swab only for combined throat and nasopharngeal sampling | | | |
| | and discard the thicker cotton swab. | | | |
| | Handwritten request for to accompany iCM request where Full respiratory Multiplex testing (except for Influenza) is required | | | |
| Comment: | During Influenza season, a Respiratory viral screen typically includes SARS Co V 2, Influenza A and B, Respiratory Syncytial Virus (RSV), Human | | | |
| | Metapneumovirus among others. | | | |
| | Influenza A & B, SARS Co V2, RSV and Human metapneumovirus are INAB accredited tests. | | | |
| | A rapid result is available when clinically indicated, but only when requested through prior consulation with the medical microbiology team. Only viral swabs will be accepted for this rapid test. | | | |
| | A negative result may not exclude infection | | | |
| Turnaround: | 24 hours in season, may be up to 5 working days out of season | | | |
| Report: | Detected, Not Detected, Inconclusive or Inhibited | | | |
| Reticulocyte C | | | | |
| Laboratory: | Haematology | | | |
| Specimen: | Blood 3mL purple Vacuette [®] (EDTA) | | | |
| • | Paediatric (1mL purple (EDTA) or 1.3 mL red) | | | |
| Comment: | The number of reticulocytes present in blood is an index of RBC production | | | |
| | by the bone marrow. Specimen must be <12 hours. | | | |
| Turnaround: | Emergency specimens: < 2 hours | | | |
| i annai e annai | Routine specimens: 8 – 24 hours | | | |
| Ref. Range: | Refer to Full Blood Count | | | |
| Ken Kungei | reference range. | | | |
| Rheumatoid Fa | | | | |
| Laboratory: | Autoimmune Serology | | | |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) | | | |
| Comment: | Quantitative Nephelometric assay. | | | |
| Turnaround: | 24 Hours | | | |
| Ref. Range: | | | | |
| Ribosomal P P | 0 - 14 IU/mL | | | |
| Laboratory: | Autoimmune Serology | | | |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) | | | |
| | | | | |
| Comment: | Qualitative Elisa. Automatically undertaken on all Anti-ENA positive sera. | | | |
| | Turnaround: 72 Hours | | | |
| Ref. Range: | Not applicable | | | |
| Rickettsia Anti | | | | |
| Laboratory: | Microbiology (Infectious Diseases Serology) | | | |
| Specimen: | 4mL clotted blood | | | |
| Comment: | Performed by a reference laboratory (Rare & Imported Pathogens Laboratory (RIPL), Porton Down) | | | |
| Turnaround: | 28 working days | | | |
| Report: | Qualitative result | | | |
| Rivaroxaban | | | | |
| | ect Oral Anti-coagulants. | | | |
| <u></u> | | | | |

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| Do (66-A) | | | | | |
|---|---|--|--|--|--|
| Ro (SS-A) | | | | | |
| Laboratory: | Autoimmune Serology | | | | |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) | | | | |
| Comment: | Qualitative Elisa. Automatically undertaken on all Anti-ENA positive sera. | | | | |
| Turnaround: | 72 Hours | | | | |
| Ref. Range: | Not applicable | | | | |
| Rotavirus / Ade | enovirus Assay | | | | |
| Laboratory: | Microbiology (Category 3 Laboratory) | | | | |
| Specimen: | Fresh faeces specimen. 1-2g is sufficient. | | | | |
| Comment: | Immunochromatographic test using anti-Adenovirus monoclonal and anti- | | | | |
| | Rotavirus monoclonal reagents. Test performed Monday to Friday 9-5pm on | | | | |
| | children <5 years. | | | | |
| Turnaround: | 24 hours. | | | | |
| | Positive reports are telephoned when available to the requesting area. | | | | |
| Report: | Positive or negative for Rotavirus and Adenovirus | | | | |
| Rubella IgG An | tibody | | | | |
| Laboratory: | Microbiology (Infectious Diseases Serology) | | | | |
| Specimen: | 4mL clotted blood | | | | |
| Comment: | This test is used in the determination of immune status to rubella. Typically, | | | | |
| | this test is done as part of an antenatal or occupational health screen. | | | | |
| Turnaround: | 36 hours | | | | |
| Report: | Quantitative value (IU/mL) | | | | |
| Rubella IgM An | tibody | | | | |
| Laboratory: | Microbiology (Infectious Diseases Serology) | | | | |
| Specimen: | 4mL clotted blood | | | | |
| Comment: | Patient history required. The presence of IgM antibodies suggests | | | | |
| | current/recent infection with the virus. | | | | |
| Turnaround: | 36 hours | | | | |
| Report: | Qualitative result | | | | |
| Salicylate | | | | | |
| Laboratory: | Clinical Biochemistry | | | | |
| Specimen: | 4.0 mL blood in a plain tube (clotted sample) | | | | |
| Turnaround: | 1 Hour | | | | |
| Ref. Range: | In adults, symptoms of Salicylate toxicity may occur at levels >300mg/L | | | | |
| SARS CoV-2 (M | | | | | |
| • | | | | | |
| , | 5, | | | | |
| | o not send through the pneumatic tube. | | | | |
| | lote: If there are two swabs in the viral swab collection kit, please use the | | | | |
| | ninner flocked swab only for combined throat and nasopharngeal sampling | | | | |
| | nd discard the thicker cotton swab. | | | | |
| Report: Rubella IgG Am Laboratory: Specimen: Comment: Turnaround: Report: Rubella IgM Am Laboratory: Specimen: Comment: Turnaround: Report: Salicylate Laboratory: Specimen: Turnaround: Ref. Range: SARS CoV-2 (M aboratory: M Specimen: V D N | Positive reports are telephoned when available to the requesting area. Positive or negative for Rotavirus and Adenovirus tibody Microbiology (Infectious Diseases Serology) 4mL clotted blood This test is used in the determination of immune status to rubella. Typical this test is used in the determination of immune status to rubella. Typical this test is done as part of an antenatal or occupational health screen. 36 hours Quantitative value (IU/mL) tibody Microbiology (Infectious Diseases Serology) 4mL clotted blood Patient history required. The presence of IgM antibodies suggests current/recent infection with the virus. 36 hours Qualitative result Clinical Biochemistry 4.0 mL blood in a plain tube (clotted sample) 1 Hour In adults, symptoms of Salicylate toxicity may occur at levels >300mg/L olecular) licrobiology iral swab (combined nasopharyngeal and throat) to not send through the pneumatic tube. Note: If there are two swabs in the viral swab collection kit, please use the hinner flocked swab only for combined throat and nasopharngeal sampling | | | | |

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| | | Author: | Mr Paul Cantwell | |
| Comment: | Nasopharyngeal swab universal transport me suitable sample types pneumatic tube . Note: If there are two thinner flocked swab o and discard the thicke A rapid SARS CoV-2 te requested through pri Only viral swabs will b | edia, viral trans for SARS-CoV- o swabs in the v only for combin r cotton swab. est is available or consulation v | port media or col 2 testing. Do nol viral swab collecti ed throat and nas when clinically in with the medical i | bas PCR media are t send through the on kit, please use the sopharngeal sampling dicated, but only when |
| | A negative result may | not exclude in | fection. | |
| Turnaround: | 24 hours, Urgent sam Microbiology medical t | | pritised with prior | approval with |
| Report: | Detected, Not detecte | <u>d, Inconclusi</u> ve | or Inhibited | |
| SARS CoV-2 / | Influenza A/B (Poir | nt of Care) | | |
| Specimen: | Viral swab (combined nasopharyngeal and throat) A cobas [®] PCR Media Dual Swab nasopharyngeal viral swab (yellow top) is the recommended sample type, a deep nasal /mid turbinate swab may be appropriate alternative in certain patient groups. | | | |
| Turnaround: Report: | < 1 hour. Detected, Not detecte | | 5 1 | |
| Specimen: | Viral swab (combined nasopharyngeal and throat) A cobas [®] PCR Media Dual Swab nasopharyngeal viral swab (yellow top) is the recommended sample type, a deep nasal /mid turbinate swab may be appropriate alternative in certain patient groups. | | | |
| Schistosoma | | | <u> </u> | |
| Laboratory: | Microbiology (Categ | orv 3 Laborato | rv) | |
| Specimen: | Collection of a term 2pm as this is the p containers without eggs may be found | inal urine spector period of maxim boric acid mustor trapped in the | timen is recomment tum schistosomal be used. In patie blood and mucus | nded (between 10am an activity). Sterile ents without haematuria, in the terminal portion lays of over 48 hours are |
| Comment: | | ollection, it is a | dvisable to add 1r | ne cannot be examined mL of undiluted formalin |
| Turnaround: | 24 hours | - | | |
| Report: | Schistosoma spp. N | ot seen or Sch | istosoma seen | |
| | Antibodies (Bilharzia | | | |
| Laboratory: | Microbiology (Infect | | Serology) | |
| Specimen: | 4mL clotted blood | | | |
| Comment: | Performed by a refe Laboratory (NPRL), | | ry (National Paras | sitology Reference |
| Turnaround: | 28 working days | | | |
| Report: | Qualitative result | | | |

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| SCL-70 | |
|--------------|--|
| Laboratory: | Autoimmune Serology |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) |
| Comment: | Qualitative Elisa. Automatically undertaken on all Anti-ENA positive sera. |
| Turnaround: | |
| Ref. Range: | - |
| Selenium | |
| Laboratory: | SAS Trace Element Unit, Southhampton University Hospitals NHS Trust |
| Specimen: | 2 ml Sod Hep Trace metal free plasma |
| Turnaround: | |
| | • |
| Report: | Up-to-date reference intervals will be applied to all Biochemistry reports as |
| Serotonin | appropriate |
| | Referred from Clinical Biochemistry to Leeds General Infirmary |
| Laboratory: | |
| Specimen: | 3 ml EDTA whole blood – FROZEN |
| Comment: | Supply platelets count info |
| | Serotonin is primarily raised in classical metastatic mid-gut carcinoid tumours. It is taken up readily by platelets or converted to 5-HIAA. Whole |
| | blood serotonin is measured and related to blood platelets. |
| Turnaround: | 20 days from receipt in Referral laboratory. |
| | |
| Ref. Range: | See report or contact Leeds General Infirmary +44 (0) 113 392 3285/3286 |
| SHBG | |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Comment: | SHBG is analysed (females only) in conjunction with testosterone. mindex |
| | (AI) is then calculated. |
| Turnaround: | |
| Ref. Range: | |
| | appropriate. |
| Sirolimus | |
| Laboratory: | Sample referred from Clinical Biochemistry to Harefield Hospital |
| Specimen: | 4.0 mL blood in an EDTA sample tube. |
| Turnaround: | |
| Ref. Range: | Interpretation of Sirolimus is dependent on time interval between sample and |
| | last dose, clinical indication for use of the drug, duration of therapy, other |
| | drug therapy and method of measurement |
| | blast Culture (Paediatric Neurology cases) |
| Laboratory: | Referred from Neuropathology to Sheffield Children's NHS Trust |
| Specimen: | 3x3mm skin bx taken into sterile culture medium |
| Comment: | Please contact Neuropathology in advance. Culture medium available from |
| | Neuropathology Lab. To arrive in Sheffield Children's NHS Trust no later than |
| | 4:30pm Mon-Fri. Protocols available on request. |
| Turnaround: | 14 weeks but may be longer depending on rate of cell line growth. |
| Skin Swab | |
| See Wound | Swab |
| Sm (Smith Ar | itigen) |
| Laboratory: | Autoimmune Serology |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) |
| Comment: | Qualitative Elisa. Automatically undertaken on all Anti-ENA positive sera. |
| Turnaround: | 72 Hours |
| Ref. Range: | Not applicable |
| Kel. Kange: | אטר מאטוורמטופ |

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| Small Round St | tructured Viruses (SRSV) |
|----------------|--|
| See Norovirus | |
| Smooth Muscle | e Antibodies |
| Laboratory: | Autoimmune Serology |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) |
| Comment: | Qualitative Immunofluorescence assay initially part of Auto Antibody Screen. Positive sera are titred to end point. Sera showing specific Anti-Actin pattern on Immunofluorescence are commented upon. |
| Turnaround: | 72 Hrs. |
| Ref. Range: | Not Applicable. |
| Sodium (Blood | |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Turnaround: | A/E or urgent sample: - 1 hour 30 mins. CUH wards, CUMH, SI, SF, SMOH, MGH: - 3 hours 182egain182. GP or OPD- Results posted within 4 days. |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate. |
| Sodium (Urina | ry) |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 24 Hr sample |
| Turnaround: | 1 Day |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate. |
| Spinal Muscula | r Atrophy (SMA) |
| Laboratory: | Referred from Biochemistry to National Centre for Medical Genetics (NCMG) |
| Specimen: | Infants: 1ml EDTA blood Adults 3-5ml EDTA blood |
| Comment: | Copy of NCMG request form available on website <u>www.genetics.ie</u> |
| Turnaround: | 6 weeks |
| Report: | Sent to referring clinician and copy of report filed in pathology |
| Sputum Cultur | |
| Laboratory: | Microbiology (Main laboratory) |
| Specimen: | Sputum from the lower respiratory tract expectorated by deep coughing. Check that specimen is of adequate quality as specimens of saliva and postnasal secretions are usually unsuitable. Ideally, the laboratory should receive a minimum volume of 1mL. The specimen should be collected into a clean, sterile, leakproof container. Sputum may be refrigerated for up to 2–3 hours without an appreciable loss of pathogens. Any delay beyond this time may allow overgrowth of Gram-negative bacilli, and <i>Haemophilus</i> species and <i>S. pneumonia</i> may die. Transport specimens ASAP. If processing is delayed, refrigeration is preferable to storage at ambient temperature. |
| Comment: | Please include any appropriate clinical details e.g. "Cystic fibrosis patient". If an unusual pathogen is suspected, the laboratory should be informed, <i>e.g.</i> <i>Burkholderia pseudomallei</i> and <i>Nocardia</i> sp require longer incubation of cultures. Refer to Mycobacteria testing for instructions for collection for TB culture. If a fungal infection is clinically suspected, please include as much information as possible regarding patient medical history, travel history and occupation, |
| Turnaround: | Prelim: 24 hours; Final: 4 days. Prolonged incubation is required for <i>Burkholderia</i> spp. And fungal culture, which are reported if positive. |
| Report: | Culture report: Any clinically significant isolate with the appropriate sensitivities. |

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| Stem cell enun | neration CD34 |
|------------------------|--|
| Laboratory: | Haematology (Flow Cytometry depatement) |
| Specimen: | 3 ml EDTA specimen peripheral blood |
| Comment: | Test performed only by prior arrangement with laboratory |
| Turnaround: | 48 hours |
| Report: | CD34 Quantitation – stem cells detected per ml |
| STD Screen | |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Tests: | Hepatitis B surface antigen, HIV Ag/Ab, syphilis antibody |
| Turnaround: | Negative samples: 36 hours. Please allow extra time for samples testing positive in house for HIV Ag/Ab and syphilis antibody (confirmatory testing required). |
| Report: | Qualitative result |
| | uid – Microscopy and Culture |
| Laboratory: | Microbiology (Main laboratory) |
| Specimen: | Specialist collection according to local protocols. Ideally, a minimum volume of 1mL should be collected into a clean, sterile, leakproof container. |
| | The volume of specimen influences the transport time that is acceptable. Large volumes of purulent material maintain the viability of anaerobes for longer. Results from delayed specimens must be interpreted with caution bearing in mind the difficulties in isolating anaerobes from these specimens Transport specimens ASAP. If processing is delayed, refrigeration is preferable to storage at ambient temperature. |
| Comment: | Test performed routinely Monday to Friday 9-5pm or by urgent request. |
| Turnaround: | Microscopy: 2 hours. Culture: Prelim: 24 hours; Final: 48-72 hours. Urgent report telephoned when available. |
| Report: | Total white cell count, differential leucocyte count (if appropriate), Gram Stain and Culture. All isolates are reported with appropriate sensitivities. Total white cell counts and differential leucocyte count are not performed on specimens containing a clot, which would invalidate the cell count. |
| Striated Muscle | |
| Laboratory: | Sample referred from Autoimmune Serology to Eurofins-Biomnis Laboratories. |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) |
| Turnaround: | Approx. 3 Weeks |
| Ref. Range: | See report form, or visit internet site <u>https://www</u> .eurofins.ie/biomnis/ for up to date referral test information |
| Strongyloides | Antibodies |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Comment: | Performed by a reference laboratory (National Parasitology Reference Laboratory (NPRL), London) |
| Turnaround: | 28 working days |
| Report: | Qualitative result |
| Strongyloides | Microscopy and Culture |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | Faeces |
| Comment: | Performed by a reference laboratory (National Parasitology Reference Laboratory (NPRL), London). Faecal specimens should NOT be refrigerated. |
| Turnaround: Report: | 28 working days Positive or negative |

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| Sweat Test | |
|-----------------|---|
| Laboratory: | Clinical Biochemistry |
| Comment: | Sweat is collected in GD ward, GC Day unit and from the Adult CF unit |
| Turnaround: | Done daily. |
| Ref. Range: | Contact CUH Immunology Laboratory |
| Synacthen Tes | |
| Laboratory: | Clinical Biochemistry |
| Specimen: | Timed serum samples |
| Comment: | Clearly indicate on request form and sample the time of sampling |
| Turnaound: | 3 days |
| Ref. range: | Up-to-date reference intervals will be applied to all Biochemistry reports as |
| | appropriate |
| Synovial Fluid | |
| | dy Fluid – Microscopy and Culture |
| Syphilis Antibo | |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Comment: | Sera positive by chemiluminescent immunoassay are further tested by RPR |
| | (Rapid Plasma Reagin) and possibly TPPA (Treponema pallidum Particle |
| | Agglutination). Positive samples may be sent to a reference laboratory for |
| | confirmation. |
| Turnaround: | Negative: 36 hours |
| | Positive samples: 14 working days |
| Report: | Qualitative result |
| t(11:14) molec | cular testing in Mantle Cell Lymphoma |
| Laboratory: | Referred by Pathology Laboratory to Cancer Molecular Diagnostics (CMD), St. |
| | James hospital |
| Specimen: | FFPE tissue block |
| Comment: | |
| Turnaround: | 18 working days (from date testing material is sent to referral institution) |
| Report: | Not applicable |
| Tacrolimus (FK | (506 / Prograf) |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in an EDTA tube |
| Comment: | Trough sample required. Analysed on Thursday. |
| Turnaround: | 1-2 days |
| Ref. Range: | Interpretation of Tacrolimus is dependent on time interval between sample |
| | and last dose, clinical indication for use of the drug, duration of therapy, |
| | other drug therapy and method of measurement. |
| TB – See Myco | bacteria testing |
| T-cell receptor | gene rearrangements (Clonality studies) |
| Laboratory: | Referred from Pathology to CMD, St. James Hospital |
| Specimen: | FFPE tissue block |
| Comment: | |
| Turnaround: | 19 working days (from date testing material is sent to referral lab) |
| Report: | Not applicable |
| Tear Duct – Cu | lture |
| See Lacrimal | |
| | |

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| Temporal Art | ery Biopsies |
|---------------|--|
| Laboratory: | Neuropathology |
| Specimen: | Formalin-fixed artery |
| Turnaround: | 3 days |
| Testosterone | |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Turnaround: | 1 Week |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as |
| | appropriate. |
| | |
| Tetanus antib | odies (IgG) |
| Laboratory: | Clinical Biochemistry |
| Specimen: | Blood 4mL red top Vacuette [®] (or similar container for clotted blood) |
| Comment: | Test performed by reference laboratory (Respiratory Infections Laboratory, Colindale, London). |
| Turnaround: | 6-7 weeks |
| Report: | Greater than 0.43IU/mL indicates previous exposure to tetanus toxoid. |
| Theophylline | |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Comment: | Take trough sample immediately before next dose. When making |
| | comparative measurements, it is advisable that sampling times be consistent |
| Turnaround: | , 5 1 |
| Ref. Range: | Therapeutic Range 10-20 mg/L Range quoted is appropriate for a trough sample. |
| Thiamine | |
| Laboratory: | Referred from Clinical Biochemistry to Biomnis Ireland, Dublin. |
| Specimen: | 2ml EDTA whole blood light protected |
| Comment: | Also referred to as Vitamin B1 or Aneurin |
| Turnaround: | 1-2 weeks |
| Report: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate. |
| Thioguanine | Nucleotides (TGN) |
| Laboratory: | Referred from Clinical Biochemistry, CUH to Purine Research Lab, St Thomas/Viapath |
| Specimen: | 4.0 mL blood EDTA sample (purple top) |
| Comment: | Store in fridge. Do not freeze |
| | Please provide a recent red blood cell result |
| Turnaround: | 3 weeks. |
| Ref. Range: | Refer to final report. |

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| Throat Swab | |
|--------------------------|--|
| Laboratory: | Microbiology (Main laboratory) |
| Specimen: | Swab the tonsillar area and/or posterior pharynx avoiding the tongue and uvula. Transport specimens ASAP in charcoal containing transport media. If processing is delayed, refrigeration is preferable to storage at ambient |
| | temperature. If diphtheria or gonorrhoea is suspected special testing should be requested. Ideally, inoculation of specimens for <i>N. gonorrhoeae</i> is made |
| | directly on to culture media at the bedside and incubated without delay. |
| | Specimens for viral isolation should be submitted in appropriate viral transport medium (available from Microbiology, CUH). |
| Comment: | Test performed routinely Monday to Friday 9-5pm or by urgent request. |
| Turnaround: | Culture Final: 24-48 hours |
| Report: | Culture for B-haemolytic streptococci, other bacteria (if appropriate), or yeasts. |
| Thrombophilia | |
| Laboratory: | Haematology |
| Specimen: | Three Blood 3mL, blue Vacuette® (sodium citrate 3.2%) and, |
| | One Blood 4mL red Vacuette (clotted specimen), |
| | One Blood 3mL purple Vacuette (EDTA specimen). Due to potential |
| | contamination of genetic material a separate EDTA sample is required. |
| | Samples must be received within 4 hours . Thrombophilia request form FOR-CUH-PAT-1575, including documentation of |
| | patient consent, must be received with all requests and is available on the |
| | CUH website. |
| | www.bcshguidelines.com/documents/Heritable_thrombophilia_bjh_07_2010.pdf |
| | Specimens that are haemolysed, underfilled or overfilled cannot be analysed, |
| | check coagulation sample bottles are not expired to ensure correct filling. Note: BCSH guidelines on Thrombophilia testing must be adhered to. |
| Comment: | Test available Mon to Fri, during routine working hours. |
| | Thrombosis occurs when activation of blood coagulation overwhelms the |
| | ability of the natural anticoagulant mechanism and fibrinolytic system to |
| | prevent thrombus formation taking place. Thrombophilia screen consists of: |
| | INR, APTT, FIB, Actin FSL, DVV test, Antithrombin 3, Protein C, Activated Protein C Resistance and Protein S assays. Anti-Cardinolipin and Beta 2- |
| | Glycoprotein 1 are also included as part of the screen if a clotted sample is |
| | received. |
| | Requests must conform with BCSH guidelines |
| | Samples without Request Form WILL NOT be processed. |
| Turnaround: | 3 – 4 weeks |
| Report: | Refer to final report for refenence intervals of individual assays |
| | & Thyroglobulin Antibodies |
| Laboratory: Specimen: | Sample referred from Clinical Biochemistry to Eurofins-Biomnis Laboratories 4.0 mL blood in Li Hep or plain tube (clotted sample) |
| Comment; | On patients with diagnosed thyroid cancer only. Consultant request only. |
| Turnaround: | 3 weeks |
| Ref. Range: | See report form, or visit internet site <u>https://www</u> .eurofins.ie/biomnis/_for up to |
| | date referral test information |
| Thyroid Antibo | dies (Anti-Thyroid Peroxidase Abs/ Anti-TPO Abs) |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in a plain tube (clotted sample) |
| Turnaround: | 4 days |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate. |
| | appropriate. |

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| Thyroid Stimul | ating Hormone (TSH) |
|----------------|--|
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Turnaround: | 4 days |
| Ref. Range: | Please contact Clinical Biochemistry lab for Paediatric and Pregnancy-related |
| 5 | Reference ranges. |
| | Up-to-date reference intervals will be applied to all Biochemistry reports as |
| | appropriate. |
| Thyroseq® | |
| Laboratory: | Referred from Cytology Laboratory in Pathology Dept. to Thyroseq |
| | International, University of Pittsburgh Medical Centre. |
| Specimen: | Thyroid FNA Thin Prep Smear or Thyroid FNA FFPE Cell Block |
| Comment: | For the diagnosis of Thy 3a/Thy 3f in Thyroid Cancers. |
| Turnaround: | 4-6 weeks |
| Tissue / Biops | v |
| Laboratory: | Microbiology (Main laboratory) |
| Specimen: | Tissue specimens for Microbiology must not be placed in formalin. The |
| 0000000 | specimen should be collected into a clean, sterile, leakproof container. For |
| | small specimens, add several drops of sterile saline to keep moist (include on |
| | label the nature of any additives e.g. 10mL saline). Do not allow tissue to dry |
| | out. Bone marrow aspirates should be inoculated directly into a blood culture |
| | bottle as per the Blood Culture guidelines. Transport specimens ASAP. If |
| | processing is delayed, refrigeration is preferable to storage at ambient |
| | temperature. It is vital that the specimen container is properly labelled. |
| Comment: | Test performed routinely Monday to Friday 9-5pm or by urgent request. The |
| | volume of specimen influences the transport time that is acceptable. Large |
| | volumes of purulent material maintain the viability of anaerobes for longer. |
| | The recovery of anaerobes is compromised if the transport time exceeds 3 |
| | hours. If a fungal infection is suspected, please include as much information |
| T | as possible regarding patient medical history, travel history and occupation. |
| Turnaround: | Culture: Prelim: 24 hours; Final: 48-72 hours |
| Report: | Culture report: Any clinically significant isolate with the appropriate sensitivities. |
| Tobramycin | sensitivities. |
| Refer to Antib | iotic Assavs |
| TORCH | |
| See Intra-Ute | rine Infection Screen |
| Toxicology / D | rug Screen: Blood |
| Laboratory: | Sample referred from Clinical Biochemistry to Department of Clinical |
| , | Biochemistry, Toxicology, Sandwell and West, posted Monday, Tuesday, |
| | Wednesday and Thursday. |
| Specimen: | 4.0 mL blood in a plain tube (clotted sample; non gel tube) |
| Comment: | Tested for Benzodiazepines, Barbiturates, Alcohol, Tricyclics. Drug screen |
| | measurement is provided for clinical purposes only. Samples will not be |
| | accepted for medicolegal or workplace testing |
| Turnaround: | 1 week |
| Ref. Range: | See report form or contact Department of Clinical Biochemistry, Toxicology, |
| | Sandwell and West |
| Toxicology / D | rug Screen: Urine |
| Laboratory: | Sample referred from Clinical Biochemistry to Toxicology Laboratory |
| | BEAUMONT Hospital Dublin, posted Monday, Tuesday, Wednesday and |
| | Thursday |

Thursday.

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| | | | | |
| Specimen: | Spot urine | | | |
| Comment: | | | | ocaine, Propoxyphene, |
| | measurement is pro | | | Alcohol. Drug screen |
| | accepted for medic | | | Samples will not be |
| Turnaround: | 1 week | | ace testing | |
| | | contact Beaum | ont Toxicology Der | nt Tel (01) 8092673 / |
| Ref. Range See report form or contact Beaumont Toxicology Dept. Tel (01) 80926 (01)8092675, Emergency after hours (087) 2590749, Fax (01) 80939 | | | | |
| Toxocara Antib | | | | , · · · · · (· _ / · · · · · · · · |
| Laboratory: | Microbiology (Infect | tious Diseases S | Serology) | |
| Specimen: | 4mL clotted blood | | | |
| Comment: | Performed by a refe | erence laborato | ry (National Parasi | tology Reference |
| | Laboratory (NPRL), | London) | | |
| Turnaround: | 28 working days | | | |
| Report: | Qualitative result | | | |
| | ondii IgG Antibody | | | |
| Laboratory: | Microbiology (Infec | tious Diseases S | Serology) | |
| Specimen: | 4mL clotted blood | | | |
| Turnaround: | 36 hours | | | |
| Report: | Qualitative result | | | |
| | ondii IgM Antibody | | | |
| Laboratory: | Microbiology (Infec | tious Diseases S | Serology) | |
| Specimen: | 4mL clotted blood | | | |
| Turnaround: | Negative samples: | | | |
| | – 28 working days | a IgM result mu | st be confirmed b | y a reference laboratory |
| Report: | Qualitative result | | | |
| TPMT Phenoty | | | | |
| Laboratory: | Sample referred fro | m Clinical Bioch | emistry to Dr Lor | etta Ford Clinical |
| Laboratory. | | | , | gham, West Midlands, |
| | B18 7QH Tel 0044 | | | |
| Specimen: | 5 – 10 mL EDTA w | | | |
| Turnaround: | 4 weeks | | | |
| Ref. Range | Contact laboratory | | | |
| Transferrin | | | | |
| Laboratory: | Clinical Biochemistr | 'Y | | |
| Specimen: | 4.0 mL blood in pla | in tube (clotted | sample) | |
| Turnaround: | 4 Days | - | | |
| Ref. Range: | Up-to-date reference | ce intervals will | be applied to all B | liochemistry reports as |
| | appropriate. | | | |
| % Transferrin | | | | |
| Laboratory: | Clinical Biochemistr | у | | |
| Specimen: | Not applicable | Tues T | F amila | |
| Comment: | Calculated from the | e fron and Trans | sierrin results. | |
| Turnaround: | 4 Days | | | |
| Ref. Range: | Contact biochemist | i y | | |
| Trichinella Ant | | tique Disesses (| Corology() | |
| Laboratory: | Microbiology (Infect | uous Diseases S | serology) | |
| Specimen: Comment: | 4mL clotted blood Performed by a refe | oronco laborato | ny (National Daras | tology Poforonco |
| comment. | Laboratory (NPRL), | | y (ivacional raidsi | torogy Relerence |
| | | Londony | | |

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| Turnaround: | 28 working days |
|-------------|--------------------|
| Report: | Qualitative result |

| Trichomonas v | aginalis |
|---|--|
| Laboratory: | Microbiology (Main laboratory) |
| Specimen: | Testing for Trichomonas vaginalis will not be performed unless a labelled |
| | slide is sent accompanying the swab. |
| | For <i>Trichomonas</i> , the posterior fornix should be swabbed. The slide should |
| | then be placed in a slide holder. |
| Comment: | This examination must be specifically requested. |
| Turnaround: | 24 hours. Trichemenae vaginalis seen er net seen |
| Report: Tricyclics | Trichomonas vaginalis seen or not seen |
| Laboratory: | Sample referred from Clinical Biochemistry to Department of Clinical |
| Laboratory. | Biochemistry, Toxicology, Sandwell and West Birmingham, posted Monday, |
| | Tuesday, Wednesday and Thursday. |
| Specimen: | Blood: 4.0 mL blood in a plain tube (clotted sample) |
| Comment: | See Toxicology / Drug Screen |
| Turnaround: | 1 week |
| Ref. Range: | See report form or contact Department of Clinical Biochemistry, Toxicology, |
| | Sandwell and West Birmingham |
| Triglycerides | |
| Laboratory: | Clinical Biochemistry |
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Comment: | Fasting sample required |
| Turnaround: | A/E or urgent sample: - 1 hour 30 minS. CUH wards, CUMH, SI, SF, SMOH, MGH: - 3 hours 189egain189. GP or OPD results posted within 4 days. |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate. |
| Troponin I – H | igh Sensitive |
| Laboratory: | Clinical Biochemistry |
| Specimen: 4.0 mL blood in plain tube (clotted sample) | |
| Turnaround: | 1 hour 15 mins |
| Ref. Range: | The 99 th . Centile is = <34 ng/L (male) |
| | is = <16 ng/L (female) Optimally for the biochemical diagnosis of MI it is recommended that two |
| | samples are taken for Troponin I (hs) measurement; the first at presentation |
| | and the second 3 to 6 hours later. |
| | In a patient with evidence of ischaemia: AMI is likely if, at least one result is |
| | > 34 ng /L (for males) or >16ng/L (for females) and Troponin I (hs) values |
| | change by 50% or more between the two samples. |
| | <i>cruzi</i> Antibodies |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| Comment: | Performed by a reference laboratory (National Parasitology Reference Laboratory (NPRL), London) |
| Turnaround: | 28 working days |
| Report: | Qualitative result |
| Tryptase (Mas | t Cell) |
| Laboratory: | Sample referred from Clinical Biochemistry to Eurofins-Biomnis Laboratories |
| Specimen: | 4.0 mL blood in Li Hep or plain tube (clotted sample) |
| | |

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| Comment: | Draw blood as soon and 8 hours after. | as possible aft | er anaphylactic sh | ock, again at 2 hours |
| Turnaround: | 3 weeks | | | |
| Ref. Range: | | visit internet s | ite https://www.euro | fins.ie/biomnis/ for up to |
| | date referral test inf | | | |
| Ttg (tissue Tra | ns Glutaminase ant | ibodies) | | |
| Laboratory: | Autoimmune Serolo | gy | | |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) | | | |
| Comment: | | Quantitative Immunoassay using Phadia Immunocap 250 analyser. Part of Coeliac screen. Anti EMA undertaken automatically on all positive sera to confirm. | | |
| Turnaround: | 24 Hours | | | |
| Ref. Range: | 0 - 2.5 AU/ML | | | |
| Tuberculosis T | esting | | | |
| Refer to Myco | bacteriology | | | |
| Tubule Antiboo | | | | |
| Laboratory: | Sample referred fro Laboratories. | | | |
| Specimen: | Blood, 4 mL red top | Vacuette (or s | imilar container fo | or clotted blood) |
| Turnaround: | Approx. 3 Weeks | visit internet a | | fine is /his mais / for up to |
| Ref. Range: | date referral test inf | | nte <u>nttps://www</u> .euro | fins.ie/biomnis/ for up to |
| U1RNP | | | | |
| Laboratory: | Autoimmune Serolo | gy | | |
| Specimen: | Blood, 4 mL red top | Vacuette (or s | imilar container fo | or clotted blood) |
| Comment: | Qualitative Elisa. Automatically undertaken on all Anti-ENA positive sera. | | | |
| Turnaround: | 72 Hours | | | |
| Ref. Range: | Not applicable | | | |
| Ulcer Swab | | | | |
| See Wound S | Swab | | | |
| Urate (Blood) | | | | |
| Laboratory: | Clinical Biochemistry | | | |
| Specimen: | 4.0 mL blood in plai | | | |
| Turnaround: | MGH: - 3 hours 190 | egain190. GP | or OPD- Results p | CUMH, SI, SF, SMOH, osted within 4 days. |
| Ref. Range: | appropriate. | | | Biochemistry reports as |
| Urate (Urinary | | | | |
| Laboratory: | Clinical Biochemistry | y | | |
| Specimen: | 24 Hour collection | | | |
| Turnaround: | 1 Day | | | |
| Ref. Range: | Up-to-date referenc appropriate. | e intervals will | be applied to all E | Biochemistry reports as |
| Urea (Blood) | | | | |
| Laboratory: | Clinical Biochemistry | | | |
| Specimen: | 4.0 mL blood in plai | • | | |
| Turnaround: | MGH: - 3 hours 190 | egain190. GP | or OPD- Results p | , CUMH, SI, SF, SMOH, osted within 4 days. |
| Ref. Range: | Up-to-date referenc appropriate. | e intervals will | be applied to all E | Biochemistry reports as |

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Urea (Urinary)

| Laboratory: | Clinical Biochemistry |
|-------------|---|
| Specimen: | Spot or 24 Hr urine sample |
| Turnaround: | 1 Day |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as |
| _ | appropriate. |

Urethral Swab

Refer to Genital swab

Urinary Legionella Antigen

| Laboratory: | Microbiology (Infectious Diseases Serology) | | | | |
|-------------------------|---|--|--|--|--|
| Specimen: | Urine | | | | |
| Comment: | Test performed only by special arrangement with Microbiology Consultant | | | | |
| Turnaround: | 36 hours | | | | |
| Report: | Positive or negative | | | | |
| Urinary Steroid Profile | | | | | |
| Laboratory: | Referred from Biochemistry to Kings College Steroid Lab, London | | | | |
| Specimen: | 24hur urine | | | | |
| Turnaround: | 5 weeks | | | | |
| Ref. Range: | See report form | | | | |
| Urinary Schisto | Urinary Schistosomiasis | | | | |

See Schistosoma haematobium

Urinary Streptococcus pneumoniae Antigen

| Laboratory: | Microbiology (Infectious Diseases Serology) |
|-------------|---|
| Specimen: | Urine |
| Turnaround: | 36 hours |
| Report: | Positive or negative |
| | |

Urine Microscopy and Culture

| Laboratory: | Microbiology (Main laboratory) | | |
|-------------|---|--|--|
| Specimen: | Ideally, a minimum of 8.5mL is required for routine culture. The specimen should be collected into a clean, sterile, leakproof 10ml BD Vacutainer® C&S Urine Tube for Culture and Sensitivity with boric acid. Note: A minimum of 8.5mL is <i>essential</i> for boric acid samples, where smaller volumes are collected, do not use a boric acid container but use a clean sterile leak-proof 20ml universal. | | |
| | Excessive fluid intake will dilute the urine and may decrease the colony count to $<10^5$ CFU/mL. | | |
| | Separate specimens must be collected for detection of Mycobacteria or <i>S. haematobium</i> . A fresh specimen is essential for the investigation of casts. | | |
| | | | |

Specimen Types

Midstream urine (**MSU**) Recommended for routine use. The first part of voided urine is discarded and without interrupting the flow, approximately 10mL is collected into a 10ml BD Vacutainer® C&S Urine Tube for Culture and Sensitivity with boric acid. The remaining urine is discarded.

Bag specimen urine (**BSU**). Used commonly for infants and young children. The sterile bags are taped over the genitalia and the collected urine is transferred to a sterile 10ml BD Vacutainer® C&S Urine Tube for Culture and Sensitivity with boric acid. There are frequent problems of contamination with this method of collection.

Clean catch urine (**CCU**). Thorough periurethral cleaning is recommended. The whole specimen is collected into a 10ml BD Vacutainer® C&S Urine Tube for Culture and Sensitivity with boric acid.

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Suprapubic aspirate (**SPA**). The use of this invasive procedure is usually reserved for clarification of equivocal results from voided urine e.g. in infants.

Catheter urine (**CSU**). May be obtained from suprapubic or per urethral catheters. The specimen should not be obtained from the collection bag.

Ileal conduit-urostomy urine is collected via a catheter passed aseptically into the stomal opening after removal of the external appliance. Results from this type may be difficult to interpret and should be performed only if there is an indication for treatment, such as pyrexia or constitutional upset.

Cystoscopy urine is obtained directly from the bladder using a cystoscope.

| Comment: | It is important that there should be minimal delay before culture. If processing is delayed >6 hours, refrigeration for up to 48 hours and use of boric acid containers is recommended. Ensure containers are filled to the line (8.5mL). | | |
|-------------|---|---|--|
| Turnaround: | Microscopy: | Routine: 24 hours. Urgent: 2 hours of receipt. | |
| | Culture: | Preliminary: 24 hours. Final: 24-72 hours | |
| Report: | Microscopy: | Report on the range of WBCs and RBCs per cmm as well as the presence of epithelial cells, casts, bacteria, yeasts and <i>Trichomonas</i> spp. (if present). | |
| | Culture: | Report bacterial growth in orgs/mL with sensitivities and comment where appropriate. Culture will only be carried out where WCC is >25/ μ L and bacteria is >200/ μ L, but the following are cultured in all cases; Antenatal, <16 year, Renal, ICU, potentially immunocompromised. | |

Valproate

| Laboratory: | Clinical Biochemistry |
|-------------|---|
| Specimen: | 4.0 mL blood in plain tube (clotted sample) |
| Comment: | Chronic oral dosing: trough sample immediately before next dose |
| Turnaround: | 1 Day |
| | |

Vancomycin

| Refer to Antib | iotic Assays | | |
|-----------------|---|--|--|
| Vancomycin Re | esistant Enterococci (VRE) | | |
| Laboratory: | Microbiology (Main laboratory) | | |
| Specimen: | Rectal swabs, placed in charcoal containing transport media. | | |
| Comment: | Test performed Monday to Friday 9-5pm. Label all Microbiology forms with | | |
| | VRE SCREEN. Indicate if the patient was previously VRE positive. Transport | | |
| | specimens ASAP. If processing of swabs is delayed, refrigeration is preferable to storage at ambient temperature. | | |
| Turnaround: | Prelim: 48 hours; Final: 48-72 hours | | |
| Report: | "VRE not isolated", | | |
| | Enterococcus faecium/faecalis (New VRE)/(VRE) isolated. | | |
| Varicella-zoste | r Virus IgG Antibody | | |
| Laboratory: | Microbiology (Infectious Diseases Serology) | | |
| Specimen: | 4mL clotted blood | | |
| Comment: | VZV IgG testing is performed on all antenatal patients | | |
| Turnaround: | 36 hours | | |
| Report: | Qualitative result | | |
| Varicella-zoste | r Virus Molecular | | |
| Laboratory: | Microbiology (Infectious Diseases Serology) | | |
| Specimen: | CSF (1mL), viral swab (skin, eye), vesicle fluid, skin scrapings | | |
| Comment: | Performed by a reference laboratory (National Virus Reference Laboratory (NVRL), Dublin) | | |
| Turnaround: | 14 working days | | |

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| Report: | Detected or not detected | | |
|-----------------|---|--|--|
| Vasculitic Scre | en | | |
| Laboratory: | Autoimmune Serology | | |
| Specimen: | Blood, 4 mL red top Vacuette (or similar container for clotted blood) | | |
| Comment: | Includes Auto Antibody Screen + Anti Neutrophil Cytoplasmic Antibody assay. | | |
| Turnaround: | 48 Hours or stat by contacting laboratory. | | |
| Ref. Range: | Not applicable. Refer to follow on tests if Screen Positive. | | |
| Very Long Cha | in Fatty acids | | |
| Laboratory: | Sample referred from Clinical Biochemistry to Willink Institute, Manchester. | | |
| Specimen: | 4.0 mL blood in EDTA or Lithium Heparin | | |
| Turnaround: | 3 weeks | | |
| Ref. Range: | See report form | | |
| Vincent's Angi | na | | |
| See Mouth Sw | vab | | |
| Viral Screen (E | Eye) | | |
| Laboratory: | Microbiology (Infectious Diseases Serology) | | |
| Specimen: | Viral swab | | |
| Tests: | Adenovirus, herpes simplex virus 1/2, varicella-zoster Virus (VZV) | | |
| Comment: | Performed by a reference lab (National Virus Reference Laboratory (NVRL), Dublin) | | |
| Turnaround: | 14 working days | | |
| Report: | Detected or not detected | | |
| Viscosity | | | |
| Laboratory: | Viscosity testing is referred from Clinical Biochemistry (Immunology section) to St. James' Hospital, Dublin | | |
| Specimen: | 2 samples in EDTA bottles. | | |
| Comment: | Viscosity >2.9 associated with Hyperviscosity Syndrome | | |
| Turnaround: | 3 Days | | |
| Ref. Range: | Refer to Haematology Dept. St. James Hospital. | | |
| Vitamin A (Ret | inol) | | |
| Laboratory: | Sample referred from Clinical Biochemistry to Nutristasis Unit, St. Thomas Hospital, London. | | |
| Specimen: | 4.0 mL blood in a plain tube (clotted sample) | | |
| Comment: | Consultant request only. Protect from light. | | |
| Turnaround: | 5 weeks | | |
| Ref. Range: | See report form, or visit internet site www.nutristasis.com for up to date referral test information | | |
| Vitamin B12 | | | |
| Laboratory: | Haematology | | |
| Specimen: | Blood 4mL red Vacuette (clotted specimen). | | |
| | | | |

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| Comment: | Test available Monday to Friday, during routine working hours. Vitamin B12 is a coenzyme necessary to the biosynthesis of DNA and RNA. Deficiency in man is associated with megaloblastic anaemia it is also vital to the normal metabolism of folic acid. It is of particular importance to recognise vitamin B12 deficiency as it causes both neurologic and psychiatric damage, which is preventable when diagnosed at an early stage. Values between 120 and 135 ng/l are considered indeterminate and should be interpreted in conjunction with full blood count results (including macrocytosis and clinical parameters). B12 and Folate should be requested for investigation of abnormal FBC results and relevant clinical syndromes. |
|----------------|---|
| | Use of haematinics for screening of well patients is not recommended. |
| | Requests should be accompanied by clinical details. See BCSH guidelines. |
| | The diagnosis of B12 and folate deficiency |
| | http://onlinelibrary.wiley.com/doi/10.1111/bjh.12959/pdf |
| Turnaround: | 7 working days |
| Ref. Range: | 140 – 844ng/L |
| | 120 – 170 ng/L indeterminate These are ADULT ranges – for guidance only |
| 1. 25 Dihvdrox | y Vitamin D (Calcitrol) |
| Laboratory: | Sample referred from Clinical Biochemistry to Eurofins-Biomnis Laboratories |
| Specimen: | MI blood in a plain tube (clotted sample) on ice, must be frozen < 1 hr. |
| | (minimum 2.0 mL serum required) |
| Comment: | Consultant request only. |
| Turnaround: | 3 weeks |
| Ref. Range: | See report form, or visit internet site https://www.eurofins.ie/biomnis/ for up to date referral test information. |
| Vitamin D (25) | Hydroxy Vitamin D) / Hydroxycholecalciferol |
| Laboratory: | Clinical Biochemistry |
| , Specimen: | 4.0 mL blood in a plain tube (clotted sample). |
| Comment: | Appropriate clinical details essential |
| Turnaround: | 10 days |
| Ref. Range: | Up-to-date reference intervals will be applied to all Biochemistry reports as appropriate. |
| Vitamin E (Toc | |
| Laboratory: | Sample referred from Clinical Biochemistry to Nutristasis Unit, St. Thomas Hospital, London |
| Specimen: | 4.0 mL blood in a plain tube (clotted sample). |
| Comment: | Sample must be separated < 1 hour. |
| Turnaround: | 5 weeks |
| Ref. Range: | See report form, or visit internet site <u>www.nutristasis.com</u> for up to date referral test information |
| Vitamin K (Phy | /tonadione) |
| Laboratory: | Sample referred from Clinical Biochemistry to Nutristasis Unit, St. Thomas Hospital, London |
| Specimen: | 4.0 mL blood in a plain tube (clotted sample) on ice, must be separated and frozen within 1 hour |
| Comment: | Protect from light. Consultant request only. |
| Turnaround: | 5 weeks |
| Ref. Range: | See report form, or visit internet site <u>www.nutristasis.com</u> for up to date referral test information |

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| Von-Willebran | d Multimers / Collagen binding |
|----------------------------------|--|
| Laboratory: | Referred from Haematology Dept. National Coagulation Laboratory, Centre for Clinical Pathology and Laboratory Medicine (CPLM), St James Hospital, Dublin 8 |
| Specimen: | Blood 3mL; blue Vacuette [®] (sodium citrate 3.2%) x 3 |
| Comment: | This is part of the Von Willebrand Screen which includes VW:Ag, VW:Rco, and Factor VIII. Multimers are only analysed in specific circumstances or on request by Coagulation Consultant. |
| Turnaround: | 90 days / 140 days (Working days) |
| Report: | Sent to referring clinician and copy filed in laboratory |
| Von Willebrand Antigen and Fa | d Screen: Ristocetin Co-factor vWF Activity, Von-Willebrand Factor actor VIII |
| Laboratory: | Haematology |
| Specimen: | Blood 3mL x 3, blue Vacuette® (sodium citrate 3.2%) Specimens that are haemolysed, underfilled or overfilled cannot be analysed, check coagulation sample bottles are not expired to ensure correct filling). |
| Comment: | Test available Monday to Friday, during routine working hours. Screen includes Factor V111 assay, vWF:ag (vW factor Ag), vWFactor Activity (Ristocetin Co-Factor) |
| Turnaraundu | Samples must be received within 4 hours |
| Turnaround: | 3 – 4 weeks |
| Ref. Range: | vWF activity: 0.55 – 1.56 IU/mL vWF Ag level: 0.50 – 1.60 IU/mL |
| | Factor VIII Adult 0.50 – 1.49 IU/mL |
| VWF Cleaving | Protease (vWFcp) Assay (ADAMTS13 Activity and Antibodies) |
| Laboratory | Refered from Haematology to HSL/TDL (Health Services Laboratories) Haemostasis Laboratory, Haematology Department, 60 Whitfield Street, London, W1T 4EU or Belfast Belfast Trust Health and Social Care Northern Ireland, Haemostasis Laboratory if Urgent |
| Specimen: Comment: | Blood 3mL blue Vacuette [®] (sodium citrate 3.2%) fill tube to mark. Request must be booked in advance with the Haematology Laboratory CUH. Requested by Consultant Haematologist for further investigation of von Willebrand Disease. ADAMTS13 Assay Request form must be completed, must be sent on dry ice and samples can only be referred Monday or Tuesday (via Eurofins-Biomnis). |
| Turnaround: | 60 days |
| Report: | Sent to referring clinician and copy filed in laboratory |
| Warfarin Plasn | na Resistance Concentration and gene |
| Laboratory: | Sample is referred from Haematology to The Centre for Haemostasis and Thrombosis, 1^{st} Floor North Wing, St Thomas' Hospital |
| Specimen: | $2 \times EDTA$ and $2 \times Citrate$, needs to be booked with the laboratory prior to sampling. |
| Comment: | Requested by Coagulation Consultant Super Warfarin (rodenticides) Vitamin K1 and PIVKA 11 are part of this profile reported and may be requested |
| Turnaround: | 21 days /80 days (Working days) |
| Report: | Sent to referring clinician and copy filed in laboratory |
| West Nile Viru | s Antibodies |
| Laboratory: | Microbiology (Infectious Diseases Serology) |
| Specimen: | 4mL clotted blood |
| | |

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| | | | | |
| Comment: | • | ference laborator | y (National Virus | Reference Laboratory |
| | (NVRL), Dublin) | | | |
| Turnaround: | By arrangement | | | |
| Report: | Qualitative result | | | |
| | ase (Tropheryma | | | |
| Laboratory: | Microbiology (Infe | | Serology) | |
| Specimen: | 4mL EDTA blood, | | | |
| Comment: | Street Hospital for | | | biology, Great Ormond |
| Turnaround: | 28 working days | | | |
| Report: | Detected or not de | | | |
| | ase (Tropheryma | | | |
| Laboratory: | Microbiology Main | Lab | | |
| Specimen: | CSF | | | |
| Comment: | PCR test performe | d by a reference | laboratory (Micro | pathology) |
| Turnaround: | 5 working days | | | |
| Report: | Detected or not de | etected | | |
| hooping Cou | - | | | |
| | a Species – Culture | | | |
| Vinter Vomitir | | | | |
| | s – Norwalk-like viru | | | . , |
| • | Skin / Abscess / D | | / Bite / Burn sv | vab) |
| Laboratory: | Microbiology (Mair | | | . |
| Specimen: | | | on request form. | Specimens of pus, if |
| | present, are prefe | | chould be cuppli | ed (ideally a minimum |
| | 1mL). | volume of zomi | | |
| | Swabs should be s | soaked in exudate | e where possible | Specimen a |
| | | | | est part of the wound, |
| | avoiding the super | | | |
| | | | | e that is accentable |
| Comment: | | cimen influences | the transport tim | |
| Comment: | The volume of spe | | | pility of anaerobes for |
| Comment: | The volume of spe Large volumes of longer. | purulent materia | maintain the vial | pility of anaerobes for |
| Comment: | The volume of spe Large volumes of longer. Specimens should the recovery of an | purulent materia be transported t aerobes is comp | l maintain the vial o the laboratory v romised. Results f | pility of anaerobes for within 3 hours after whi rom delayed specimen |
| Comment: | The volume of spe Large volumes of longer. Specimens should the recovery of an must be interprete | purulent materia be transported t aerobes is comp ed with caution b | l maintain the vial o the laboratory v romised. Results f | pility of anaerobes for within 3 hours after whi rom delayed specimen |
| Comment: | The volume of spectrum Large volumes of longer. Specimens should the recovery of an must be interprete anaerobes from the | purulent materia be transported t aerobes is comp ed with caution b sese specimens. | l maintain the vial o the laboratory v romised. Results f earing in mind the | pility of anaerobes for vithin 3 hours after whi rom delayed specimen e difficulties in isolating |
| Comment: | The volume of spectrum Large volumes of longer. Specimens should the recovery of an must be interprete anaerobes from the Routine processing | purulent materia be transported t aerobes is comp ed with caution b lese specimens. g of superficial sv | l maintain the vial o the laboratory v romised. Results f earing in mind the vabs of ulcers sho | pility of anaerobes for within 3 hours after whi rom delayed specimen e difficulties in isolating uld be discouraged. |
| Comment: | The volume of spec- Large volumes of longer. Specimens should the recovery of an must be interprete anaerobes from the Routine processing Swabbing dry crus | be transported t aerobes is comp ed with caution b lese specimens. g of superficial sy sted areas is unli | I maintain the vial o the laboratory v romised. Results f earing in mind the vabs of ulcers sho kely to be helpful. | pility of anaerobes for within 3 hours after whi rom delayed specimen e difficulties in isolating uld be discouraged. If specimens are take |
| Comment: | The volume of spec Large volumes of longer. Specimens should the recovery of an must be interprete anaerobes from th Routine processing Swabbing dry crus from ulcers the de | be transported t aerobes is completed with caution b bese specimens. g of superficial sw sted areas is unlil bris on the ulcer | I maintain the vial o the laboratory v romised. Results f earing in mind the vabs of ulcers sho kely to be helpful. should be remove | pility of anaerobes for within 3 hours after wh rom delayed specimen e difficulties in isolating uld be discouraged. If specimens are take ed, the ulcer cleaned w |
| Comment: | The volume of spec Large volumes of longer. Specimens should the recovery of an must be interprete anaerobes from the Routine processing Swabbing dry cruss from ulcers the de saline and either a | be transported t aerobes is completed with caution b bese specimens. g of superficial sw sted areas is unlil bris on the ulcer biopsy, or prefe | I maintain the vial o the laboratory v romised. Results f earing in mind the vabs of ulcers sho kely to be helpful. should be remove rably a needle asp | pility of anaerobes for within 3 hours after whi rom delayed specimen e difficulties in isolating uld be discouraged. If specimens are taken ed, the ulcer cleaned w piration of the edge of |
| Comment: | The volume of spec Large volumes of longer. Specimens should the recovery of an must be interprete anaerobes from the Routine processing Swabbing dry cruss from ulcers the de saline and either a the wound taken. | be transported t aerobes is comp ed with caution b bese specimens. g of superficial sy sted areas is unlil bris on the ulcer biopsy, or prefe A less invasive ir | I maintain the vial o the laboratory v romised. Results f earing in mind the vabs of ulcers sho kely to be helpful. should be remove rably a needle asp rigation-aspiration | bility of anaerobes for within 3 hours after wh from delayed specimen e difficulties in isolating uld be discouraged. If specimens are taken ed, the ulcer cleaned w biration of the edge of n method may be |
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| Comment: | The volume of spec Large volumes of longer. Specimens should the recovery of an must be interprete anaerobes from the Routine processing Swabbing dry cruss from ulcers the de saline and either a the wound taken. preferred. Using a ulcer margin and i | be transported t aerobes is completed with caution b bese specimens. g of superficial sy sted areas is unlil bris on the ulcer a biopsy, or prefe A less invasive ir small needle-les rrigate gently with | I maintain the vial o the laboratory v romised. Results f earing in mind the vabs of ulcers sho kely to be helpful. should be remove rably a needle asp rigation-aspiration s syringe, place th th at least 1mL st | within 3 hours after which rom delayed speciment e difficulties in isolating uld be discouraged. If specimens are taken ed, the ulcer cleaned with piration of the edge of the method may be the syringe tip under the erile saline without |
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| | The volume of spec Large volumes of p longer. Specimens should the recovery of an must be interprete anaerobes from the Routine processing Swabbing dry cruss from ulcers the de saline and either a the wound taken. preferred. Using a ulcer margin and i preservative. After further 1mL sterile approximately 0.2 Urgent microscopy (pus /fluid): Culture: | be transported t aerobes is completed with caution b bese specimens. g of superficial sw sted areas is unlil bris on the ulcer biopsy, or prefe A less invasive ir small needle-less rrigate gently wir massage of the saline. Massage 5mL of the fluid Within 2 hou | I maintain the vial o the laboratory v romised. Results f earing in mind the vabs of ulcers sho kely to be helpful. should be remove rably a needle asp rigation-aspiration s syringe, place th th at least 1mL sto ulcer margin, rep the ulcer margin and place in a ste urs of receipt. | vithin 3 hours after whi rom delayed specimen e difficulties in isolating uld be discouraged. If specimens are taken ed, the ulcer cleaned w piration of the edge of n method may be ne syringe tip under the erile saline without eat the irrigation with again, aspirate rile, leakproof containe Final report: 24-72 hou |
| | The volume of spec Large volumes of longer. Specimens should the recovery of an must be interprete anaerobes from the Routine processing Swabbing dry crus from ulcers the de saline and either a the wound taken. preferred. Using a ulcer margin and i preservative. After further 1mL sterile approximately 0.2 Urgent microscopy (pus /fluid): Culture: Microscopy: Rej | be transported t aerobes is completed with caution b bese specimens. g of superficial sw sted areas is unlil bris on the ulcer biopsy, or prefe A less invasive ir small needle-less rrigate gently wir massage of the saline. Massage 5mL of the fluid Within 2 hou | I maintain the vial o the laboratory v romised. Results f earing in mind the vabs of ulcers sho kely to be helpful. should be remove rably a needle asp rigation-aspiration s syringe, place th th at least 1mL st ulcer margin, rep the ulcer margin and place in a ste urs of receipt. | within 3 hours after whi rom delayed specimen e difficulties in isolating uld be discouraged. If specimens are taken ed, the ulcer cleaned w biration of the edge of n method may be ne syringe tip under the erile saline without eat the irrigation with again, aspirate rile, leakproof containe |

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| | Culture: | "No growth" or "skin flora" or report any clinically significant organism isolated with sensitivities. | |
|-------------|---|--|--|
| Zika Virus | | | |
| Laboratory: | Microbiology | / (Infectious Diseases Serology) | |
| Specimen: | 4mL clotted | blood (Serology), 4mL EDTA blood (Molecular) | |
| Comment: | Performed b (NVRL), Dub | by a reference laboratory (National Virus Reference Laboratory plin) | |
| Turnaround: | 14 days | | |
| Report: | Qualitative result (Serology), Detected or Not Detected (Molecular) | | |
| Zinc | | | |
| Laboratory: | Referred fro Guildford | m Clinical Biochemistry to SAS Laboratory for Trace Elements, | |
| Specimen: | 4.0 mL bloo | d in a metal-free plain tube (clotted sample). | |
| Turnaround: | 3 weeks | | |
| Ref. Range: | Up-to-date appropriate | reference intervals will be applied to all Biochemistry reports as | |

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14 GLOSSARY OF ABBREVIATIONS

The abbreviations used in this handbook include names of tests are in accordance with current use and accepted recommendations.

| current use | e and accepted recommendations. |
|-------------|--|
| ACE | Angiotensin converting enzyme |
| ACTH | Adrenocorticotrophic hormone |
| ADH | Antidiuretic hormone |
| AFB | Acid fast bacilli |
| AFP | Alpha-Fetoprotein |
| ALT | Alanine aminotransferase |
| ALP | Alkaline phosphatase |
| ANCA | Antineutrophil 198riiodothyr antibody |
| ANF | Antinuclear Factor |
| APC | Activated protein C |
| APTT | Activated partial Thromboplastin time |
| ASOT | Antistreptolysin O titre |
| AST | Aspartate aminotransferase |
| BJP | Bence Jones Protein |
| C3 | Third component of complement |
| C4 | Fourth component of complement |
| CA | Carbohydrate antigen (tumour markers) |
| CEA | Carcinoembryonic antigen |
| CK | Creatine kinase |
| CMV | Cytomegalovirus |
| CPE | Carbapenemase Producing Enterbacteriales |
| CRP | C-reactive protein |
| CSF | Cerebrospinal fluid |
| DDI | D-Dimers |
| DHEA | Dehydroepiandrosterone |
| DHEAS | Dehydroepiandrosterone sulphate |
| DVVT | Dilute Viper Venom test |
| EBV | Epstein Barr virus |
| EDTA | Ethylene diamine tetra-acetic acid |
| EGFR | Epidermal Growth Factor Receptor |
| EMA | Endomycial Antibodies |
| ENA | Extractable Nuclear Antigens |
| EPO | Erythropoietin |
| ESR | Erythrocyte sedimentation rate |
| FISH | Flourescence In Situ Hybridisation |
| FBC | Full blood count, full blood examination, complete blood count |
| FNAB | Fine needle aspiration biopsy |
| FSH | Follicle stimulating hormone |
| FT3 | Free Triiodothyronine (T3) |
| FT4 | Free thyroxine (T4) |
| GBM(Q) | Glomerular Basement Membrane Antibodies (Quick test) |
| GC | Gonococci |
| GGT | Gamma glutamyl transferase (transpeptidase) |
| GTT | Glucose tolerance test |
| HAV | Hepatitis A virus |
| Hb | Haemoglobin |
| HbA1c | Glycated haemoglobin |
| | |

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| HbA2 | Hoomoglobin A2 |
|------------------|---|
| HbF | Haemoglobin A2 Haemoglobin F, fetal haemoglobin |
| HbS | Sickle haemoglobin, haemoglobin S |
| HBsAg | Hepatitis B surface antigen |
| HBV | Hepatitis B virus |
| hCG | Human chorionic gonadotrophin |
| HCO₃ | Bicarbonate |
| HCT | Haematocrit, packed cell volume |
| HCV | Hepatitis C virus |
| HDL | High density lipoprotein |
| HDNB | Haemolytic Disease of the Newborn |
| hGH | Human growth hormone |
| HIAA | 5-Hydroxyindole acetate |
| HLA | Human leucocyte antigen |
| HMMA | 4-hydroxy-3-methoxymandelate |
| HPV | Human papillomavirus |
| HSV | Herpes simplex virus |
| HVA | Homovanillate |
| HVS | High Vaginal Swab |
| HZV | Herpes zoster virus (varicella-zoster) |
| ICCS | Intercellular cement substance |
| Ig | Immunoglobulin |
| IGF | Insulin-like growth factor |
| INR | International normalised ratio |
| IUCD | Intrauterine Contraceptive Device |
| kg | Kilogram |
| kPa | Kilopascal |
| KRAS | KRAS gene |
| LD | Lactate dehydrogenase |
| LDL | Low density lipoprotein |
| LGV | Lymphogranuloma venereum |
| LH | Luteinising hormone |
| MCH | Mean cell haemoglobin |
| MCHC | Mean cell haemoglobin concentration |
| MCV | Mean cell volume |
| MGUS | Monoclonal gammopathy of unknown significance |
| MMR | Measles, mumps, rubella IgG antibodies |
| MRSA | Methicillan-Resistant Staph aureus |
| MSI | Microsatellite Instability |
| MSU | , Midstream Urine |
| MTHFR | Methyltetrahydrofolate Reductase |
| PCR | Polymerase chain reaction |
| pCO ₂ | Partial pressure of carbon dioxide (CO ₂) |
| PCP | Pneumocystis jirovecii |
| PCV | Packed cell volume |
| PDL1 | Programmed Death Ligand-1 |
| PIE | Pulmonary infiltration with eosinophilia |
| PNH | Paroxysmal nocturnal haemoglobinuria |
| pO ₂ | Partial pressure of oxygen (O ₂) |
| PR | Prothrombin ratio |
| PSA | Prostate specific antigen |
| | |

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| PT PTH PTHrP RAST RCC RDW RFLP RPR RSV SHBG SLE | Prothrombin time Parathyroid hormone Parathyroid hormone related peptide Radioallergosorbent test- see specific IgE Red cell count Red cell distribution width Restriction fragment length polymorphism Rapid Plasma Reagin Respiratory syncytial virus Sex hormone binding globulin Systemic lupus erythematosus |
|---|---|
| SM | Smith Antigen |
| STI | Sexually transmitted infection |
| Т3 | Triiodothyronine |
| T4 | Thyroxine (tetraiodothyronine) |
| TBG | Thyroxine binding globulin |
| TORCH | Toxoplasma, rubella, cytomegalovirus, parvovirus B19 |
| TPPA | Treponema pallidum Particle Agglutination |
| TRH | Thyrotropin releasing hormone |
| TSH | Thyroid stimulating hormone |
| tTG | Tissue Trans Glutaminase Antibodies |
| VCA | Viral capsid antigen (EBV) |
| VIP | Vasoactive intestinal polypeptide |
| VRE | Vancomycin- Resistant Enterococci |
| vWf | von Willebrand factor |
| vWfAg | von Willebrand factor antigen |
| WCC | white cell count, leucocyte count |
| XDP | Cross linked fibrin degradation products, D-dimer |

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15 NAMES AND ADDRESSES OF REFERRAL LABORATORIES

| Name | Address | Referring Dept |
|---|---|--------------------------|
| Alpha One Foundation | RCSI Building, Beaumont Hospital, Dublin 9 | Biochemistry |
| Anaerobe Reference Laboratory | | |
| Analytical Services International Ltd | St. George's University Of London Cranmer Terrace, London SW17 ORE | Biochemistry |
| Antimicrobial Reference Laboratory | Department of Medical Microbiology Southmead Hospital Westbury on Trym Bristol BS10 5NB | Clinical Microbiology |
| Belfast City Hospital (CLL) | Molecular Haematology, Haematology Department, Belfast City Hospital, Belfast Health and Social Care Trust, 51 Lisburn Road, Belfast, UK BT9 7AB. | Haematology |
| Beaumont Hospital | Biochemistry Lab, Beaumont, Dublin 9 | Biochemistry |
| Biochemical Genetics Unit | Box 247Addenbrooke's Hospital Hills RoadCambridgeCB2 2QQ | Biochemistry |
| Biochemistry Department, St. James's Hospital | James's Street, Dublin 8, Ireland | Biochemistry |
| Biochemistry, Mater Misericordiae University Hospital (MMUH) | Eccles St., Dublin 7 | Biochemistry |
| Bristol Genetics Laboratory | North Bristol NHS Trust, Bristol Genetics Lab, Pathology Sciences, Southmead Hospital, Westbury-On-Trym, Bristol, BS10 5NB | Haematology |
| Brucella Reference Unit (BRU) | Liverpool Clinical Laboratories, Royal Liverpool and Broadgreen University Hospitals NHS Trust, Duncan Building, Prescot St., Liverpool L7 8XP, England | Clinical Microbiology |
| Cancer Molecular Diagnostics (CMD), St James Hospital | Cancer Molecular Diagnostics, SJH, LabMed Directorate, St. James's Hospital, Dublin 8, D08 W9RT | Haematology |
| Cardiff and Vale University University Hospital of Wales Cardiff CF 14 4XY Health Board, Dept of Medical Biochemistry | | Biochemistry |
| Central PathologyHaematology Laboratory, Central Pathology LaboratoryHaematology, St James's(CPL) Building, LabMed Directorate, St. James's Hospital, Dublin 8, D08 W9RT | | Haematology |
| Cholinesterase Investigation Unit | Pathology Sciences Building Southmead Hospital Westbury- on-Trym Bristol BS10 5NBUnited Kingdom | Biochemistry |
| City Hospital Birmingham Dr Jonathan Berg / Dr Loretta Ford City Hospital, Dudley Road, Birmingham, B18 7QH, UK | | Biochemistry |
| Clinical and Molecular Genetics Unit | Institute of Child Health.30 Guildford Street, London United Kingdom | Biochemistry |

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| Name | Address | Referring Dept |
|--|--|---|
| Clinical Biochemistry Department | Kings College Hospital Denmark Hill, LondonSE5 9RS, United Kingdom020 3299 9000 | Biochemistry |
| Viapath/Synnovis Consultant Renal Pathologist, Beamount | EM/Histopathology Department, Beaumont Hospital, Beaumont Road, Dublin 9, D09 A0KH | Pathology (Electron Microscopy) |
| Department of Cellular Pathology, The Adelaide and Meath Hospital | Department of Cellular Pathology, The Adelaide and Meath Hospital incorp. The National Children's Hospital, Tallaght, Dublin 24 | Pathology |
| Department of Clinical Chemistry and Newborn Screening, Sheffield | Sheffield Children's NHS Trust Western Bank Sheffield S10 2TH, United Kingdom | Biochemistry, Neuropathology |
| Department of Immunology,North General Hospital | Herries Road, Sheffield S5 7AU | Immunolgy |
| Department of Microbiology | Old Medical School, Leeds General Infirmary, Thoresby Place, Leeds LS1 3EX, England | Clinical Microbiology |
| Eurofins-Biomnis Ireland | Three Rock Road, Sandyford Business Estate, Dublin 18, Ireland | Biochemistry, Haematology, Microbiology |
| Eurofins-Biomnis SELA S | 17/19 Avenue Tony Garnier, 69007, Lyon, France | Neuropathology |
| Freeman Hospital | Freeman Hospital Freeman Road High Heaton Newcastle Upon Tyne NE7 7DNUnited Kingdom | Biochemistry |
| Galateau-Salle, Prof Francoise, | Department of Pathologique Route de la DeDelivandre CHU-Cote de Nacre,14033-CAENCEDEX, France | Pathology |
| Gastrointestinal Bacteria Reference Unit (GBRU) | Bacteriology Reference Department, UK Health Security Agency, 61 Colindale Avenue, London NW9 5HT, England | Clinical Microbiology |
| Genomic Health, Inc. | Genomic Health, Inc.,301 Penobscot Drive, Redwood City, CA 94063,USA | Pathology |
| Great Ormond Street Immunology | Great Ormond Street Immunology, Departments of Immunology and Clinical Molecular Genetics, Level 4 Camelia Botnar Laboratories, Great Ormond Street Hospital Great Ormond Street, NHS Trust, WC1N 3JH | Haematology |
| GSTS Pathology Kingspath Hospital, King's College Hospital NHS Foundation Trust | Mr Christopher Lambert, Red Cell Laboratory, c/o Main Pathology CSR, Viapath Analytics, Ground Floor Bessemer Wing, King's College Hospital, Denmark Hill, London SE5 9RS, United Kingdom | Haematology |
| Haematology, Our Lady's Hospital Crumlin | Our Lady's Children's Hospital, Division of Cytogenrtics (Oncology), Crumlin, Dublin 12, Ireland | Haematology |
| Haemostasis Molecular Diagnostics (HMD), St James Hospital | Haematology Dept. to Haemostasis Molecular Diagnostics (HMD), National Coagulation Laboratory, Centre for Clinical and Laboratory Medicine, CPLM, St James Hospital, Dublin 8 | Haematology |
| Harefield Hospital | Mr Neil Leaver Principal Clinical Scientist, Harefield Hospital,Harefield 90 UB United Kingdom | Biochemistry |
| Histopathology Department, SVUH | Histopathology Department, St. Vincent's University Hospital, Dublin | Pathology |

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| Name | Address | Referring Dept |
|--|---|--------------------------|
| HPA/PHE Laboratory | P.O. Box 209Manchester Medical Microbiology Partnership Clinical Sciences Building Manchester Royal Infirmary Oxford Road | Biochemistry |
| HSL (Health Services Laboratories) | HSL (Health Services Laboratories) Haemostasis Laboratory, Haematology Department, 60 Whitfield Street, London, W1T 4EU | Haematology |
| HSL (Health Services Laboratories) Advanced Diagnostics | HSL- AD, Ground Floor, 60 Whitfield Street, London, W1T 4EU | Pathology |
| Immunology Department and Protein Reference Unit | P.O Box 894 Sheffield S5 7YTUnited Kingdom | Biochemistry |
| Irish Meningitis & Sepsis Reference Laboratory (IMSRL) | The Children's University Hospital, Temple St, Dublin 1, Ireland | Clinical Microbiology |
| Irish Mycobacterial Reference Laboratory | Clinical Microbiology, St. James's Hospital, James's Street, Dublin 8 | Clinical Microbiology |
| Leeds Cancer Centre | HMDS, Leeds Cancer Centre, Bexley Wing, Beckett Street, Leeds LS9 7TF | Haematology |
| Leeds Endocrinology Laboratory | Department of Specialist Laboratory Medicine Block 46 St James Hospital Leeds Gen LS9 7TF | Biochemistry |
| LMH, King's Haematological Malignancies Diagnostic Centre (KHMDC), | Molecular Haemato-Oncology (LMH), Department of Haematological Medicine, King's College Hospital, The Rayne Institute, 123 Coldharbour Lane, London SE5 9NU (BCR-ABL1 Kinase Domain Mutations using renal Sequencing) | Haematology |
| Malaria Reference Laboratory | PHE Malaria Reference Laboratory, Faculty of Infectious & Tropical Diseases, London School of Hygiene & Tropical Medicine, Keppel Street, LONDON, WC1E 7HT | Haematology |
| Manchester Centre for Genomic Medicine | Genomic Diagnostics Laboratory, 6th floor, St Mary's hospital, Oxford Road, Manchester M13 9WL, UK. | Pathology |
| Med Lab Pathology | Unit 3, Sandyford Business Centre, Sandyford Business Park, Dublin 18 | Biochemistry |
| Metabolic Investigation Laboratory, Children's Health Ireland | Temple St., Dublin 1 | Biochemistry |
| Microbiology | Great Ormond Street Hospital for Children, Great Ormond Street, London WC1N 3JH, England | Clinical Microbiology |
| Micropathology Ltd | University of Warwick Science Park, Venture Centre, Sir William Lyons Road, Coventry CV4 7EZ | Clinical Microbiology |
| Microbiology, Central Pathology Laboratory | St James's Hospital, James's St., Dublin 8 | Clinical Microbiology |
| Mitochondrial NCG Diagnostic Service | The Medical School, Newcastle University, Framlington Place, Newcastle upon Tyne NE2 4HH, UK | Pathology |
| Molecular Histopathology Laboratory, Beaumont Hospital | Molecular Histopathology Laboratory, Department of Pathology, R.C.S.I. Education & Research Centre, Beaumont Hospital, Dublin 9 | Pathology |

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| Name | Address | Referring Dept |
|---|--|----------------------------|
| Molecular Microbiology, Central Pathology Laboratory | St James's Hospital, James's St., Dublin 8 | Clinical Microbiology |
| Molecular Pathology Diagnostic Service | Queen Elizabeth Hospital Birmingham Mindelsohn Way Edgbaston Birmingham, B15 2GW United Kingdom | Pathology |
| MRSA National Reference Laboratory | St. James's Hospital, James's Street, Dublin 8. | Clinical Microbiology |
| Munich Leukaemia Laboratory (MLL) | MLL Münchner Leukämielabor GmbH, Max-Lebsche-Platz 31, 81377 München, Postfach 20 14 53, 80014 Munich, Germany | Haematology |
| Mycology Reference Centre | Old Medical School, Thoresby Place, Leeds LS1 3EX, England | Clinical Microbiology |
| National Amyloidosis Centre | Royal Free Hospital Rowland Hill Street London, NW3 2PF | Biochemistry, Pathology |
| National Centre for Medical Genetics | National Centre for Medical Genetics Our Lady's Children's Hospital Crumlin Dublin 12, Ireland | Biochemistry |
| National Centre for Medical Genetics | National Centre for Medical Genetics Our Lady's Children's Hospital Crumlin Dublin 12, Ireland | Haematology |
| National Coagulation Laboratory | National Coagulation Laboratory, Centre for Clinical Pathology and Laboratory Medicine, (CPLM), St James Hospital, Dublin 8 | Haematology |
| National Carbapenemase Producing Enterobacteriales Reference Laboratory | Carbapenemase Producing Enterobacteriales (CPE) Reference Laboratory, Department of Medical Microbiology, University Hospital Galway, Galway | Clinical Microbiology |
| National Mycobacterium Reference Laboratory | Abernethy Building Institute of Cell and Molecular Science (ICMS)2 Newark Street London E1 2AT | Clinical Microbiology |
| National Parasitology Reference Laboratory (NPRL) | Department of Clinical Parasitology, Hospital for Tropical Diseases, Mortimer Market, Capper Street, London WC1E 6JB, England | Clinical Microbiology |
| National Salmonella, Shigella & Listeria Reference Laboratory | Department of Medical Microbiology, University Hospital Galway, Galway | Clinical Microbiology |
| National Virus Reference Laboratory (NVRL) | University College Dublin, Belfield, Dublin 4, Ireland | Clinical Microbiology |
| Neuroimmunology Dept | National Hospital for Neural and Neurosurgery, Queen Square, London WC1N 3BG | Biochemistry |
| NHSBT Centre Bristol | NHSBT Centre, 500 North Bristol Park, Northway, Filton, Bristol, BS34 7QH, UK | Haematology |
| Nutristasis Unit | Haemostasis and Thrombosis GSTS Pathology4th floor, North Wing St Thomas' Hospital Westminster Bridge Road London SE1 7EH | Biochemistry |

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| Name | Address | Referring Dept |
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| OLCH, National Centre for Medical Genetics (NCMG) Crumlin | Division of Cytogenetics (Oncology), National Centre for Medical Genetics (NCMG), Our Lady's Hospital, Department of Clinical Genetics, Children's Health Ireland at Crumlin Dublin D12 N512 | Biochemistry |
| Oxford University Hospitals NHS Foundation Trust | National Haemoglobin Reference Laboratory, Oxford Haemophilia Centre, Churchill Hospital, Oxford OX3 7LJ | Haematology |
| Oxford Genetics Laboratories | Oxford Regional Genetics Laboratories, Churchill Hospital, Headington, Oxford, OX3 7LE, United Kingdom | Haematology |
| Oncology Cytogenetics | Cytogenetics Oncology, 5 th Floor Tower Wing, Guy's Hospital, Great Maze Pond, London, SE1 9RT UK Tel: 020 7188 1709 | Haematology |
| Oncology Cytogenetics | Dr. Jonathan Shanks, Oncology Cytogenetics, The Christie Foundation, Manchester, United Kingdom | Pathology |
| Poundbury Cancer Institute | Dr Corrado D'Arrigo, Poundbury Cancer Institute, Dorset, United Kingdom | Pathology |
| Primary Ciliary Dyskinesia (PCD) Diagnostic Service, University Hospital Southampton | Patricia Goggin/Regan Doherty PCD EM Scientists Biomedical imaging Unit Mail point 12South Academic Block Southampton General Hospital UK SO166YD | Pathology |
| Public Health Laboratory, Cherry Orchard Hospital | PHL Cherry Orchard Hospital, Ballyfermot, Dublin 10 | Clinical Microbiology |
| Purine Research Laboratory | Dr Lynette Fairbanks, 4th Floor, North Wing, St. Thomas's Hospital, London SE1 7EH | Biochemistry |
| Rare and Imported Pathogens Laboratory (RIPL) | UK Health Security Agency, Porton Down, Salisbury, Wiltshire SP4 0JG, England | Clinical Microbiology |
| Respiratory and Vaccine Preventable Bacteria Reference Unit (RVPBRU) | | Clinical Microbiology, Biochemistry |
| Rotunda Hospital | Rotunda Hospital , Parnell Street, Dublin 1, DO1 P5W9 | Haematology |
| Royal Free Hospital HSL | Haematology Laboratory, Royal Free Hospital HSL Analytics LLP, Katharine Dormandy Haemophilia Centre and Thrombosis Unit First Floor, Royal Free Hospital, Pond Street, London NW3 2QG, U.K. | Haematology |
| Royal Marsden Hospital NHS Foundation TR | RMH HMDS, The Centre for Molecular Pathology, The Royal Marsden NHS Foundation Trust, Cotswold Road, Sutton, Surrey, SM2 5NG | Haematology |
| Salamanca University | | Haematology |
| SAS Centre | c/o Ground Floor Oncology Charing Cross Hospital Fulham Palace RoadLONDONW6 8RF | Biochemistry |
| SAS Peptide Hormones, Royal Surrey County Hospital | Clinical Laboratory, Royal Surrey County Hospital, Egerton Road,GUILDFORDGU2 5XX | Biochemistry |

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| Name | Address | Referring Dept |
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| SAS Trace Element Unit | Division of Laboratory Medicine Southampton University Hospitals NHS Trust Mail Point 804Southampton General Hospital Tremona RoadSOUTHAMPTONSO16 6YD | Biochemistry |
| Sexually Transmitted Bacteria Reference Laboratory (STBRL) | Bacteriology Reference Department, UK Health Security Agency, 61 Colindale Avenue, London NW9 5HT, England | Clinical Microbiology |
| Sheffield Diagnostic Genetics Service | Sheffield Children's NHS Foundation Trust Western Bank, Sheffield S10 2TH Sheffield Diagnostic Genetics Service, C Floor Blue Lifts, Sheffield Children's NHS Foundation Trust, Clarkson Street, Sheffield, S10 2TQ | Haematology |
| TDL Genetic Referrals | The Doctor's Laboratory Genetics,60 Whitfield Street, London W1T 4EU | Biochemistry |
| The Antimicrobial Resistance and Healthcare Associated Infections Reference Unit (AMRHAI) | Bacteriology Reference Department, UK Health Security Agency, 61 Colindale Avenue, London NW9 5HT, England | Clinical Microbiology |
| The National Creutzfeldt- Jakob Disease Research & Surveillance Unit | Room FU 529, First Floor, Chancellor's Building, 49 Little France Crescent, Edinburgh EH16 4SB, UK | Pathology |
| Thyroseq International | University of Pittsburg Medical Centre, 200 Meyran Ave # 318, Pittsburgh, PA 15213, United States | Pathology (Cytology) |
| Toxicology Laboratory, Beaumont Hospital | Beaumont, Dublin 9 | Biochemistry |
| Toxoplasma Reference Laboratory (TRL) | Singleton Hospital, Swansea SA2 8QA, Wales | Clinical Microbiology |
| Trace Element Laboratory | Centre of Clinical Science & Measurement, School of Biological Sciences, University of Surrey, Guildford GU2 5XHEndocrine Laboratory | Biochemistry |
| Trace Element Unit, King's Healthcare Trust | Dr Raja, Trace Element Unit, Dept. of Clinical Biochemistry King's Healthcare Trust Denmark Hill London, SE5 9RSEngland | Pathology |
| UKHSA Mycology Reference Laboratory | UKHSA South West Laboratory, Science Quarter, Southmead Hospital, Bristol BS10 5NB, England | Clinical Microbiology |
| Viapath, GSTS Pathology | Viapath, GSTS Pathology Centre, The Human Nutristasis Unit, Haemostasis Laboratories, 4th Floor North Wing, St Thomas' Hospital, Westminster Bridge Road, London SE1 7EH, United Kingdom | Haematology |
| Virology Reference Department | UK Health Security Agency, 61 Colindale Avenue, London NW9 5HT, England | Clinical Microbiology |
| Wessex Regional Genetics Laboratory | Leukaemia Research Group, Wessex Regional Genetics Laboratory, Salisbury District Hospital, Salisbury, Wiltshire, SP2 8BJ | |
| Wellchild Laboratory | Wellchild Research Laboratory, 12th floor, Guy's Hospital, London SE1 9RT | Biochemistry |
| Willink Biochemical Genetics Unit | Genetic Medicine, 6th Floor, St Mary's Hospital, Oxford Road, Manchester M13 9WL | Biochemistry |
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